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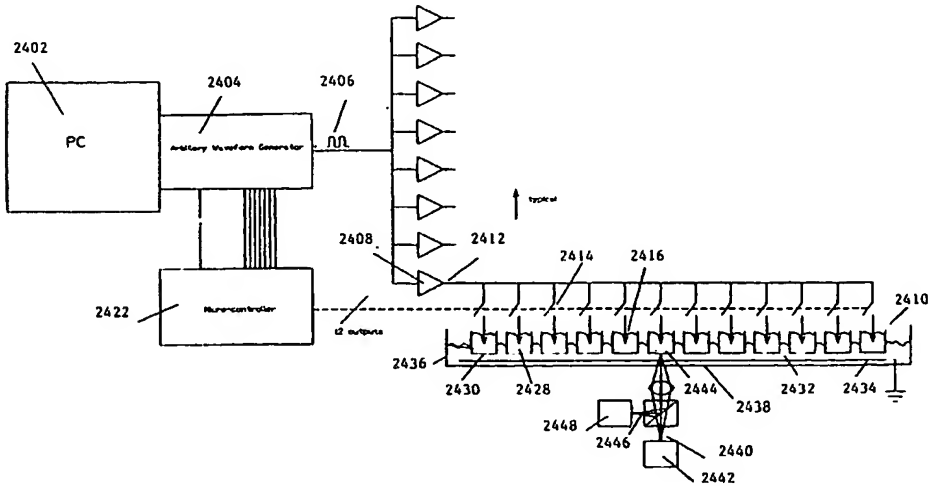
(54) Title: ELECTRICAL FIELD STIMULATION OF EUKARYOTIC CELLS



(57) Abstract: Methods of identifying activators and inhibitors of voltage-gated ion channels are provided in which the methods employ electrical field stimulation of the cells in order to manipulate the open/close state transition of the voltage-gated ion channels. This allows for more convenient, more precise experimental manipulation of these transitions, and, coupled with efficient methods of detecting the result of ion flux through the channels, provides methods that are especially suitable for high throughput screening.

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For two-letter codes and other abbreviations, refer to the "Guidance Notes on Codes and Abbreviations" appearing at the beginning of each regular issue of the PCT Gazette.

TITLE OF THE INVENTION

ELECTRICAL FIELD STIMULATION OF EUKARYOTIC CELLS

CROSS-REFERENCE TO RELATED APPLICATIONS

The subject application is related to co-pending provisional application no. 60/304,955, filed July 12, 2001, to which priority is claimed under 35 USC § 119(e).

STATEMENT REGARDING FEDERALLY-SPONSORED R&D

Not applicable.

REFERENCE TO MICROFICHE APPENDIX

Not applicable.

FIELD OF THE INVENTION

The present invention is directed to methods and associated apparatuses for stimulating eukaryotic cells by the application of electric fields. The electric fields are produced by certain arrangements of electrodes that create an electric potential difference in the environment of the cells, resulting in a change in membrane potential of the cells. The change in membrane potential affects various physiological processes within the cells, including the opening and closing of voltage-gated ion channels. The ability to alter the open/close transitions of voltage-gated ion channels by the application of electric fields as described herein provides for novel methods of screening compounds for the ability to modulate the activity of voltage-gated ion channels.

BACKGROUND OF THE INVENTION

Certain molecular events in eukaryotic cells depend on the existence or magnitude of an electric potential gradient across the plasma (*i.e.*, outer) membrane of the cells. Among the more important of such events is the movement of ions across the plasma membrane through voltage-gated ion channels. Voltage-gated ion channels form transmembrane pores that open in response to changes in cell membrane potential and allow ions to pass through the membrane. Voltage-gated ion channels have many physiological roles. They have been shown to be involved in

maintaining cell membrane potentials and controlling the repolarization of action potentials in many types of cells (Bennett et al., 1993, Cardiovascular Drugs & Therapy 7:195-202; Johnson et al., 1999, J. Gen. Physiol. 113:565-580; Bennett & Shin, "Biophysics of voltage-gated sodium channels," in Cardiac Electrophysiology: From Cell to Bedside, 3rd edition, D. Zipes & J. Jalife, eds., 2000, W.B. Saunders Co., pp.67-86; Bennett & Johnson, "Molecular physiology of cardiac ion channels," Chapter 2 in Basic Cardiac Electrophysiology and Pharmacology, 1st edition, A. Zasa & M. Rosen, eds., 2000, Harwood Academic Press, pp. 29-57). Moreover, mutations in sodium, calcium, or potassium voltage-gated ion channel genes leading to defective channel proteins have been implicated in a variety of disorders including the congenital long QT syndromes, ataxia, migraine, muscle paralysis, deafness, seizures, and cardiac conduction diseases, to name a few (Bennett et al., 1995, Nature 376:683-685; Roden et al., 1995, J. Cardiovasc. Electrophysiol. 6:1023-1031; Kors et al., 1999, Curr. Opin. Neurol. 12:249-254; Lehmann et al., 1999, Physiol. Rev. 79:1317-1372; Holbauer & Heufelder, 1997, Eur. J. Endocrinol. 136:588-589; Naccarelli & Antzelevitch, 2000, Am. J. Med. 110:573-581).

Several types of voltage-gated ion channels exist. Voltage-gated potassium channels establish the resting membrane potential and modulate the frequency and duration of action potentials in neurons, muscle cells, and secretory cells. Following depolarization of the membrane potential, voltage-gated potassium channels open, allowing potassium efflux and thus membrane repolarization. This behavior has made voltage-gated potassium channels important targets for drug discovery in connection with a variety of diseases. Dysfunctional voltage-gated potassium channels have been implicated in a number of diseases and disorders. Wang et al., 1998, Science 282:1890-1893 have shown that the voltage-gated potassium channels KCNQ2 and KCNQ3 form a heteromeric potassium ion channel known as the "M-channel." Mutations in KCNQ2 and KCNQ3 in the M-channel are responsible for causing epilepsy (Biervert et al., 1998, Science 279:403-406; Singh et al., 1998, Nature Genet. 18:25-29; Schroeder et al., Nature 1998, 396:687-690).

Voltage-gated sodium channels are transmembrane proteins that are essential for the generation of action potentials in excitable cells (Catterall, 1993, Trends Neurosci. 16:500-506). In mammals, voltage-gated sodium channels consist of a macromolecular assembly of α and β subunits with the α subunit being the pore-forming component. α subunits are encoded by a large family of related genes, with

some α subunits being present in the central nervous system (Noda et al., 1986, Nature 322:826-828; Auld et al., 1988, Neuron 1:449-461; Kayano et al., 1988, FEBS Lett. 228:187-194) and others in muscle (Rogart et al., 1989, Proc. Natl. Acad. Sci. USA 86:8170-8174; Trimmer et al., 1989, Neuron 3:33-49).

Voltage-gated calcium channels are transmembrane proteins that in the open configuration allow the passive flux of Ca^{2+} ions across the plasma membrane, down the electrochemical gradient. They mediate various cell functions, including excitation-contraction coupling, signal transduction, and neurotransmitter release.

Current methods of drug discovery often involve assessing the biological activity (*i.e.*, screening) of tens or hundreds of thousands of compounds in order to identify a small number of those compounds having a desired activity. In many high throughput screening programs, it is desirable to test as many as 50,000 to 100,000 compounds per day. Unfortunately, current methods of assaying the activity of voltage-gated ion channels are ill suited to the needs of a high throughput screening program. Current methods often rely on electrophysiological techniques. Standard electrophysiological techniques involve "patching" or sealing against the cell membrane with a glass pipette followed by suction on the glass pipette, leading to rupture of the membrane patch (Hamill et al., 1981, Pflugers Arch. 391:85-100). This has limitations and disadvantages. Accessing the cell interior may alter the cell's response properties. The high precision optical apparatuses necessary for micromanipulating the cells and the pipettes make simultaneous recording from more than a few cells at a time impossible. Given these difficulties, the throughput that can be achieved with electrophysiological techniques falls far short of that necessary for high throughput screening.

Various techniques have been developed as alternatives to standard methods of electrophysiology. For example, radioactive flux assays have been used in which cells are loaded with a radioactive tracer (*e.g.*, $^{86}Rb^+$, $^{22}Na^+$, [^{14}C]-guanidinium) and the efflux of the dye is monitored. Cells loaded with the tracer are exposed to compounds and those compounds that either enhance or diminish the efflux of the tracer are identified as possible activators or inhibitors of ion channels in the cells' membranes.

Assays that measure the change in a cell's membrane potential due to the change in activity of an ion channel have been developed. Such assays often employ voltage sensitive dyes that redistribute between the extracellular environment

and the cell's interior based upon a change in membrane potential and that have a different fluorescence spectrum depending on whether they are inside or outside the cell. A related assay method uses a pair of fluorescent dyes capable of fluorescence resonance energy transfer to sense changes in membrane potential. For a description of this technique, see González & Tsien, 1997, Chemistry & Biology 4:269-277. See also González & Tsien, 1995, Biophys. J. 69:1272-1280 and U.S. Patent No. 5,661,035. Other methods employ ion selective indicators such as calcium dependent fluorescent dyes to monitor changes in Ca^{2+} influx during opening and closing of calcium channels.

Ideally, methods of screening against voltage-gated ion channels require that the transmembrane potential of the cells being assayed be controlled and/or that the ion channels studied be cycled between open and closed states. This has been done in various ways. In standard electrophysiological techniques, the experimental set-up allows for direct manipulation of membrane potential by the voltage clamp method (Hodgkin & Huxley, 1952, J. Physiol. (Lond.) 153:449-544), *e.g.*, changing the applied voltage or injecting various ions into the cell. In other methods, changing the extracellular K^+ concentration from a low value (*e.g.*, 5 mM) to a higher value (*e.g.*, 70-80 mM) results in a change in the electrochemical potential for K^+ due to the change in the relative proportion of intracellular and extracellular potassium. This results in a change in the transmembrane electrical potential towards a more depolarized state. This depolarization can activate many voltage-gated ion channels, *e.g.*, voltage-gated calcium, sodium, or potassium channels. Alternatively, Na^+ channels can be induced into an open conformation by the use of toxins such as veratridine or scorpion venom (Strichartz et al., 1987, Ann. Rev. Neurosci. 10:237-267; Narahashi & Harman, 1992, Meth. Enzymol. 207:620-643). While sometimes effective, such experimental manipulations may alter the channel pharmacology, can be awkward to perform, and can lead to artifactual disturbances in the system being studied.

Electrical field stimulation of cells has been performed on a single cell by sealing a glass microelectrode to the cell membrane. Rupture of the sealed patch of cell membrane resulted in an electrical connection between the interior fluid in the glass microelectrode and the fluid within the cell that was used to stimulate the cell via an electronic pulse generator. The electrophysiological response of the cell was measured via a sensitive electronic amplifier. The disadvantage of this technique is

that only one cell at a time was tested and it is a tedious and time consuming operation to seal the microelectrode to an individual cell.

HEK293 cells have been grown on a silicon chip made up of an array of field-effect transistors. Some of the cells were positioned over the gate region of the transistors, thus having portions of their plasma membranes overlying the source and the drain. When a patch pipette in such cells manipulated the intracellular voltage, Maxi-K potassium channels in the cells' plasma membranes were opened. This led to current flow in the region between the cells' membrane and the transistor. This current flow modulated the source-drain current, which could be detected by an appropriate device. The chip plus cells was said to have potential as a sensor and as a prototype for neuroprosthetic devices. See Straub et al., 2001, Nature Biotechnol. 19:121-124; Neher, 2001, Nature Biotechnol. 19:114.

SUMMARY OF THE INVENTION

The present invention is directed to methods of identifying activators and inhibitors of voltage-gated ion channels in which the methods employ electrical field stimulation of the cells via extracellular electrodes in order to manipulate the open/close state transitions of the voltage-gated ion channels. This allows for more convenient, more precise manipulation of these transitions, and, coupled with efficient methods of detecting ion flux or membrane potential, results in methods that are especially suitable for high throughput screening in order to identify substances that are activators or inhibitors of voltage-gated ion channels.

The present invention also provides apparatuses for use in the above-described methods. In particular, modifications of standard multiwell tissue culture plates are provided where the modified multiwell tissue culture plates have electrodes that can alter the transmembrane electric potential of cells in the wells of the plates, thus altering the ratio of open/close states of voltage-gated ion channels in the cells.

BRIEF DESCRIPTION OF THE DRAWINGS

Figure 1A shows a top view of one embodiment of the present invention. This embodiment comprises a glass slide 1 in which or upon which are a gold positive electrode 2 and a gold negative electrode 3 spaced such that a gap 4 of about 25 μm to 100 μm exists between the electrodes. The electrodes together with spacers 5 (here shown as plastic strips) arranged generally at right angles to the

electrodes define a series of wells 6 about 100 μm deep into which cells can be placed and/or grown. Figure 1B shows a cross-sectional side view of the embodiment of Figure 1A. In this embodiment, the identities of the positive and negative electrodes can be interchanged, if desired. The electrodes need not be made from gold; other conductive materials may be used. Also, the spacers need not be plastic; other non-conductive materials may be used.

Figure 2A shows a top view of an embodiment of the present invention in which a typical 96 well plate contains electrodes within each well. Figure 2B shows a cross-sectional side view of one of the wells in Figure 2A. The well has a first electrode 1 (here shown as a positive electrode) on the side 2 of the well, a second electrode 3 (here shown as a negative electrode) on the bottom 4 of the well, a strip of an optional insulating material 5 on the bottom of the well, and a cell 6 at the bottom of the well. A single cell is shown merely for convenience of illustration; in most cases a plurality of cells would be in the bottom of the well. The sides 2 of the well are made of a non-conducting material such as plastic and the bottom of the well is made from a conducting material such as indium tin oxide (ITO). The well is shown with a fluid level 7 sufficient to completely cover the cell 6 and the second electrode 3 at the bottom 4 of the well and to reach the first electrode 1 on the side 2 of the well. The well is not drawn to scale with respect to Figure 2A. Figure 2C shows an alternative arrangement of electrodes in a well. In this embodiment, both the positive electrode 1 and the negative electrode 2 are in the bottom 3 of the well. In this embodiment, the sides 4 and bottom 3 of the well are made of non-conducting material such as plastic. The fluid level 5 is such as to cover the cells 6 as well as the positive 1 and negative 2 electrodes.

Figure 3 shows a single well 1 from an embodiment of the invention where first 2 and second 3 electrodes are interdigitating and have been chemically etched on a layer of conductive material on the surface of a glass substrate 4. The well is generally circular with a 3 mm diameter. The electrodes are 10 μm wide and have a spacing of 160 μm. Either the first 2 or the second 3 electrodes may function as the positive electrode. The width of the electrodes and the spacing between the electrodes can be varied. The width is preferably between 1 and 10 μm; the spacing between the electrodes is preferably 5 μm to 160 μm. In particularly preferred embodiments, the spacing between the electrodes is at least as great as a typical diameter of a eukaryotic cell (*i.e.*, about 40 μm to 50 μm).

Figure 4A and 4B illustrates an embodiment in which wells are formed by attaching a well frame onto the substrate. Figure 4A shows an exploded view of the embodiment containing a well frame 1 the openings 2 of which form the wells on the substrate 3 where the well frame 1 is attached to the substrate 3 (*e.g.*, by gluing it in place), a contact guide plate 5 with a spring loaded contact 6, and a printed circuit board (PCB) 7. The substrate holder 4 is used to hold the assembled device in position on a measuring instrument such as a microscope or fluorescent plate reader (not shown). The PCB 7 contains connections through which the electrodes (not shown) can be linked to a pulse generator (not shown). Figure 4B shows an assembled view.

Figure 5 shows an arrangement of interdigitating electrodes formed upon a substrate that contains virtual wells. Virtual wells are described further herein.

Figure 6 shows a single well from an embodiment of the invention where two substantially parallel plates 1 have their opposing surfaces coated with conductive layers 2 between which is sandwiched a droplet of fluid containing the cells to be tested 3. One conductive layer is a positive electrode (here the upper conductive layer 4) while the other conductive layer is a negative electrode (here the lower conductive layer 5). Of course, the identity of the electrodes could be reversed, with the upper conductive layer being the negative electrode and the lower conductive layer being the positive electrode). In particular versions of this embodiment, the plates are glass and the conductive layer is indium tin oxide (ITO). The conductive layer preferably has a thickness of about 200 Å to 2,000 Å, or 500 Å to 1,500 Å, or 800 Å to 1,200 Å.

Figure 7 shows a single well 3 from an embodiment of the invention where one of the electrodes is a thin coating of conductive material 2 on the surface of a flat substrate 1 and forms the bottom 10 of the well. The other electrode 7 enters the well 3 from above and makes contact with the fluid 5 within the well 3. Electrode 7 is shown in cut-away view. Electrode 7 contains a central conductive material portion 8 that is surrounded by an insulator 6. For the sake of simplicity, a single cell 4 is shown in the well. Generally, at least 10^5 cells would be present in the well. The conductive layer preferably has a thickness of about 200 Å to 2,000 Å, or 500 Å to 1,500 Å, or 800 Å to 1,200 Å.

Figure 8 shows a single well 4 from an embodiment of the invention where the bottom of the well 4 is a filter membrane 12 upon which cells can be

grown. For simplicity, a single cell 8 is shown on the filter membrane 12. The well 4 is located in a trough 2 having a glass bottom 1 and filled with a first fluid 3. One electrode 7 enters the well 4 from above and makes contact with a second fluid 5 within the well 4. Electrode 7 contains a central conductive material portion that is surrounded by an insulator 6 and is connected to a pulse generator (not shown) by a first lead 9. A second electrode 11 is positioned within the first fluid 3 and is connected to the pulse generator by a second lead 10. The second electrode 11 is shown in cut-away view. The second electrode 11 actually forms a circle in the bottom of the well 4. Either the first electrode 7 is the positive electrode while the second electrode 11 is the negative electrode or the first electrode 7 is the negative electrode while the second electrode 11 is the positive electrode.

Figure 9A shows a single well 2 from an embodiment of the invention where both the positive 5 and negative 8 electrodes enter the well 2 from above. The well 2 contains fluid 3 in which a single cell 9 is shown, although generally a plurality of cells will be present in the well 2. The positive electrode 5 is connected to a pulse generator (not shown) by a positive lead 6. The negative electrode 8 is connected to the pulse generator by a negative lead 7. Both electrodes are embedded in an insulator 4. The positive 5 and negative 8 electrodes traverse the interior of the insulator 4 such that the positive 5 and negative 8 electrodes are generally perpendicular to a glass plate 1 that forms the bottom of the well 2. However, when the positive 5 and negative 8 electrodes exit the bottom 10 of the insulator 4, the positive 5 and negative 8 electrodes are each bent into a 90° angle so that they lie on and parallel to the bottom 10 of the insulator 4. Figure 9B is a view looking up from the glass plate 1 that forms the bottom of the well 2 and shows the arrangement of the bent portion of the positive 5 and negative 8 electrodes lying on bottom of the insulator 4.

Figure 10A shows an embodiment of the invention where both the positive 5 and negative 8 electrodes enter the well 2 from above and the positive 5 and negative 8 electrodes are arranged in a manner similar to that of a co-axial cable. The positive electrode 5 is embedded in an insulator 4 with the negative electrode 8 coating the outside of the insulator 4. The positive electrode 5 is connected to a pulse generator (not shown) by a positive lead 6. The negative electrode 8 is connected to the pulse generator by a negative lead 7. The well 2 contains fluid 3 in which a single cell 9 is shown, although generally a plurality of cells will be present in the well 2. A

glass plate 1 forms the bottom of the well 2. Figure 10B shows a view looking up from below the positive 5 and negative 8 electrodes.

Figure 11 shows an embodiment of the invention similar to the embodiment shown in Figure 8 except that in Figure 11 the electrode 7 that enters the well from above is not surrounded by an insulator but instead is within a pipette tip 6 and makes contact with a first fluid 5 also within the pipette tip 6 that is co-extensive with the first fluid 5 in the well 4. This arrangement has the advantage of minimizing the formation of bubbles in the first fluid 5 in the area at the end of the electrode 7. The bottom of the well 4 is a filter membrane 12 upon which cells can be grown. For simplicity, a single cell 8 is shown on the filter membrane 12. The well 4 sits in a trough 2 having a glass bottom 1 and filled with a second fluid 3. Electrode 7 is connected to a pulse generator (not shown) by a first lead 9. A second electrode 11 is positioned within the second fluid 3 and is connected to the pulse generator by a second lead 10. The second electrode 11 is shown in cut-away view. The second electrode 11 actually forms a circle in the bottom of the well 4. Either electrode can be the positive or negative electrode.

Figure 12A-B shows an embodiment that is similar to the embodiment of Figure 7 in having one electrode enter from above while the other electrode forms the bottom of the wells. Figure 12A is a side cross-sectional view that shows a substrate that is a 96-well microtiter plate in which one electrode 1 is a layer of a conductive material such as ITO that forms the bottom of the wells 2. The other electrode 3 enters the wells from above and makes contact with the fluid in the wells (fluid not shown). The electrodes are connected to an electrical pulse generator 4 by leads 5. Either electrode may be the positive or negative electrode. An alternative embodiment, similar to that shown, is to replace the bottom of standard 96, 384, 1536, or 3456 well plates with a conductive material such as ITO, which forms one electrode. The second electrode is lowered into each well from above. Contact to the ITO electrode can be made via electrically conducting silver epoxide or by placing a 3 M KCl (or similar salt solution) in alternate wells as the contact to the ITO bottoms from a platinum wire. Figure 12B shows a top view of the substrate.

Figure 13A-B shows an embodiment comprising two multiwell substrates containing virtual wells. Figure 13A is a side cross-sectional view that shows the top substrate 1 approaching the bottom substrate 2. The top electrode 3 is made of a conducting material such as ITO and forms the bottom of the virtual wells 4

of the top substrate 1. Similarly, the bottom electrode 5 is made of a conducting material such as ITO and forms the bottom of the virtual wells 6 of the bottom substrate 2. A thin layer of TEFLON® or a similar hydrophobic material 11 covers the surfaces of the conducting material on the substrates. Circular areas of the surface of the substrate that lack TEFLON® are relatively hydrophilic and form the virtual wells. The TEFLON® layer is about 0.5 μm to 100 μm thick. The top 3 and bottom 5 electrodes are connected to an electrical pulse generator 6 by leads 7. The left most wells of the apparatus are shown containing fluid drops. The top drop 8 might contain a substance such as a drug or a compound to be tested while the bottom drop 9 might contain cells expressing a voltage-gated ion channel. Figure 13B shows the apparatus after the top 1 and bottom 2 substrates have moved close enough together so that the top 8 and bottom 9 drops have mixed. 10 is a spacer (not shown in Figure 13A) that helps to align the top 1 and bottom 2 substrates and keeps the substrates the proper distance apart for mixing of the drops.

Figure 14 illustrates the principles of electrical field stimulation of cells.

Figure 15 shows two wells from an embodiment where one electrode enters the wells from above 1 while the second electrode is formed from the transparent ITO-coated bottom 2 of the transparent substrate 3 that is in contact with a highly conductive metal grounding grid 4. The dashed lines with arrowheads illustrate how current flows from the electrodes that enter from above 1 through a buffered salt solution 5 and the cells 6 and through the ITO layer 2 and the metal grounding grid 4. Arrows 7 within the substrate 3 illustrate how light from a source used in the detection system (not shown) would pass in the upward direction through the transparent substrate 3 and the ITO layer 2 into the cells 6 and then be re-emitted by the cells 6 as fluorescence and pass downward to a detector (not shown). Optional adhesive seals 8 that can be used to attach the wells to the ITO-coated substrate 3 are shown. The thickness of the ITO layer is preferably about 200 Å to 2,000 Å, or 500 Å to 1,500 Å, or 800 Å to 1,200 Å.

Figure 16A shows two wells of a multiwell embodiment having a conductive layer 1 such as ITO that forms the bottom of the wells. The positive electrode 2 enters the left well 3 from above while the negative electrode 4 enters the right well 5 from above. The transparent layer of a conductive material 1 such as ITO coats a transparent substrate 7 such as glass. The dotted line with an arrowhead

shows the path of current flow. Of course, the identity of the positive and negative electrodes could be reversed. Cells 8 are shown in fluid 9 within the wells. Optional adhesive seals 10 that can be used to attach the wells to the ITO-coated substrate 7 are shown. Light path is indicated by arrows in the substrate. Figure 16B shows a side cut-away view of this embodiment that illustrates how the positive 2 and negative 4 electrodes might be connected to a pulse generator 11. Also shown is the transparent conductive layer 6 coating the transparent substrate 7. Figure 16C shows a top view of the embodiment that illustrates the alternating pattern of positive and negative electrodes. Figure 16D is a photograph of this embodiment that has been partially disassembled. The wells are formed by a well frame 12 that is attached to the glass substrate 13 that is has been coated with ITO. During normal operation, the substrate will cover all the wells. For the purpose of illustration, this view shows only part of the substrate.

Figure 17 shows a graphical representation of data obtained from an embodiment of the invention similar to that depicted in Figure 16. The data represent Ca^{2+} influx into HEK293 cells that have been transfected to express the human α1H T-type voltage-gated calcium channel (GenBank accession no. AF073931). Ca^{2+} influx occurred when the T-type channels opened and was measured by detecting fluorescent emission at 520-560 nm of the calcium indicator dye Fluo4 that had been excited at 480 nm. At the time points indicated, a preselected voltage was applied through the electrodes. This resulted in the opening of a portion of the T-type channels, allowing Ca^{2+} influx. This caused a spike in the fluorescent emission at 520-560 nm by the calcium indicator dye Fluo4. The spike gradually decayed, as shown.

Figure 18A-B shows a nucleotide sequence encoding the human PN3 sodium channel (SEQ.ID.NO.:1). Figure 18C shows the corresponding amino acid sequence (SEQ.ID.NO.:2). From GenBank accession no. AF117907.

Figure 19A-B shows a nucleotide sequence encoding the α1H subunit of the human T-type calcium channel (SEQ.ID.NO.:3). Figure 19C shows the corresponding amino acid sequence (SEQ.ID.NO.:4). From GenBank accession no. AF073931.

Figure 20A-B shows a nucleotide sequence encoding a splice variant of the α1B subunit of the human N-type calcium channel (SEQ.ID.NO.:5). Figure

20C shows the corresponding amino acid sequence (SEQ.ID.NO.:6). From GenBank accession no. M94172.

Figure 21A-B shows a nucleotide sequence encoding a splice variant of the α1B subunit of the human N-type calcium channel (SEQ.ID.NO.:7). Figure 21C shows the corresponding amino acid sequence (SEQ.ID.NO.:8). From GenBank accession no. M94173.

Figure 22A-B shows a nucleotide sequence encoding the human calcium channel α1A isoform 1A-1 subunit (SEQ.ID.NO.:9). Figure 22C shows the corresponding amino acid sequence (SEQ.ID.NO.:10). From GenBank accession no. AF004884.

Figure 23A-B shows a nucleotide sequence encoding the human calcium channel α1A isoform 1A-2 subunit (SEQ.ID.NO.:11). Figure 23C shows the corresponding amino acid sequence (SEQ.ID.NO.:12). From GenBank accession no. AF004883.

Figure 24 shows a schematic diagram of one embodiment of a EFS system utilizing a computer, voltage generator, amplifier, membrane bottom wells, common trough, and fluorescence detector, *inter alia*.

Figure 25 is a photograph showing an electrode head embodiment especially adapted for use with a 96 well tray.

Figure 26 is a photograph showing a trough embodiment for use in conjunction with the electrode head embodiment shown in Figure 25.

Figure 27 is a photograph showing the trough embodiment of Figure 26 with a multi-screen well tray positioned therein.

Figure 28 is a photograph showing the assembled electrode head, trough and multiscreen.

Figure 29 shows a graphical representation of data obtained from an embodiment of the invention similar to that depicted in Figure 28. The data represent a membrane potential change in HEK293 cells that have been transfected to express

human PN1 voltage-gated sodium channel. Each plot represents a row (12wells) A-H of a 96-well plate. Each column of the 96-well plate data was acquired for 15 seconds on a VIPR™. Stimulation pulse protocol was applied during the data acquisition as follows; 2s baseline was followed with a 2ms square pulse, Amplitude = 20mA, Frequency = 10 Hz, Duration = 5s.

Figure 30 is a bar graph representation of the peak ration change of data depicted in Figure 29. 1 μM TTX a specific and potent blocker of tetrodotoxin (TTX)-sensitive voltage-gated sodium channels is present in wells E1, F1, G1, H1, A12, B12, C12 and D12. In addition well A11 contains an internal standard for blocking TTX-sensitive voltage-gated sodium channels. Z-score is a measure of the difference in the uninhibited and inhibited signal divided by the sum of the standard deviations.

Figure 31 shows the effects of increasing concentrations of TTX (upper panel) and of Compound A (lower panel) on the EFS-stimulated depolarization signal in HEK293/PN1 cells. The IC_{50}s obtained in these experiments are comparable to those obtained through other techniques. The high Hill coefficients, nH, result from the threshold nature of the stimulation protocol.

Figure 32 is a photograph showing an alternative embodiment. Figure 32 shows an electrode head similar to that shown in Figure 25, and a copper electrode plate. This embodiment is especially adapted for use with Caco-2 multiscreens (Millipore, Beford, MA).

Figure 33 is a photograph similar to that shown in Figure 32 except that the copper electrode plate has been turned over to show conducting pins (note: pins extend out of page toward reader).

Figure 34 is a photograph showing the copper electrode plate placed on top of an assembled Caco-2 membrane bottom well and receiver tray.

Figure 35 is a photograph showing the assembled embodiment of Figure 34, i.e., electrode head, copper electrode plate with pins, Caco-2 membrane bottom well, and Caco-2 receiver tray.

Figure 36 depicts a novel electrode embodiment that comprises a dielectric disc sandwiched between two conductive discs. Figure 36A shows an expanded view of the novel electrode embodiment. Figure 36B shows the novel electrode embodiment electrically connected to a concentric lead. Figure 36C shows the novel electrode embodiment electrically connected to edge leads.

DETAILED DESCRIPTION OF THE INVENTION

The present invention provides equipment and techniques to implement electric field stimulation (EFS) of cells while monitoring a biological response of the cells. Preferably, the biological response is monitored by fluorescence detection. The cells are grown and/or attached to specially designed substrates such as, *e.g.*, glass slides which contain preferably transparent, electrically conductive electrodes or multiwell tissue culture plates containing electrodes so disposed that when a preselected voltage is applied across the electrodes the transmembrane potential of cells within the wells of the multiwell tissue culture plates is altered.

In general terms, the present invention involves providing a substrate upon which living eukaryotic cells, preferably mammalian cells, are present where the cells express voltage-gated ion channels in their plasma membranes. Positive and negative electrodes are positioned either on or near the substrate so that when a voltage is applied through the electrodes the voltage-gated ion channels either open or close, thereby modulating the flow of at least one type of ion through the plasma membranes of the cells. This modulation of ion flow, or a change in membrane potential that results from the modulation of ion flow, is detected, either directly or indirectly, preferably by the use of fluorescent indicator compounds in the cells.

Collections of substances, *e.g.*, combinatorial libraries of small organic molecules, natural products, phage display peptide libraries, etc., are brought into contact with the voltage-gated ion channels in the plasma membranes of the cells and those substances that are able to affect the modulation of ion flow are identified. In this way, the present invention provides methods of screening for activators and inhibitors of voltage-gated ion channels. Such activators and inhibitors are expected to be useful as pharmaceuticals or as lead compounds from which pharmaceuticals can be developed by the usual processes of drug development, *e.g.*, medicinal chemistry.

During an applied extracellular electrical field, the cell membrane electrical capacitance will charge or discharge depending upon the polarity and orientation of the cell relative to the field. This results in a transient change in the transmembrane potential in a given patch of membrane. These transient changes in transmembrane potential will vary continuously around each cell depending upon the orientation of each patch of membrane relative to the applied field and the existing transmembrane potential. In each membrane patch, membrane potential will be perturbed away from the resting value by the applied external field. This change in membrane potential will in turn affect the proportion of open and closed voltage-gated ion channels in each local patch of membrane, which will affect the conductance of the voltage-gated ion channels and thus change the membrane potential further. This process is expected to vary around each cell such that, in any given cell, different patches of membrane and the embedded voltage-gated ion channels will experience different membrane potentials. In general, the membrane potential in a given patch of membrane will change at a rate that is proportional to its resistance (1/conductance) and its capacitance (C_m) such that $dV/dt = I/C_m$ where I is the total current flow (I=V/R) across the patch of membrane.

Figure 14 illustrates these concepts. For the sake of simplicity, the plasma membrane of the cell shown in Figure 14 is divided into four patches: left, top, right, and bottom. Current will flow between the electrodes if a voltage difference is applied. This will alter the cell membrane potential. If electrode 1 is positive and electrode 2 is negative, the membrane patch at the bottom of the cell will be hyperpolarized but the top patch will be depolarized. The left and right patches will "see" no change in membrane potential. If polarity is reversed, the opposite will occur.

In reality, of course, the cell's plasma membrane is a continuum of individual patches rather the simplified system of four patches depicted in Figure 14. The applied voltage alters the membrane potentials of the various patches to many different values such that the embedded voltage-gated ion channels "sample" the many different potentials and are driven through their various conformational states. These include open states, closed states, high affinity drug bound states, and low affinity drug bound states.

Accordingly, the present invention provides a method for identifying modulators of the activity of a voltage-gated ion channel comprising:

(a) altering the transmembrane potential of at least a portion of the membrane of a cell expressing the voltage-gated ion channel by applying a voltage to the cells through extracellular electrodes while monitoring ion flow through the voltage-gated ion channel;

(b) exposing the cell in step (a) to a substance and monitoring ion flow through the voltage-gated ion channel;

(c) comparing the ion flow through the voltage-gated ion channel in step (a) and step (b);

where a difference in the ion flow through the voltage-gated ion channel in step (a) and step (b) indicates that the substance is a modulator of the voltage-gated ion channel.

A variation of the method comprises:

(a) dividing a plurality of cells expressing the voltage-gated ion channel into a control portion and a test portion;

(b) altering the transmembrane potential of the control portion of cells by applying a voltage to the cells through extracellular electrodes while monitoring ion flow through the voltage-gated ion channel;

(c) altering the transmembrane potential of the test portion of cells by applying the voltage to the cells through extracellular electrodes in the presence of a substance while monitoring ion flow through the voltage-gated ion channel;

(d) comparing the ion flow through the voltage-gated ion channel in step (b) and step (c);

where a difference in the ion flow through the voltage-gated ion channel in step (b) and step (c) indicates that the substance is modulator of the voltage-gated ion channel.

For the sake of simplicity, the above methods are described in terms of "a" voltage-gated ion channel although those skilled in the art will understand that in actual practice the cells will express a plurality of the voltage-gated ion channels for which modulators are sought. Generally, each cell will express at least 10^2, 10^3, 10^4, 10^5, 10^6 or more molecules of the voltage-gated ion channel. Also, ion flow will be monitored through the plurality of the voltage-gated ion channels rather than through a single voltage-gated ion channel. Similarly, the methods will generally be practiced by employing a plurality of cells, even though the methods are described above in terms of "a" cell.

Generally, the methods of the present invention will be carried out on a substrate that is a modified version of a standard multiwell tissue culture plate or microtiter plate. Such substrates will have a place for the cells to be tested (generally the wells of the tissue culture plate or microtiter plate) and will have positive and negative electrodes (either built into the plate or nearby) in such an orientations with respect to the cells that the electrodes can deliver a voltage potential that causes an alteration in the open/close state of the voltage-gated ion channels in the cells. The electrodes are extracellular, *i.e.*, they do not penetrate into or across the plasma membranes of the cells although they may touch the outside of the plasma membranes in certain embodiments. Extracellular electrodes do not include electrodes which form a continuous connection with a cell's interior, *e.g.*, patch/clamp electrodes.

Therefore, the present invention provides a method of identifying activators of a voltage-gated ion channel comprising:

(a) providing a substrate upon which are living eukaryotic cells that express a plurality of the voltage-gated ion channels in their plasma membranes;

(b) providing positive and negative electrodes positioned either on or near the substrate such that when a preselected voltage is applied through the positive and negative electrodes the transmembrane electrical potential of the cells is altered such that at least a portion of the voltage-gated ion channels are closed;

(c) applying the preselected voltage through the positive and negative electrodes;

(d) determining a control value for the flow of ions through the voltage-gated ion channels of the cells in step (c);

(e) exposing the cells of step (c) to a substance for a period sufficient and under conditions such that a detectable number of the portion of the

voltage-gated ion channels that are closed become open and allow ion flow through the detectable number of voltage-gated ion channels if the substance is an activator of the voltage-gated ion channels;

(f) determining a test value for the flow of ions through the voltage-gated ion channels of the cells of step (e);

(g) comparing the control value to the test value;

where if the control value is less than the test value, then the substance is an activator of the voltage-gated ion channel.

The above-described method can be easily modified to provide a method for identifying inhibitors of the voltage-gated ion channel. The voltage applied through the electrodes is preselected such that it alters the electrical field around the cells and consequently alters the transmembrane electrical field. This in turn changes the states of the embedded voltage-gated ion channels such that instead of the voltage-gated ion channels being closed, the voltage-gated ion channels may open. Substances are then screened for the ability to close or inhibit the channels.

Accordingly, the present invention provides a method of identifying inhibitors of a voltage-gated ion channel comprising:

(a) providing a substrate upon which are living eukaryotic cells that express a plurality of the voltage-gated ion channels in their plasma membranes;

(b) providing positive and negative electrodes positioned either on or near the substrate such that when a preselected voltage is applied through the positive and negative electrodes the transmembrane electrical potential of the cells is altered such that at least a portion of the voltage-gated ion channels are open;

(c) applying the preselected voltage through the positive and negative electrodes;

(d) determining a control value for the flow of ions through the voltage-gated ion channels of the cells in step (c);

(e) exposing the cells of step (c) to a substance for a period sufficient and under conditions such that a detectable number of the portion of the voltage-gated ion channels that are open become closed and restrict ion flow through the detectable number of voltage-gated ion channels if the substance is an inhibitor of the voltage-gated ion channels;

(f) determining a test value for the flow of ions through the voltage-gated ion channels of the cells of step (e);

(g) comparing the control value to the test value;

where if the control value is greater than the test value, then the substance is an inhibitor of the voltage-gated ion channel.

In the above-described method for identifying activators, the terms "a portion of the voltage-gated ion channels are closed" and "a detectable number" are related and have relative rather than absolute values. Similarly, in the above-described method for identifying inhibitors, the terms "a portion of the voltage-gated ion channels are open" and "a detectable number" are also related and have relative rather than absolute values. What is meant is that a portion of the voltage-gated ion channels will be open or closed such that when the substance acts on the channels, a change in the open/closed state of a sufficient number of channels (*i.e.*, "a detectable number") occurs such that a difference in ion flow that is large enough to be measured by the detection system employed takes place. There is no need to determine the actual number of ion channels that constitutes the "portion" of voltage-gated ion channels that are closed or open or the "detectable number" so long as the difference in ion flow can be measured. The actual portion of channels that will be open or closed as well as the actual value of "detectable number" in order for the methods to be practiced will depend on such variables as the channel that is being studied, the concentrations of the substances tested, the nature of the detection system for ion flow, and so forth. Adjusting the voltage applied through the electrodes to take into account such variables so that control and test values can be obtained is a matter of routine experimentation in which the skilled artisan will be guided by knowledge in the art such as, *e.g.*, the known voltage dependence of the open/close transition of the voltage-gated ion channel under study, the nature and sensitivity of the detection system employed to monitor the flow of ions, the level of expression of the ion channel in the cells, and so forth.

The electrodes can be arranged in a variety of ways in order to provide for the proper stimulus. A number of arrangements are described herein and illustrated in the accompanying figures. These include arrangements where the cells are present in wells in the substrate and:

(a) both a positive and negative electrode is present in each well;

(b) one electrode is present in the well and the other electrode enters the fluid medium in the well from above without touching the sides or bottom of the well;

(c) the electrodes form part of the sides or bottom of the wells;

(d) a pattern of interdigitating electrodes has been formed on the surface of the substrate and at least some of the cells are positioned between the interdigitating branches of the positive and negative electrodes.

The skilled person will recognize that it is generally beneficial to run controls together with the methods described herein. For example, it will usually be helpful to have a control in which the substances are tested in the methods against cells that preferably are essentially identical to the cells that are used in the methods except that these cells would not express the voltage-gated ion channels of interest. In this way it can be determined that substances which are identified by the methods are really exerting their effects through the voltage-gated ion channels of interest rather than through some unexpected non-specific mechanism. One possibility for such control cells would be to use non-recombinant parent cells where the cells of the actual experiment express the voltage-gated ion channels of interest due to the recombinant expression of those voltage-gated ion channels of interest.

Other types of controls would involve taking substances that are identified by the methods of the present invention as activators or inhibitors of voltage-gated ion channels of interest and testing those substances in the methods of the prior art in order to confirm that those substances are also activators and inhibitors when tested in those prior art methods.

One skilled in the art would recognize that, where the present invention involves comparing control values for the flow of ions to test values for the flow of ions and determining whether the control values are greater or less than the test values, a non-trivial difference is sought. For example, if in the methods of identifying inhibitors, the control value were found to be 1% greater than the test value, this would not indicate that the substance is an inhibitor. Rather, one skilled in the art would attribute such a small difference to normal experimental variance. What is looked for is a significant difference between control and test values. For the purposes of this invention, a significant difference fulfills the usual requirements for a statistically valid measurement of a biological signal. For example, depending upon the details of the experimental arrangement, a significant difference might be a difference of at least 10%, preferably at least 20%, more preferably at least 50%, and most preferably at least 100%.

One skilled in the art would understand that the cells that give rise to the control values need not be physically the same cells that give rise to the test values, although that is possible. What is necessary is that the cells that give rise to the control values be substantially the same type of cell as the cells that give rise to the test values. A cell line that has been transfected with and expresses a certain voltage-gated ion channel could be used for both the control and test cells. Large numbers of such cells could be grown and a portion of those cells could be exposed to the substance and thus serve as the cells giving rise to the test value for ion flow while a portion would not be exposed to the substance and would thus serve as the cells giving rise to the control value for ion flow. No individual cell itself would be both control and test cell but the virtual identity of all the cells in the cell line ensures that the methods would nevertheless be reliable.

Accordingly, the present invention provides a method of identifying activators of a voltage-gated ion channel comprising:

(a) providing a substrate upon which are living eukaryotic cells that express a plurality of the voltage-gated ion channels in their plasma membranes;

(b) providing positive and negative electrodes positioned either on or near the substrate such that when a preselected voltage is applied through the positive and negative electrodes the transmembrane electrical potential of the cells is altered such that at least a portion of the voltage-gated ion channels are closed;

(c) applying the preselected voltage through the positive and negative electrodes to a control sample of the cells;

(d) determining a control value for the flow of ions through the voltage-gated ion channels of the control sample of the cells in step (c);

(e) applying the preselected voltage through the positive and negative electrodes to a test sample of the cells while exposing the test sample of the cells to a substance for a period sufficient and under conditions such that a detectable number of the portion of the voltage-gated ion channels that are closed in the test sample become open and allow ion flow through the detectable number of voltage-gated ion channels if the substance is an activator of the voltage-gated ion channels;

(f) determining a test value for the flow of ions through the voltage-gated ion channels of the test sample of cells of step (e);

(g) comparing the control value to the test value;

where if the control value is less than the test value, then the substance is an activator of the voltage-gated ion channel.

Similarly, the present invention provides a method of identifying inhibitors of a voltage-gated ion channel comprising:

(a) providing a substrate upon which are living eukaryotic cells that express a plurality of the voltage-gated ion channels in their plasma membranes;

(b) providing positive and negative electrodes positioned either on or near the substrate such that when a preselected voltage is applied through the positive and negative electrodes the transmembrane electrical potential of the cells is altered such that at least a portion of the voltage-gated ion channels are open;

(c) applying the preselected voltage through the positive and negative electrodes to a control sample of the cells;

(d) determining a control value for the flow of ions through the voltage-gated ion channels of the control sample of the cells in step (c);

(e) applying the preselected voltage through the positive and negative electrodes to a test sample of the cells while exposing the test sample of the cells to a substance for a period sufficient and under conditions such that a detectable number of the portion of the voltage-gated ion channels that are open in the test sample become closed and restrict ion flow through the detectable number of voltage-gated ion channels if the substance is an inhibitor of the voltage-gated ion channels;

(f) determining a test value for the flow of ions through the voltage-gated ion channels of the test sample of cells of step (e);

(g) comparing the control value to the test value;

where if the control value is greater than the test value, then the substance is an inhibitor of the voltage-gated ion channel.

"Substances" can be any substances that are generally screened in the pharmaceutical industry during the drug development process. For example, substances may be low molecular weight organic compounds (*e.g.*, having a molecular weight of less than about 1,000 daltons); RNA, DNA, antibodies, peptides, or proteins.

The conditions under which cells are exposed to substances in the methods described herein are conditions that are typically used in the art for the study of protein-ligand interactions: *e.g.*, physiological pH; salt conditions such as those represented by such commonly used buffers as PBS or in tissue culture media; a

temperature of about 4°C to about 55°C; incubation times of from several seconds to several hours. Generally, the cells are present in wells in the substrate and the substances are added directly to the wells, optionally after first washing away the media in the wells.

Determining the values of ion flow in the methods of the present invention can be accomplished through the use of fluorescent indicator compounds. One type of fluorescent indicator compound is sensitive to the level of intracellular calcium ions in the cells used in the present invention. This type of fluorescent indicator compound can be used when the methods are directed to those voltage-gated ion channels whose activity results in a change in intracellular calcium levels. Such voltage-gated ion channels include not only voltage-gated calcium channels but also other types of voltage-gated ion channels where the activity of those channels is naturally or can be coupled to changes in intracellular calcium levels. Many types of voltage-gated potassium channels can be so coupled. When using this approach to study a voltage-gated ion channel of interest that is not a voltage-gated calcium channel, it may be desirable to engineer the cells employed so as to recombinantly express voltage-gated calcium channels that are coupled to the voltage-gated ion channel of interest.

Fluorescent indicator compounds suitable for measuring intracellular calcium levels include various calcium indicator dyes (*e.g.*, fura-2, fluo-3, indo-1, Calcium Green; see Veliçelebi et al., 1999, Meth. Enzymol. 294:20-47).

Calcium indicator dyes are substances which show a change in a fluorescent characteristic upon binding calcium, *e.g.*, greatly increased intensity of fluorescence and/or a change in fluorescent spectra (*i.e.*, a change in emission or excitation maxima). Fluo-3, fura-2, and indo-1 are commonly used calcium indicator dyes that were designed as structural analogs of the highly selective calcium chelators ethylene glycol-bis(β-aminoethyl ether) N,N,N',N'-tetraacetic acid (EGTA) and 1,2-bis(2-aminophenoxy) ethane-N,N,N',N'-tetraacetic acid (BAPTA). The fluorescence intensity from fluo-3 increases by more than 100-fold upon binding of calcium. While the unbound dye exhibits very little fluorescence, calcium-bound fluo-3 shows strong fluorescence emission at 526 nm. Fura-2 is an example of a dye that exhibits a change in its fluorescence spectrum upon calcium binding. In the unbound state, fura-2 has an excitation maximum of 362 nm. This excitation maximum shifts to 335 nm upon calcium binding, although there is no change in emission maximum. Binding of

calcium to fura-2 can be monitored by excitation at the two excitation maxima and determining the ratio of the amount of fluorescence emission following excitation at 362 nm compared to the amount of fluorescence emission following excitation at 335 nm. A smaller ratio (*i.e.*, less emission following excitation at 362 nm) indicates that more fura-2 is bound to calcium, and thus a higher internal calcium concentration in the cell.

The use of calcium indicator dyes entails loading cells with the dye, a process which can be accomplished by exposing cells to the membrane-permeable acetoxymethyl esters of the dyes. Once inside the plasma membrane of the cells, intracellular esterases cleave the esters, exposing negative charges in the free dyes. This prevents the free dyes from crossing the plasma membrane and thus leaves the free dyes trapped in the cells. Measurements of fluorescence from the dyes are then made, the cells are treated in such a way that the internal calcium concentration is changed (*e.g.*, by exposing cells to an activator or inhibitor of a voltage-gated ion channel), and fluorescence measurements are again taken.

Fluorescence from the indicator dyes can be measured with a luminometer or a fluorescence imager. One preferred detection instrument is the Fluorometric Imaging Plate Reader (FLIPR) (Molecular Devices, Sunnyvale, CA). The FLIPR is well suited to high throughput screening using the methods of the present invention as it incorporates integrated liquid handling capable of simultaneously pipetting to 96 or 384 wells of a microtiter plate and rapid kinetic detection using a argon laser coupled to a charge-coupled device imaging camera.

A typical protocol for use of calcium indicator dyes would entail plating cells expressing a voltage-gated ion channel of interest into an appropriate substrate (*e.g.*, clear, flat-bottom, black-wall 96 well plates that have a suitable arrangement of positive and negative electrodes) and allowing the cells to grow overnight in standard tissue culture conditions (*e.g.*, 5% CO_2, 37°C). The cells are generally plated at a density of about 10,000 to 100,000 cells per well in appropriate growth medium. On the day of the assay, growth medium is removed and dye loading medium is added to the wells.

If the calcium indicator dye is fluo-3, *e.g.*, dye loading medium could be prepared by solubilizing 50 µg of fluo-3-AM ester (Molecular Probes F-1242) in 22 µl DMSO to give a 2 mM dye stock. Immediately before loading the cells, 22 µl 20% pluronic acid (Molecular Probes P-3000) is added to the dye. The tube

containing the dye is mixed with a vortex mixer and 42 ml of the dye/pluronic acid solution is added to 10.5 ml of Hanks Balanced Salt Solution (Gibco/BRL Cat # 14025-076) with 20 mM HEPES (Gibco/BRL Cat # 1560-080), 2.5 mM probenecid (Sigma Cat # P-8761), and 1% fetal bovine serum (Gibco/BRL Cat # 26140-087; not BSA)). The dye and the loading medium are mixed by repeated inversion (final dye concentration about 4 μM).

Growth medium can be removed from the cells by washing (wash medium is Hanks Balanced Salt Solution (Gibco/BRL Cat # 14025-076) with 20 mM HEPES (Gibco/BRL Cat # 1560-080), 2.5 mM probenecid (Sigma Cat # P-8761), and 0.1% bovine serum albumin (Sigma Cat # A-9647; not FBS) three times, leaving 100 μl residual medium in the wells after the fourth wash. Then 100 μl of the dye in the loading medium is added to each well. The cells are then incubated for 60 minutes to allow for dye loading.

Following dye loading, fluorescent measurements of the cells are taken prior to exposure of the cells to substances that are to be tested. The cells are then exposed to the substances and those substances that cause a change in a fluorescent characteristic of the dye are identified. The measuring instrument can be a fluorescent plate reader such as the FLIPR (Molecular Devices). Substances that cause a change in a fluorescent characteristic in the test cells but not the control cells are possible activators or inhibitors of the voltage-gated ion channel.

The exact details of the procedure outlined above are meant to be illustrative. One skilled in the art would be able to optimize experimental parameters (cell number, dye concentration, dye loading time, temperature of incubations, cell washing conditions, and instrument settings, etc.) by routine experimentation depending on the particular relevant experimental variables (*e.g.*, type of cell used, identity of dye used). Several examples of experimental protocols that can be used are described in Veliçelebi et al., 1999, Meth. Enzymol. 294:20-47. Other suitable instrumentation and methods for measuring transmembrane potential changes via optical methods includes microscopes, multiwell plate readers and other instrumentation that is capable of rapid, sensitive ~~ratiometric~~ fluorescence detection. For example, the VIPR (Aurora Biosciences, San Diego, CA) is an integrated liquid handler and kinetic fluorescence reader for 96-well and greater multiwell plates. The VIPR reader integrates an eight channel liquid handler, a multiwell positioning stage and a fiber-optic illumination and detection system. The system is designed to measure fluorescence from a column of eight wells simultaneously before, during and after the introduction of liquid

sample obtained from another microtiter plate or trough. The VIPR reader excites and detects emission signals from the bottom of a multiwell plate by employing eight trifurcated optical bundles (one bundle for each well). One leg of the trifurcated fiber is used as an excitation source, the other two legs of the trifurcated fiber being used to detect fluorescence emission. A ball lens on the end of the fiber increases the efficiency of light excitation and collection. The bifurcated emission fibers allow the reader to detect two emission signals simultaneously and are compatible with rapid signals generated by the FRET-based voltage dyes.

Photomultiplier tubes then detect emission fluorescence, enabling sub-second emission ratio detection.

In particular embodiments, the calcium indicator dye is selected from the group consisting of: fluo-3, fura-2, fluo-4, fluo-5, calcium green-1, Oregon green, 488 BAPTA, SNARF-1, and indo-1.

In particular embodiments, the change in fluorescent characteristic is an increase in intensity of a fluorescence emission maximum. In other embodiments, the change in fluorescent characteristic is a shift in the wavelength of an absorption maximum.

In particular embodiments, the cells naturally express the voltage-gated ion channel of interest and/or calcium channels. In other embodiments, the cells do not naturally express the voltage-gated ion channel of interest and/or calcium channels but instead have been transfected with expression vectors that encode the voltage-gated ion channel of interest and/or calcium channels so that the cells recombinantly express the voltage-gated ion channel of interest and/or calcium channels. Transfection is meant to include any method known in the art for introducing expression vectors into the cells. For example, transfection includes calcium phosphate or calcium chloride mediated transfection, lipofection, infection with a retroviral construct, and electroporation.

An alternative to the use of calcium indicator dyes is the use of the aequorin system. The aequorin system makes use of the protein apoaequorin, which binds to the lipophilic chromophore coelenterazine forming a combination of apoaequorin and coelenterazine that is known as aequorin. Apoaequorin has three calcium binding sites and, upon calcium binding, the apoaequorin portion of aequorin

changes its conformation. This change in conformation causes coelenterazine to be oxidized into coelenteramide, CO_2, and a photon of blue light (466 nm). This photon can be detected with suitable instrumentation.

Since the gene encoding apoaequorin has been cloned (U.S. Patent No. 5,541,309; U.S. Patent No. 5,422,266; U.S. Patent No. 5,744,579; Inouye et al., 1985, Proc. Natl. Acad. Sci. USA 82:3154-3158; Prasher et al., 1985, Biochem. Biophys. Res. Comm. 126:1259-1268), apoaequorin can be recombinantly expressed in cells in which it is desired to measure the intracellular calcium concentration. Alternatively, existing cells that stably express recombinant apoaequorin can be used. Such cells derived from HEK293 cells and CHO-K1 cells are described in Button & Brownstein, 1993, Cell Calcium 14:663-671. For example, the HEK293/aeq17 cell line can be used as follows.

The HEK293/aeq17 cells are grown in Dulbecco's Modified Medium (DMEM, GIBCO-BRL, Gaithersburg, MD, USA) with 10% fetal bovine serum (heat inactivated), 1 mM sodium pyruvate, 500 μg/ml Geneticin, 100 μg/ml streptomycin, 100 units/ml penicillin. Expression vectors encoding the voltage-gated ion channel of interest as well as, optionally, the desired voltage-gated calcium channel subunits (α 1A, α 1B, α 1C, α 1D, α 1E, α 1G, α 1H, α 1I, $\alpha_2\delta$, β_1, β_2, β_3, β_4, etc.) can be transfected into the HEK293/aeq17 cells by standard methods in order to express the desired voltage-gated ion channel subunits and voltage-gated calcium channel subunits in the HEK293/aeq17 cells. The cells are washed once with DMEM plus 0.1 % fetal bovine serum, and then charged for one hour at 37°C /5% CO_2 in DMEM containing 8 μM coelenterazine cp (Molecular Probes, Eugene, OR, USA) and 30 μM glutathione. The cells are then washed once with Versene (GIBCO-BRL, Gaithersburg, MD, USA), detached using Enzyme-free cellissociation buffer (GIBCO-BRL, Gaithersburg, MD, USA), diluted into ECB (Ham's F12 nutrient mixture (GIBCO-BRL) with 0.3 mM $CaCl_2$, 25 mM HEPES, pH7.3, 0.1% fetal bovine serum). The cell suspension is centrifuged at 500 x g for 5 min. The supernatant is removed, and the pellet is resuspended in 10 ml ECB. The cell density is determined by counting with a hemacytometer and adjusted to 500,000 cells/ml in ECB. The substances to be tested are diluted to the desired concentrations in ECB and aliquoted into the assay plates, preferably in triplicate, at 0.1 ml/well. The cell suspension is injected at 0.1 ml/well, read and integrated for a total of 400 readings using a luminometer (Luminoskan Ascent, Labsystems Oy, Helsinki, Finland).

Alternatively, the cells may first be placed into the assay plates and then the substances added. Data are analyzed using the software GraphPad Prism Version 3.0 (GraphPad Software, Inc., San Diego, CA, USA).

It will be understood by those skilled in the art that the procedure outlined above is a general guide in which the various steps and variables can be modified somewhat to take into account the specific details of the particular assay that is desired to be run. For example, one could use semisynthetic coelenterazine (Shimomura, 1989, Biochem. J. 261:913-920; Shimomura et al., 1993, Cell Calcium 14:373-378); the time of incubation of the cells with coelenterazine can be varied somewhat; somewhat greater or lesser numbers of cells per well can be used; and so forth.

For reviews on the use of aequorin, see Créton et al., 1999, Microscopy Research and Technique 46:390-397; Brini et al., 1995, J. Biol. Chem. 270:9896-9903; Knight & Knight, 1995, Meth. Cell. Biol. 49:201-216. Also of interest may be U.S. Patent No. 5,714,666 which describes methods of measuring intracellular calcium in mammalian cells by the addition of coelenterazine co-factors to mammalian cells that express apoaequorin.

Another way to measure ion flow is to monitor changes in transcription that result from the activity of voltage-gated ion channels by the use of transcription based assays. Transcription-based assays involve the use of a reporter gene whose transcription is driven by an inducible promoter whose activity is regulated by a particular intracellular event such as, *e.g.*, changes in intracellular calcium levels, that are caused by the activity of a voltage-gated ion channel. Transcription-based assays are reviewed in Rutter et al., 1998, Chemistry & Biology 5:R285-R290. Transcription-based assays of the present invention rely on the expression of reporter genes whose transcription is activated or repressed as a result of intracellular events that are caused by the interaction of a activator or inhibitor with a voltage-gated ion channel.

An extremely sensitive transcription-based assay is disclosed in Zlokarnik et al., 1998, Science 279:84-88 (Zlokarnik) and also in U.S. Patent No. 5,741,657. The assay disclosed in Zlokarnik and U.S. Patent No. 5,741,657 employs a plasmid encoding β-lactamase under the control of an inducible promoter. This plasmid is transfected into cells together with a plasmid encoding a receptor for which it is desired to identify agonists. The inducible promoter on the β-lactamase is chosen

so that it responds to at least one intracellular signal that is generated when an agonist binds to the receptor. Thus, following such binding of agonist to receptor, the level of β-lactamase in the transfected cells increases. This increase in β-lactamase is measured by treating the cells with a cell-permeable dye that is a substrate for cleavage by β-lactamase. The dye contains two fluorescent moieties. In the intact dye, the two fluorescent moieties are physically linked, and thus close enough to one another that fluorescence resonance energy transfer (FRET) can take place between them. Following cleavage of the dye into two parts by β-lactamase, the two fluorescent moieties are located on different parts, and thus can diffuse apart. This increases the distance between the fluorescent moieties, thus decreasing the amount of FRET that can occur between them. It is this decrease in FRET that is measured in the assay.

The assay described in Zlokarnik and U.S. Patent No. 5,741,657 can be modified for use in the methods of the present invention by using an inducible promoter to drive β-lactamase where the promoter is activated by an intracellular signal generated by the opening or closing of a voltage-gated ion channel. Cells expressing a voltage-gated ion channel and the inducible promoter-driven β-lactamase are placed in the apparatus of the present invention, where the open or closed state of the voltage-gated ion channels can be controlled. The cells are exposed to the cell-permeable dye and then exposed to substances suspected of being activators or inhibitors of the voltage-gated ion channel. Those substances that cause a change in the open or closed state of the voltage-gated ion channel are identified by their effect on the inducible promoter-driven β-lactamase and thus on FRET. The inducible promoter-driven β-lactamase is engineered with a suitable promoter so that β-lactamase is induced when the substance is either an activator or an inhibitor, depending upon the nature of the assay.

The flow of ions through voltage-gated ion channels can also be measured by measuring changes in membrane potential via the use of fluorescent voltage sensitive dyes. The changes in membrane potential will depend on the ion channels in the cell membrane. The resultant membrane potential will depend on the net properties of all the channels and the change caused by inhibiting (through a substance that is an inhibitor or antagonist) or activating (through a substance that is an activator or an agonist) the voltage-gated ion channel of interest. One knowledgeable in cellular and membrane biophysics and electrophysiology will

understand the directions of the changes in membrane potential since those changes depend on the ion channels present and the inhibition or activation of those channels by test substances. In many cases when using fluorescent voltage sensitive dyes, the experimental system can be calibrated by using known activators or inhibitors of the voltage-gated ion channel of interest.

The present invention therefore includes assays that monitor changes in ion flow caused by activators or inhibitors of voltage-gated ion channels based upon FRET between a first and a second fluorescent dye where the first dye is bound to one side of the plasma membrane of a cell expressing a voltage-gated ion channel of interest and the second dye is free to move from one face of the membrane to the other face in response to changes in membrane potential. In certain embodiments, the first dye is impenetrable to the plasma membrane of the cells and is bound predominately to the extracellular surface of the plasma membrane. The second dye is trapped within the plasma membrane but is free to diffuse within the membrane. At normal (*i.e.*, negative) resting potentials of the membrane, the second dye is bound predominately to the inner surface of the extracellular face of the plasma membrane, thus placing the second dye in close proximity to the first dye. This close proximity allows for the generation of a large amount of FRET between the two dyes. Following membrane depolarization, the second dye moves from the extracellular face of the membrane to the intracellular face, thus increasing the distance between the dyes. This increased distance results in a decrease in FRET, with a corresponding increase in fluorescent emission derived from the first dye and a corresponding decrease in the fluorescent emission from the second dye. See figure 1 of González & Tsien, 1997, Chemistry & Biology 4:269-277. See also González & Tsien, 1995, Biophys. J. 69:1272-1280 and U.S. Patent No. 5,661,035.

In certain embodiments, the first dye is a fluorescent lectin or a fluorescent phospholipid that acts as the fluorescent donor. Examples of such a first dye are: a coumarin-labeled phosphatidylethanolamine (*e.g.*, N-(6-chloro-7-hydroxy-2-oxo-2H--1-benzopyran-3-carboxamidoacetyl)-dimyristoylphosphatidyl-ethanolamine) or N-(7-nitrobenz-2-oxa-1,3-diazol-4-yl)-dipalmitoylphosphatidylethanolamine); a fluorescently-labeled lectin (*e.g.*, fluorescein-labeled wheat germ agglutinin). In certain embodiments, the second dye is an oxonol that acts as the fluorescent acceptor. Examples of such a second dye are: bis(1,3-dialkyl-2-thiobarbiturate)trimethineoxonols (*e.g.*, bis(1,3-dihexyl-2-

thiobarbiturate)trimethineoxonol) or pentamethineoxonol analogues (*e.g.*, bis(1,3-dihexyl-2-thiobarbiturate)pentamethineoxonol; or bis(1,3-dibutyl-2-thiobarbiturate)pentamethineoxonol). See González & Tsien, 1997, Chemistry & Biology 4:269-277 for methods of synthesizing various dyes suitable for use in the present invention. In certain embodiments, the assay may comprise a natural carotenoid, *e.g.*, astaxanthin, in order to reduce photodynamic damage due to singlet oxygen.

The use of such fluorescent dyes capable of moving from one face of the plasma membrane to the other is especially appropriate when the methods of the present invention are directed to inwardly rectifying potassium channels. Activation of inwardly rectifying potassium channels results in increased potassium current flow across the plasma membrane. This increased current flow results in a hyperpolarization of the cell membrane that can be detected by use of the technique described above since such hyperpolarization will result in greater FRET.

A large number of possible combinations of types of substrates and electrodes; physical arrangement of electrodes; number, shape, and arrangement of wells for holding the cells are suitable for use in the present invention.

Figure 1 illustrates an embodiment of the invention where the electrodes are generally parallel wires or strips of conductive material such as gold. The electrodes lie on the surface of a glass substrate and, together with the spacers, form the walls of the wells. For clarity, only a single series of wells is shown in Figure 1. Generally, substantially the entire surface of the glass substrate would be covered by wells formed in the manner shown. Cells are placed in the wells and grown in suitable media until an appropriate number of cells is present in the wells. Alternatively, an appropriate number of cells may be placed into the wells and used without further growth.

Figure 2B illustrates an embodiment of the invention where the wells are cavities or depressions in the surface of the substrate, as in typical multiwell tissue culture plates. Each well has an electrode at the bottom of the well and another electrode that is aligned along a side of the well. The cells are shown in Figure 2B as attached at the bottom of the well but in certain embodiments the cells may be suspension cells dispersed in the fluid in the well.

Figure 2C illustrates an embodiment of the invention similar to that shown in Figure 2B except that in Figure 2C both electrodes are at the bottom of the wells.

Figure 3 illustrates an embodiment of the invention where an array of interdigitating transparent electrodes has been chemically etched onto the surface of a glass substrate. The electrode array, comprising a comb of positive and negative electrodes, has been chemically etched onto an indium tin oxide (ITO) coated glass plate. The thin layer of ITO (about 200 Å to 2,000 Å, or 500 Å to 1,500 Å, preferably 1,200 Å thick) forms a transparent conductive coating on the surface of the glass. Although not essential, it is preferred that the layer of ITO be thin enough to be transparent. The chemical etching process removes the ITO from selected areas, resulting in an array of transparent ITO electrodes bonded to the glass. Multiple reaction wells may be contained on a single glass plate by forming fluid retention wells at the different electrode array sites. The wells can be formed by attaching (*e.g.*, gluing) a well frame to the glass substrate or by forming virtual wells on the glass plate by a method such as screening hydrophobic ink onto the plate.

Figure 4A and 4B illustrates an embodiment in which wells are formed by attaching a well frame onto the substrate.

Figure 6 illustrates an embodiment in which a droplet of fluid containing cells that express a voltage-gated ion channel is sandwiched between two plates. The plates, which can be glass plates, are each coated with a thin layer of conductive material such as indium tin oxide (ITO). The layers of conductive material are connected to a pulse generator such that one layer functions as a positive electrode and the other layer functions as a negative electrode.

Figures 7 and 8 illustrate embodiments in which one of the electrodes enters the well from above. In Figures 9 and 10, both electrodes enter from above.

The substrates for use in the present invention may contain virtual wells. Virtual wells are formed when a surface is patterned to have relatively hydrophilic domains within relatively hydrophobic fields so that an aqueous sample is physically constrained by surface tension to the more hydrophilic domains by the edges of the more hydrophobic fields. The hydrophilic domains can be small circles that form a pattern similar to the wells of a conventional microtiter plate. Virtual wells provide a location in which samples can be confined without the deep indentations found in conventional microtiter plates. Figure 5 illustrates a surface for

use in the present invention that is a derivatized glass surface upon which virtual wells have been formed and upon which a pattern of interdigitated electrodes has also been formed. Figure 3 shows an individual well from this surface. International Patent Publication WO 99/39829 describes virtual wells and how they can be made.

"Interdigitating" refers to an arrangement of positive and negative electrodes where the positive and negative electrodes contain branches that are arranged such that, if a line were drawn from one branch of a positive electrode to the adjacent branch of the positive electrode, the line would cross a branch of the negative electrode. Similarly, if a line were drawn from one branch of a negative electrode to the adjacent branch of the negative electrode, the line would cross a branch of the positive electrode. Generally, each interdigitating positive or negative electrode has at least 2, or at least 4, or at least 10, or at least 20 interdigitating branches. An example of interdigitating electrodes is shown in Figure 3.

Various additional arrangements of electrodes formed from conductive materials on glass substrates are possible. One arrangement has the positive and negative electrodes formed on two parallel glass substrates. For example, instead of having the positive and negative electrodes on a single glass substrate, two ITO coated glass substrates can be utilized by placing the glass substrates parallel to one another and placing the biologic fluid containing the cells in the gap between the glass substrates. In this arrangement, one conductive glass substrate serves as the positive electrode while the second glass substrate serves as the negative electrode. The electrode field is formed at a right angle to the surface of the plates. This arrangement would allow fluid containing the cells to be either dispensed in between the plates or drawn into the gap via capillary action. The detector's light beam would enter perpendicular to the glass substrates and pass into the gap between the glass substrates, illuminating the fluid and cells. The fluorescence transmission from the cells would be collected by the detector in a similar manner. Figure 6 illustrates one version of this arrangement. Another version is shown in Figure 13 where an embodiment comprising two ITO-coated plates each containing multiple virtual wells is depicted. The ITO forms the bottom of the wells as well as the electrodes.

Another arrangement has the positive and negative electrodes formed by a single glass substrate and a reference electrode. This arrangement utilizes a single glass substrate coated with a conductive material such as ITO as one electrode. A well holding the biological fluid and cells is formed on the surface of the

conductive material coating the glass substrate. A wire or similar conducting member placed into the well serves as the second electrode. Figure 7 illustrates a single well of a version of this arrangement. Figure 12 depicts this type of arrangement as it is usually practiced, in a multiwell format. Figure 15 shows a modification of this arrangement where one electrode is a highly conductive metal grid that is in contact with the ITO layer.

Another arrangement has the single conductive glass substrate acting as the conductor to the current generated by a positive and negative electrode pair placed in adjacent wells. See Figure 16A-D. This arrangement does not use a grounding grid. The current flows from a first electrode in a first well through the ITO bottom of the first well to the ITO bottom of an adjacent second well and through a second electrode in the second well. Adjacent electrodes are alternately positive and negative. See Figure 16A and 16C.

In certain embodiments using interdigitating electrodes, the spacing and width of the branches of the electrodes are on the same order of magnitude as the size of individual cells. Cells may be grown and attached to the substrate in such a manner that, if a cell attaches between a pair of positive and negative electrode branches, a lower applied stimulus pulse can be utilized. The advantage of this close electrode spacing is that it results in less shunting of the stimulus current pulse through the fluid medium and less fluid heating while stimulating the cells. The use of transparent interdigitating electrodes offers the advantage of passing light from a fluorescent emission light source through the preferably glass substrate and transparent electrodes onto the cell and light passage of the fluorescence signal back to the light detector. While making the electrodes from a transparent material such as indium tin oxide (ITO) has advantages in certain embodiments, the electrodes may also be made from non-transparent conductive materials such as platinum, silver, or gold. If the electrode material is not transparent, fluorescence measurements are still possible because light can pass through the glass in between the electrodes.

Regardless of the arrangement of electrodes, stimulus pulses are generated by a pulse generator and applied to either a single well electrode array or to multiple well electrode arrays. Various commercial pulse generators can be utilized that permit waveform generation and amplitude adjustment. Constant voltage or constant current waveforms can be applied to the electrodes. Commercially available

power supplies that can be used in the present invention include the STG 1004 or STG 1008 Stimulus Generator or the National Instruments PCI 6713 8 channel pcb.

In using the pulse generator to stimulate the cells, particular attention should be paid to the amplitude, pulse width, and polarity used. For certain extreme field strengths, electroporation of the biological membrane can occur, and this should be avoided. When changing the external electrical field, the desired goal is a change in the trans-membrane field (Vm) by less than approximately $\pm$ 100 mV. As such the amount of charge added or removed from the cell membrane capacitance is critical. Adjustment of the pulse amplitude and duration is necessary to ensure a change in Vm without electroporation of the cells. Typically the voltage changes across the electrodes may be on the order of $\pm$ 10 volts, preferably less than $\pm$ 5 volts, and if possible less than $\pm$ 1 volt. These values can be adjusted empirically, by routine experimentation, in order to optimize the cellular membrane potential change without electroporation of the cell membrane. In general, the amount of charge change on the cell membrane will depend upon the local field changes, which depend upon the electrical current. Adjusting the area (the current-time integral) of the applied current as determined by the change in external electric field can be readily optimized empirically. In general, if the goal is to stimulate a cellular action potential, the pulse duration will be kept brief and the amplitude will be increased up to a point that exceeds the threshold for action potential generation. This will be affected by the relative levels of ion channels expressed in the cells and will vary accordingly, requiring empirical adjustment. A typical value might be a pulse duration of 1 millisecond and a pulse amplitude of 5 volts; this might be varied to increase the duration to 2 milliseconds and decrease the amplitude to 2.5 volts, or to decrease the duration and increase the amplitude, etc. In general, there is an inverse parabolic relationship between the duration and the amplitude of the applied pulse, where the area of the applied current-time integral remains constant. Because ion channel kinetics and action potentials can be rapid and brief, minimizing the pulse duration is useful. These parameters will also depend upon the manufactured electrodes, their capacitance and resistance, the geometrical relationship to the cells, the ionic strength and composition of the solutions used, and the electrical coupling to the cells. Because of these many variables, an empirical approach based upon the above guidelines is best.

Electrode arrangements can be adapted to 12-well, 24-well, 96-well, 384-well, 1,536-well, 3,456-well, and other plate formats, permitting the present invention to be used in high throughput screening applications.

In embodiments of the invention such as that illustrated in Figure 12 where multiple wells are present in the substrate and each well has an electrode associated with it, the stimulus delivered to each well through the electrodes can be individually controlled by the application of suitable software that governs the pulse generator. Such software is well known in the art or can be readily designed by one skilled in the art.

Particular embodiments of the present invention employ an arrangement of electrodes and wells on a substrate such that the substrate has the same form factor as a typical multiwell tissue culture plate that is used for high throughput screening, *e.g.*, a 96 well plate. The spacing of the wells on the substrate can be such that the center-to-center distances of the wells on the substrate is the same as the typical center-to-center distances between wells on typical 96 well plates that are used for high throughput screening. This facilitates the use of the present invention with current equipment used in high throughput screening such as plate handlers, detectors, automatic pipettors, etc. Substrates can be manufactured by modifying the well-known manufacturing processes generally used to make multiwell tissue culture plates by adding electrodes to the plates according to one of the configurations of electrodes disclosed herein.

In particular embodiments of the present invention, the substrate is not silicon or a field effect transistor.

In particular embodiments of the present invention, cells are utilized that have been transfected with expression vectors comprising DNA that encodes a voltage-gated ion channel. Preferably, the cells do not naturally express corresponding voltage-gated ion channels. For example, if the expression vectors direct the expression of a voltage-gated calcium channel, the cells will not naturally express voltage-gated calcium channels. Alternatively, if the cells naturally express corresponding voltage-gated ion channels, those corresponding voltage-gated ion channels can be distinguished from the transfected voltage-gated ion channels in some manner, *e.g.*, by the use of appropriate inhibitors, by manipulation of membrane potential. A preferred cell line for use in the present invention is the HEK293 cell line (ATCC 1573) since this cell line naturally expresses endogenous potassium

channels, which may be beneficial for electrical field stimulation experiments with channels that cause membrane potential depolarization (*e.g.*, sodium or calcium channels).

Cells are generally eukaryotic cells, preferably mammalian cells. The cells may be grown to the appropriate number on the substrates or they may be placed on the substrate and used without further growth. The cells may be attached to the substrate or, in those embodiments where the cells are placed or grown in wells, the cells may be suspension cells that are suspended in the fluid in the wells. Primary cells or established cell lines may be used.

Suitable cells for transfection with expression vectors that direct the expression of voltage-gated ion channels include but are not limited to cell lines of human, bovine, porcine, monkey and rodent origin. The cells may be adherent or non-adherent. Cells and cell lines which are suitable and which are widely available, include but are not limited to: L cells L-M(TK$^-$) (ATCC CCL 1.3), L cells L-M (ATCC CCL 1.2), HEK293 (ATCC CRL 1573), Raji (ATCC CCL 86), CV-1 (ATCC CCL 70), COS-1 (ATCC CRL 1650), COS-7 (ATCC CRL 1651), CHO-K1 (ATCC CCL 61), 3T3 (ATCC CCL 92), NIH/3T3 (ATCC CRL 1658), HeLa (ATCC CCL 2), C127I (ATCC CRL 1616), BS-C-1 (ATCC CCL 26), MRC-5 (ATCC CCL 171), CPAE (ATCC CCL 209), Saos-2 (ATCC HTB-85), ARPE-19 human retinal pigment epithelium (ATCC CRL-2302), GH3 cells, and primary cardiac myocytes.

A variety of voltage-gated ion channels may be used in the present invention. For example, voltage-gated sodium channels, voltage-gated potassium channels, and voltage-gated calcium channels are suitable.

In certain embodiments of the present invention, the cells used do not naturally express the voltage-gated ion channel of interest. Instead, DNA encoding the voltage-gated ion channel is transfected into cells in order to express the voltage-gated ion channel in the plasma membrane of the cells. DNA encoding voltage-gated ion channels can be obtained by methods well known in the art. For example, a cDNA fragment encoding a voltage-gated ion channel can be isolated from a suitable cDNA library by using the polymerase chain reaction (PCR) employing suitable primer pairs. The cDNA fragment encoding the voltage-gated ion channel can then be cloned into a suitable expression vector. Primer pairs can be selected based upon the known DNA sequence of the voltage-gated ion channel it is desired to obtain.

Suitable cDNA libraries can be made from cellular or tissue sources known to contain mRNA encoding the voltage-gated ion channel.

One skilled in the art would know that for certain voltage-gated ion channels, it is desirable to transfect, and thereby express, more than one subunit in order to obtain a functional voltage-gated ion channel. For example, N-type calcium channels are composed of a multisubunit complex containing at least an α1B, an α2δ, and a β1 subunit. On the other hand, T-type calcium channels are functional with only a single subunit, *e.g.*, α1G, α1H, or α1I. Common knowledge in the art of the subunit composition of a voltage-gated ion channel of interest will lead the skilled artisan to express the correct subunits in the transfected cells.

One skilled in the art could use published voltage-gated ion channel sequences to design PCR primers and published studies of voltage-gated ion channel expression to select the appropriate sources from which to make cDNA libraries in order to obtain DNA encoding the voltage-gated ion channels. The following publications may be of use in this regard:

U.S. Patent No. 5,380,836 describes nucleic acid sequences encoding a rat cardiac voltage-gated sodium channel;

U.S. Patent No. 6,030,810 describes a number of voltage-gated, tetrodotoxin-sensitive sodium channels;

U.S. Patent No. 6,184,349 B1 discloses a human tetrodotoxin-resistant peripheral nerve voltage-gated sodium channel known as PN3; see also GenBank accession no. AF117907;

Isom et al., 1994, Neuron 12:1183-1194 discloses a rat voltage-gated sodium channel β subunit;

McClatchey et al., 1993, Hum. Molec. Gen. 2:745-749 discloses a human voltage-gated sodium channel β1 subunit (hSCNβ1);

Isom et al., Science, 1992, 256:839-842 discloses a rat brain voltage-gated sodium channel β1 subunit (rSCNβ1);

Misgeld et al., 1995, Prog. Neurobiol. 46:423-462; North, 1989, Br. J. Pharmacol. 98:13-23; Gahwiler et al.,1985, Proc. Natl. Acad. Sci USA 82:1558-1562; and Andrade et al., 1986, Science 234:1261-1265 disclose inwardly rectifying voltage-gated potassium channels that are suitable for use in the methods of the present invention.

U.S. Patent No. 5,874,236 and U.S. Patent No. 5,429,921 describe various α1 and β subunits of human voltage-gated calcium channels;

U.S. Patent No. 5,407,820 and U.S. Patent No. 5,710,250 describe α2 subunits of human voltage-gated calcium channels;

International Patent Publication WO 98/13490 describes a brain-specific P/Q-type human voltage-gated calcium channel involved in familial hemiplagic migraine;

Table 1 provides a list of ion channel genes that are suitable for use in the present invention.

TABLE 1

Some ion channel genes of interest for EFS experiments				
Symbol	Full Name	Cytogenetic Location	MIM Number	PubMed ID
SCN1	symbol withdrawn, see SCN1A			
SCN1A	sodium channel, voltage-gated, type I, alpha polypeptide	2q24	182389	8062593
SCN1B	sodium channel, voltage-gated, type I, beta polypeptide	19	600235	8394762
SCN2A1	sodium channel, voltage-gated, type II, alpha 1 polypeptide	2q22-q23	182390	1317301
SCN2A2	sodium channel, voltage-gated, type II, alpha 2 polypeptide	2q23-q24	601219	1317301
SCN2A	symbol withdrawn, see SCN2A1	-		
SCN2B	sodium channel, voltage-gated, type II, beta polypeptide	11q22-qter	601327	10198179
SCN3A	sodium channel, voltage-gated, type III, alpha polypeptide	2q24	182391	9589372
SCN4A	sodium channel, voltage-gated, type IV, alpha polypeptide	17q23-q25.3	603967	1654742
SCN4B	sodium channel, voltage-gated, type IV, beta polypeptide	reserved		
SCN5A	sodium channel, voltage-gated, type V, alpha polypeptide (long (electrocardiographic) QT syndrome 3)	3p21	600163	
SCN6A	sodium channel, voltage-gated, type VI, alpha polypeptide	2q21-q23	182392	10198179
SCN7A	symbol withdrawn, see SCN6A	-		
SCN8A	sodium channel, voltage gated, type VIII, alpha polypeptide	12q13.1	600702	7670495
SCN9A	sodium channel, voltage-gated, type IX, alpha polypeptide	2q24	603415	7720699

TABLE 1 (Continued)

Some ion channel genes of interest for EFS experiments				
Symbol	Full Name	Cytogenetic Location	MIM Number	PubMed ID
SCN10A	sodium channel, voltage-gated, type X, alpha polypeptide	3p21-p22	604427	9839820
SCN11A	sodium channel, voltage-gated, type XI, alpha polypeptide	3p21-p24	604385	10444332
SCN12A	sodium channel, voltage-gated, type XII, alpha polypeptide	3p23-p21.3		10623608
SCNN1	symbol withdrawn, see SCNN1A	-		
SCNN1A	sodium channel, nonvoltage-gated 1 alpha	12p13	600228	7896277
SCNN1B	sodium channel, nonvoltage-gated 1, beta (Liddle syndrome)	16p12.2-p12.1		600760
SCNN1D	sodium channel, nonvoltage-gated 1, delta	1p36.3-p36.2	601328	8661065
SCNN1G	sodium channel, nonvoltage-gated 1, gamma	16p12	600761	7490094
CACNA1A	calcium channel, voltage-dependent, P/Q type, alpha 1A subunit	19p13	601011	8825650
CACNA1B	calcium channel, voltage-dependent, L type, alpha 1B subunit	9q34	601012	8825650
CACNA1C	calcium channel, voltage-dependent, L type, alpha 1C subunit	12pter-p13.2	114205	1650913
CACNA1D	calcium channel, voltage-dependent, L type, alpha 1D subunit	3p14.3	114206	1664412
CACNA1E	calcium channel, voltage-dependent, alpha 1E subunit	1q25-q31	601013	8388125
CACNA1F	calcium channel, voltage-dependent, alpha 1F subunit	Xp11.23-p11.22	300110	9344658
CACNA1G	calcium channel, voltage-dependent, alpha 1G subunit	17q22	604065	9495342

TABLE 1 (Continued)

Some ion channel genes of interest for EFS experiments				
Symbol	Full Name	Cytogenetic Location	MIM Number	PubMed ID
CACNA1H	calcium channel, voltage-dependent, alpha 1H subunit	16p13.3		9670923
CACNA1I	calcium channel, voltage-dependent, alpha 1I subunit	22q12.3-13.2		10454147
CACNA1S	calcium channel, voltage-dependent, L type, alpha 1S subunit	1q31-q32	114208	7916735
CACNA2	symbol withdrawn, see CACNA2D1	-		
CACNA2D1	calcium channel, voltage-dependent, alpha 2/delta subunit 1	7q21-q22	114204	8188232
CACNA2D2	calcium channel, voltage-dependent, alpha 2/delta subunit 2	reserved		
CACNB1	calcium channel, voltage-dependent, beta 1 subunit	17q21-q22	114207	8381767
CACNB2	calcium channel, voltage-dependent, beta 2 subunit	10p12	600003	9254841
CACNB3	calcium channel, voltage-dependent, beta 3 subunit	12q13	601958	8119293
CACNB4	calcium channel, voltage-dependent, beta 4 subunit	2q22-q31	601949	9628818
CACNG1	calcium channel, voltage-dependent, gamma subunit 1	17q24	114209	8395940
CACNG2	calcium channel, voltage-dependent, gamma subunit 2	reserved	602911	
CACNG3	calcium channel, voltage-dependent, gamma subunit 3	reserved		
CACNG4	calcium channel, voltage-dependent, gamma subunit 4	17q24		10613843
CACNG5	calcium channel, voltage-dependent, gamma subunit 5	17q24		10613843

TABLE 1 (Continued)

Some ion channel genes of interest for EFS experiments				
Symbol	Full Name	Cytogenetic Location	MIM Number	PubMed ID
CACNG6	calcium channel, voltage-dependent, gamma subunit 6	19q13.4		11170751
CACNG7	calcium channel, voltage-dependent, gamma subunit 7	19q13.4		11170751
CACNG8	calcium channel, voltage-dependent, gamma subunit 8	19q13.4		11170751
KCNA1	potassium voltage-gated channel, shaker-related subfamily, member 1 (episodic ataxia with myokymia)	12p13	176260	1349297
KCNA1B	literature alias, see KCNAB1	-		
KCNA2	potassium voltage-gated channel, shaker-related subfamily, member 2	12	176262	
KCNA2B	literature alias, see KCNAB2	-		
KCNA3	potassium voltage-gated channel, shaker-related subfamily, member 3	1p13.3 or 13	176263	2251283
KCNA3B	literature alias, see KCNAB3	-		
KCNA4	potassium voltage-gated channel, shaker-related subfamily, member 4	11p14	176266	2263489
KCNA4L	potassium voltage-gated channel, shaker-related subfamily, member 4-like	11q14		8449523
KCNA5	potassium voltage-gated channel, shaker-related subfamily, member 5	12	176267	
KCNA6	potassium voltage-gated channel, shaker-related subfamily, member 6	reserved	176257	
KCNA7	potassium voltage-gated channel, shaker-related subfamily, member 7	19	176268	
KCNA8	literature alias, see KCNQ1	-		

TABLE 1 (Continued)

Some ion channel genes of interest for EFS experiments				
Symbol	Full Name	Cytogenetic Location	MIM Number	PubMed ID
KCNA9	symbol withdrawn, see KCNQ1	-		
KCNA10	potassium voltage-gated channel, shaker-related subfamily, member 10	reserved	602420	
KCNAB1	potassium voltage-gated channel, shaker-related subfamily, beta member 1	3q26.1	601141	8838324
KCNAB2	potassium voltage-gated channel, shaker-related subfamily, beta member 2	1p36.3	601142	8838324
KCNAB3	potassium voltage-gated channel, shaker-related subfamily, beta member 3	17p13.1	604111	9857044
KCNB1	potassium voltage-gated channel, Shab-related subfamily, member 1	20q13.2	600397	7774931
KCNB2	potassium voltage-gated channel, Shab-related subfamily, member 2	8		9612272
KCNC1	potassium voltage-gated channel, Shaw-related subfamily, member 1	11p15	176258	8449507
KCNC2	potassium voltage-gated channel, Shaw-related subfamily, member 2	12 and 19q13.4	176256	8111118
KCNC3	potassium voltage-gated channel, Shaw-related subfamily, member 3	19	176264	1740329
KCNC4	potassium voltage-gated channel, Shaw-related subfamily, member 4	1p21	176265	1920536
KCND1	potassium voltage-gated channel, Shal-related subfamily, member 1	Xp11.23-p11.3	300281	10729221
KCND2	potassium voltage-gated channel, Shal-related subfamily, member 2	7q31-32	605410	10551270
KCND3	potassium voltage-gated channel, Shal-related subfamily, member 3	1p13.2	605411	10942109
KCNE1	potassium voltage-gated channel, Isk-related family, member 1	21q22.1-q22.2	176261	8432548

TABLE 1 (Continued)

Some ion channel genes of interest for EFS experiments				
Symbol	Full Name	Cytogenetic Location	MIM Number	PubMed ID
KCNE1L	potassium voltage-gated channel, Isk-related family, member 1-like	Xq22.3	300328	10493825
KCNE2	potassium voltage-gated channel, Isk-related family, member 2	21q22.1	603796	10219239
KCNE3	potassium voltage-gated channel, Isk-related family, member 3	reserved	604433	10219239
KCNE4	potassium voltage-gated channel, Isk-related family, member 4	reserved		10219239
KCNF1	potassium voltage-gated channel, subfamily F, member 1	2p25	603787	9434767
KCNF2	literature alias, see KCNG2	-		
KCNF	symbol withdrawn, see KCNF1	-		
KCNG1	potassium voltage-gated channel, subfamily G, member 1	20q13	603788	9434767
KCNG2	potassium voltage-gated channel, subfamily G, member 2	18q22-18q23	605696	10551266
KCNG	symbol withdrawn, see KCNG1	-		
KCNH1	potassium voltage-gated channel, subfamily H (eag-related), member 1	1q32-41	603305	9738473
KCNH2	potassium voltage-gated channel, subfamily H (eag-related), member 2	7q35-q36	152427	7842012
KCNH3	potassium voltage-gated channel, subfamily H (eag-related), member 3	12q13	604527	10455180
KCNH4	potassium voltage-gated channel, subfamily H (eag-related), member 4	reserved	604528	10455180
KCNH5	potassium voltage-gated channel, subfamily H (eag-related), member 5	14	605716	9738473
KCNIP1	Kv channel interacting protein 1	reserved		10676964
KCNIP2	Kv channel-interacting protein 2	10		10676964

TABLE 1 (Continued)

Some ion channel genes of interest for EFS experiments				
Symbol	Full Name	Cytogenetic Location	MIM Number	PubMed ID
KCNIP3	literature alias, see CSEN	-		
KCNJ1	potassium inwardly-rectifying channel, subfamily J, member 1	11q24	600359	7680431
KCNJ2	potassium inwardly-rectifying channel, subfamily J, member 2	17q23.1-q24.2	600681	7696590
KCNJ3	potassium inwardly-rectifying channel, subfamily J, member 3	2q24.1	601534	8088798
KCNJ4	potassium inwardly-rectifying channel, subfamily J, member 4	22q13.1	600504	8016146
KCNJ5	potassium inwardly-rectifying channel, subfamily J, member 5	11q24	600734	
KCNJ6	potassium inwardly-rectifying channel, subfamily J, member 6	21q22.1	600877	7796919
KCNJ7	symbol withdrawn, see KCNJ6	-		
KCNJ8	potassium inwardly-rectifying channel, subfamily J, member 8	12p11.23	600935	8595887
KCNJ9	potassium inwardly-rectifying channel, subfamily J, member 9	1q21-1q23	600932	8575783
KCNJ10	potassium inwardly-rectifying channel, subfamily J, member 10	1q	602208	9367690
KCNJ11	potassium inwardly-rectifying channel, subfamily J, member 11	11p15.1	600937	7502040
KCNJ12	potassium inwardly-rectifying channel, subfamily J, member 12	17p11.1	602323	7859381
KCNJ13	potassium inwardly-rectifying channel, subfamily J, member 13	2q37	603208	9878260
KCNJ14	potassium inwardly-rectifying channel, subfamily J, member 14	19q13	603953	9592090

TABLE 1 (Continued)

Some ion channel genes of interest for EFS experiments				
Symbol	Full Name	Cytogenetic Location	MIM Number	PubMed ID
KCNJ15	potassium inwardly-rectifying channel, subfamily J, member 15	21q22.2	602106	9299242
KCNJ16	potassium inwardly-rectifying channel, subfamily J, member 16	17q23.1-q24.2	605722	11240146
KCNJN1	channel, subfamily J, inhibitor 1	17p11.2-p11.1	602604	8647284
KCNK1	potassium channel, subfamily K, member 1 (TWIK-1)	1q42-q43	601745	8661042
KCNK2	potassium channel, subfamily K, member 2 (TREK-1)	1q41	603219	9721223
KCNK3	potassium channel, subfamily K, member 3 (TASK-1)	2p23	603220	9312005
KCNK4	potassium inwardly-rectifying channel, subfamily K, member 4	11q13	605720	10767409
KCNK5	potassium channel, subfamily K, member 5 (TASK-2)	6p21	603493	9812978
KCNK6	potassium channel, subfamily K, member 6 (TWIK-2)	19q13.1	603939	10075682
KCNK7	potassium channel, subfamily K, member 7	11q13	603940	10206991
KCNK9	potassium channel, subfamily K, member 9 (TASK-3)	8	605874	10734076
KCNK10	potassium channel, subfamily K, member 10	reserved	605873	
KCNK12	potassium channel, subfamily K, member 12	2p22-2p21		
KCNK13	potassium channel, subfamily K, member 13	14q24.1-14q24.3		11060316

TABLE 1 (Continued)

Some ion channel genes of interest for EFS experiments				
Symbol	Full Name	Cytogenetic Location	MIM Number	PubMed ID
KCNK14	potassium channel, subfamily K, member 14	2p22-2p21		11060316
KCNK15	potassium channel, subfamily K, member 15	reserved		
KCNMA1	potassium large conductance calcium-activated channel, subfamily M, alpha member 1	10	600150	7987297
KCNMB1	potassium large conductance calcium-activated channel, subfamily M, beta member 1	5q34	603951	8799178
KCNMB2	symbol withdrawn, see KCNMB3	-		
KCNMB2	potassium large conductance calcium-activated channel, subfamily M, beta member 2	reserved	605214	10097176
KCNMB2L	symbol withdrawn, see KCNMB3L	-		
KCNMB3	potassium large conductance calcium-activated channel, subfamily M beta member 3	3q26.3-q27	605222	10585773
KCNMB3L	potassium large conductance calcium-activated channel, subfamily M, beta member 3-like	22q11		10585773
KCNMB4	potassium large conductance calcium-activated channel, subfamily M, beta member 4	reserved	605223	
KCNMBL	symbol withdrawn, see KCNMB3	-		
KCNMBLP	symbol withdrawn, see KCNMB3L	-		
KCNN1	potassium intermediate/small conductance calcium-activated channel, subfamily N, member 1	19p13.1	602982	8781233

TABLE 1 (Continued)

Some ion channel genes of interest for EFS experiments				
Symbol	Full Name	Cytogenetic Location	MIM Number	PubMed ID
KCNN2	potassium intermediate/small conductance calcium-activated channel, subfamily N, member 2	reserved	605879	
KCNN3	potassium intermediate/small conductance calcium-activated channel, subfamily N, member 3	22q11-q13.1	602983	9491810
KCNN4	potassium intermediate/small conductance calcium-activated channel, subfamily N, member 4	19q13.2	602754	9380751
KCNQ1	potassium voltage-gated channel, KQT-like subfamily, member 1	11p15.5	192500	8528244
KCNQ1OT1	KCNQ1 overlapping transcript 1	11p15.5	604115	10220444
KCNQ2	potassium voltage-gated channel, KQT-like subfamily, member 2	20q13.3-2 20q13.3	121200	9425895
KCNQ3	potassium voltage-gated channel, KQT-like subfamily, member 3	8q24	121201	9425900
KCNQ4	potassium voltage-gated channel, KQT-like subfamily, member 4	1p34	603537	10025409
KCNQ5	potassium voltage-gated channel, KQT-like subfamily, member 5	6q14		10787416
KCNS1	potassium voltage-gated channel, delayed-rectifier, subfamily S, member 1	reserved	602905	9305895
KCNS2	potassium voltage-gated channel, delayed-rectifier, subfamily S, member 2	8q22	602906	9305895
KCNS3	potassium voltage-gated channel, delayed-rectifier, subfamily S, member 3	reserved	603888	10484328

PCR reactions can be carried out with a variety of thermostable enzymes including but not limited to AmpliTaq, AmpliTaq Gold, or Vent polymerase. For AmpliTaq, reactions can be carried out in 10 mM Tris-Cl, pH 8.3, 2.0 mM $MgCl_2$, 200 μM of each dNTP, 50 mM KCl, 0.2 μM of each primer, 10 ng of DNA template, 0.05 units/μl of AmpliTaq. The reactions are heated at 95°C for 3 minutes and then cycled 35 times using suitable cycling parameters, including, but not limited to, 95°C, 20 seconds, 62°C, 20 seconds, 72°C, 3 minutes. In addition to these conditions, a variety of suitable PCR protocols can be found in PCR Primer, A Laboratory Manual, edited by C.W. Dieffenbach and G.S. Dveksler, 1995, Cold Spring Harbor Laboratory Press; or PCR Protocols: A Guide to Methods and Applications, Michael et al., eds., 1990, Academic Press.

It is desirable to sequence the DNA encoding voltage-gated ion channels obtained by the herein-described methods, in order to verify that the desired voltage-gated ion channel has in fact been obtained and that no unexpected changes have been introduced into its sequence by the PCR reactions. The DNA can be cloned into suitable cloning vectors or expression vectors, *e.g.,* the mammalian expression vector pcDNA3.1 (Invitrogen, San Diego, CA) or other expression vectors known in the art or described herein.

A variety of expression vectors can be used to recombinantly express DNA encoding voltage-gated ion channels for use in the present invention. Commercially available expression vectors which are suitable include, but are not limited to, pMC1neo (Stratagene), pSG5 (Stratagene), pcDNAI and pcDNAIamp, pcDNA3, pcDNA3.1, pCR3.1 (Invitrogen, San Diego, CA), EBO-pSV2-neo (ATCC 37593), pBPV-1(8-2) (ATCC 37110), pdBPV-MMTneo(342-12) (ATCC 37224), pRSVgpt (ATCC 37199), pRSVneo (ATCC 37198), pCI.neo (Promega), pTRE (Clontech, Palo Alto, CA), pV1Jneo, pIRESneo (Clontech, Palo Alto, CA), pCEP4 (Invitrogen, San Diego, CA), pSC11, and pSV2-dhfr (ATCC 37146). The choice of vector will depend upon cell type in which it is desired to express the voltage-gated ion channels, as well as on the level of expression desired, and the like.

The expression vectors can be used to transiently express or stably express the voltage-gated ion channels. The transient expression or stable expression of transfected DNA is well known in the art. See, *e.g.*, Ausubel et al., 1995, "Introduction of DNA into mammalian cells," in Current Protocols in Molecular Biology, sections 9.5.1-9.5.6 (John Wiley & Sons, Inc.).

As an alternative to the above-described PCR methods, cDNA clones encoding ion channels can be isolated from cDNA libraries using as a probe oligonucleotides specific for the desired voltage-gated ion channels and methods well known in the art for screening cDNA libraries with oligonucleotide probes. Such methods are described in, *e.g.*, Sambrook *et al.*, 1989, *Molecular Cloning: A Laboratory Manual*; Cold Spring Harbor Laboratory, Cold Spring Harbor, New York; Glover, D.M. (ed.), 1985, *DNA Cloning: A Practical Approach*, MRL Press, Ltd., Oxford, U.K., Vol. I, II. Oligonucleotides that are specific for particular voltage-gated ion channels and that can be used to screen cDNA libraries can be readily designed based upon the known DNA sequences of the voltage-gated ion channels and can be synthesized by methods well-known in the art.

The present invention also provides apparatuses for use with the methods disclosed herein. For example, the present invention provides a multiwell tissue culture plate where a plurality of the wells of the plate contain a pair of electrodes disposed such that when a preselected voltage is applied across the electrodes the transmembrane potential of cells within the wells is altered.

In certain embodiments, the multiwell tissue culture plate contains one of the pair of electrodes on the bottom of the wells and the other of the pair of electrodes on the side of the wells. This embodiment is depicted in Figure 2B.

In other embodiments, the multiwell tissue culture plate contains both of the pair of electrodes on the bottom of the wells. This embodiment is depicted in Figure 2C.

In other embodiments of the multiwell tissue culture plate, one of the pair of electrodes is a layer of conductive material that forms the bottom of the wells and the other of the pair of electrodes enters the wells from above. This embodiment is depicted in Figures 7, 12, and 16.

In other embodiments of the multiwell tissue culture plate, both of the pair of electrodes are embedded in an insulator and enter the wells from above. This embodiment is depicted in Figures 9 and 10.

In other embodiments of the multiwell tissue culture plate, the electrode that enters the wells from above has a central conductive material portion that is surrounded by an insulator. This embodiment is depicted in Figure 8.

In other embodiments of the multiwell tissue culture plate, one of the pair of electrodes forms the bottom of the wells and the other of the pair of electrodes enters the wells from above. This embodiment is depicted in Figures 7 and 10.

In other embodiments of the multiwell tissue culture plate, the pairs of electrodes form an alternating pattern of positive and negative electrodes in the wells. This embodiment is depicted in Figure 16.

In other embodiments of the multiwell tissue culture plate, the layer of conductive material that forms the bottom of the wells is a layer of indium tin oxide that overlays a glass substrate. Preferably, the layer of conductive material and the glass substrate are transparent.

In other embodiments of the multiwell tissue culture plate, a plurality of the wells of the plate contain interdigitating electrodes. This embodiment is depicted in Figures 3 and 5.

The present invention provides a multiwell tissue culture plate where:

the bottom of the wells is a filter membrane upon which cells can be grown;

the wells are located in a trough that can contain fluid;

the trough contains a first electrode;

a second electrode enters the wells from above;

where the first and second electrodes are so disposed that when a preselected voltage is applied across the electrodes the transmembrane potential of cells within the wells is altered. This embodiment is depicted in Figure 8.

The present invention also provides a combination of the multiwell tissue culture plates disclosed herein and a fluorescent imager where the multiwell tissue culture plate and the fluorescent imager are positioned relative to one another such that the fluorescent imager can obtain fluorescent readings from the wells of the multiwell tissue culture plate.

The present invention also provides a combination of a top substrate and a bottom substrate where the top and bottom substrates each contain:

a plurality of virtual wells; and

a layer of conductive material that forms the bottoms of the virtual wells; where the layers of conductive material in the top and bottom substrates are connected to a pulse generator such that the layers of conductive material function as electrodes such that when a preselected voltage is applied across the electrodes the

transmembrane potential of cells within the virtual wells is altered. Such a combination is depicted in Figures 6 and 13.

The present invention also provides a substrate having square or rectangular wells formed by a plurality of generally parallel positive and negative electrodes and a plurality of spacers arranged generally at right angles to the electrodes, where:

one wall of the wells is formed by a positive electrode and the opposite wall of the well is formed by a negative electrode;

the spacers form the walls of the wells that are at right angles to the walls formed by the electrodes;

where the electrodes are so disposed that when a preselected voltage is applied across the electrodes the transmembrane potential of cells within the wells is altered. Such a substrate is depicted in Figure 1.

An example of another embodiment of the present invention comprises:

a substrate having an upper surface upon which are present at least 10^3 living eukaryotic cells which have a voltage-gated ion channel of interest in their plasma membranes;

a plurality of positive electrodes and a plurality of negative electrodes positioned either on or near the substrate such that when a voltage is applied through the positive and negative electrodes the transmembrane potential of the cells is altered;

at least one substance that is suspected of being an activator or an inhibitor of the voltage-gated ion channel;

where the cells contain a fluorescent indicator compound.

An example of another embodiment of the present invention comprises:

a multiwell tissue culture plate having a plurality of wells in which are present at least 10^3 living eukaryotic cells per well of the plurality which cells have a voltage-gated ion channel of interest in their plasma membranes;

a plurality of positive electrodes and a plurality of negative electrodes positioned such that when a preselected voltage is applied through the positive and negative electrodes, the transmembrane potential of the cells is altered;

at least one substance that is suspected of being an activator or an inhibitor of the voltage-gated ion channel in at least one of the plurality of the wells;

where the cells contain a fluorescent indicator compound or a voltage sensitive membrane dye.

The following non-limiting examples are presented to better illustrate the invention.

Example 1

In Figure 24, a preferred system for conducting high throughput screening using EFS stimulation is shown. The system consist of a computer 2402 that comprises an arbitrary waveform generator card 2404 electronically associated with the computer 2402. Custom software was written on the computer 2402 which causes the arbitrary generator card 2404 to generate a pulse voltage waveform (2406) of the appropriate electrical stimulus. The voltage waveform (2406) is applied to the input of eight constant current amplifiers 2408. Each constant current amplifier 2408 services a row on the 96-well sample filter plate 2410. The outputs from the amplifiers 2412 pass through the contacts of electrical relays 2414 allowing the current pulse to be applied to the electrodes 2416.

The waveform generator card 2404 also generates a 7-bit binary transistor-transistor logic TTL value (2418) that represents the address of the well to be excited by the stimulus. In addition, a trigger pulse 2420 is generated. Microprocessor controller 2422, waits for the trigger pulse 2420, interprets the binary value (2418) and then switches on the appropriate relay 2414 which then directs the constant current pulse (2424) to the particular electrode 2416 or electrodes, via electrode connecting wire(s) 2417 in the sample well 2426. Current flows from the amplifier's output (2424), through the relay contact 2414 through the electrode 2416 the liquid in the well 2428, through the well's membrane 2430 and returns via fluid 2432 beneath the membrane 2430 and a return wire 2434. One large common current return trough 2436 services

all 96-electrodes. Other arrangements are possible where each sample well has its own isolated current return trough and return wire. See Example 2 below.

The current return trough 2436 beneath the membranes 2430 has a clear glass bottom 2438 that permits excitation light (2440) from a light source 2442 to pass through the glass bottom 2438, through the transparent membrane 2430 and illuminate cells 2444 adhered to the membrane 2430. Fluorescent light (2446) from the cells 2444 returns back through the membrane 2430 and the glass bottom 2438 entering into the detector 2448. Suitable detectors include those described *supra*. The preferred detector is the FLIPR (Molecular Devices) fluorescence image~~r~~ of the VIPR (Aurora Biosciences)

When the pulse sequence is completed, the microprocessor controller 2422 switches off the relays 2414 isolating the constant current amplifiers' pulses (2424) from the electrodes 2416.

Turning to Figures 25 and 26, Figure 25 represents a photograph of an electrode head 2500 embodiment comprising top electrodes 2516 and first electrode connecting wires 2517. The electrode head comprises a ground contact rod 2510. Figure 26 represents a photograph of a trough embodiment 2600 for use in conjunction with the electrode head 2500 embodiment shown in Figure 25. The trough 2600 comprises bracing posts 2610 to assist in aligning and attachment of the electrode head through apertures 2520 in the electrode head 2500 (see Figure 28). A bottom electrode wire (hidden) is positioned in the trough which when submerged in the salt/buffer solution, upon assembly of the EFS system (see Figure 28) acts as bottom electrode for each of the wells. The bottom electrode wire is in electrical communication with a return connection wire 2620 at position 2630. The return connection wire is secured to the ground contact rod 2510 upon assembly of the EFS system. The trough 2600 also comprises a transparent bottom portion 2640 preferably made of glass.

Figure 27 represents a photograph of the trough embodiment 2600 wherein a Multiscreen^{TM}-Black CM 96 wellplate 2700, with 96 wells 2710, is positioned in the

trough 2600. Information concerning Millipore's multiscreen plates and biopore membranes is found, e.g., at http://www.millipore.com/catalogue.nsf/docs/C7781 and http://www.millipore.com/publications.nsf/docs/tn062.

Figure 28 is a photograph of the assembled EFS system 2800 comprising the trough 2600 with well plate 2700 in place. The electrode head 2500 is secured to the top of the trough 2600 such that the electrodes 2416 are inserted into the wells 2710, one electrode per well. The electrode head 2500 is secured down onto bracing posts 2610 (hidden) by fasteners 2810. The fasteners are preferable threaded nuts. Preferably, prior to assembly, each well 2710 (hidden) has been loaded with cells which have been cultured to canvas the bottom of the wells 2710 (hidden). After cells have been cultured under standard and known conditions, and before assembly of the EFS unit 2800, each well is preferably washed to remove cell media and then loaded with the predetermined buffer solution as discussed above.

Figure 29 shows a graphical representation of data obtained from an embodiment of the invention similar to that depicted in Figure 28. The data represent a membrane potential change in HEK293 cells that have been transfected to express human PN1 voltage-gated sodium channel. Each plot represents a row (12wells) A-H of a 96-well plate. Each column of the 96-well plate data was acquired for 15 seconds on a VIPR™. Stimulation pulse protocol was applied during the data acquisition as follows; 2s baseline was followed with a 2ms square pulse, Amplitude = 20mA, Frequency = 10 Hz, Duration = 5s. Those skilled in the art will readily appreciate, in view of the teachings herein, that the subject system may generate a pulse between 1μs to 1s. Preferably, the pulse generated is between about 0.1ms and about 100ms.

Figure 30 is a bar graph representation of the peak ratio change of data depicted in Figure 29. 1 μM TTX a specific and potent blocker of tetrodotoxin (TTX)-sensitive voltage-gated sodium channels is present in wells E1, F1, G1, H1, A12, B12,

C12 and D12. In addition well A11 contains an internal standard for blocking TTX-sensitive voltage-gated sodium channels. Z-score is a measure of the difference in the uninhibited and inhibited signal divided by the sum of the standard deviations.

Figure 31 shows the effects of increasing concentrations of TTX (upper panel) and of Compound A (lower panel) on the EFS-stimulated depolarization signal in HEK293/PN1 cells. The IC_{50}s obtained in these experiments are comparable to those obtained through other techniques. The high Hill coefficients, nH, result from the threshold nature of the stimulation protocol.

Example 2

Figure 32 represents a photograph of an EFS embodiment 3200 pertaining to an alternative EFS system configuration. The electrode head 2500 is similar to that described above in Figure 25. However, the configurations of the electrodes, wells and trough are configured differently to further isolate the electrical fields. This reduces cross-talk and interference between wells. For this embodiment, the inventors have adapted Millipore's Multiscreen™ Caco-2 Assay System for use as a EFS system. Information concerning the Multiscreen™ Caco-2 Assay System can be found at http://www.millipore.com/publications.nsf/docs/PF1780EN00. The standard commercially available Caco-2 plate system comprises two plates: a membrane-bottom cell growth plate and a 96-well receiver tray. One of the unique characteristics of the Caco-2 system is that it each well has an individual corresponding trough that is accessed basolaterally to each well. Therefore, it supplants the need for a common trough into which all of the wells sit. According to this embodiment, the top electrodes 2516 are disposed into each of the wells in the membrane-bottom cell growth plate (hidden). To establish the bottom electrode for each well, a conductive electrode plate 3220 is provided. The conductive electrode plate 3220 comprise a series of well apertures 3230, providing access of the top electrodes 2410 into the individual wells during assembly. The conductive electrode

plate 3220 also comprises a series of conductive pins (hidden) secured thereto and extending downward at positions 3240. These conductive pins are inserted through the basolateral access port of the membrane-bottom cell growth plate (not shown).

Figure 33 is a depiction of the bottom of the conductive electrode plate 3220 and shows the conductive pins 3310 , which are extending out of the page toward the reader. Figure 34 shows a side-view of the conductive electrode plate 3220 properly positioned atop of the membrane-bottom cell growth plate 3410 and 96 well receiver tray 3420. When the electrode conductive plate 3220 is properly positioned on top of the membrane-bottom cell growth plate 3410, the conductive pins 3310 are inserted through the basolateral access port (not shown) into the individual trough area (not shown) of the 96 well receiver tray. When the individual trough area is filled with the appropriate solution it contacts the bottom of each well and individual pin. Therefore, when the well and trough area are filled with solution, current may flow from the top electrode to the bottom electrode during operation. Figure 35 is a side-view of the assembled EFS system. The assembled system comprises the membrane-bottom cell growth plate 3410 positioned in the 96 well tray 3420. The electrode plate 3220 is mounted on top of the membrane bottom well plate 3410. The electrode head 2500 is shown mounted on top of the electrode plate 3220.

One clear advantage to the EFS systems described in Examples 1 and 2 above, and elsewhere in the present application, is the ability to generate a uniform field across the cells, as opposed to tangential to the cells. Generating an electrical field across the cells is made possible by the novel "top to bottom" placement of the electrodes in a multiwell format.

Example 3

Figure 36 shows a novel electrode embodiment 3600. Figure 36A depicts an expanded view of the electrode 3600. The electrode 3600 comprises two parallel

plates 3610 and 3630 with a low dielectric plate or disc 3620 between them. Optionally, the electrode may be coated with an insulating material. Potential advantages of this design are that special multiwell plates are not required, i.e., any plate that the cells will stick to and that the stimulation and emission light will pass through may be used. There is no filter in the well that may absorb compound or pass compound during long incubations. In the case of the coated electrode, very little current is used and ohmic heating is diminished, even for dc current and even for extended periods of stimulation. The capacitance current is low enough that this advantage applies to ac current as well. The sealed electrode permits placement very close to the cell layer for more uniform stimulation.

Not to be bound by any theory, it is believed that the more uniform the electrical field presented to the cells is, a more accurate indication of potential modulation to the cells will be achieved. In other words, the more uniform the electrical field is, the potential modulation as observed by any of the methods presented herein, e.g., fluorescence, will more directly correlate to actual modulation of ion channels in the cell membrane, and less correlate with background noise in the system caused by cross-interference, cross-illumination, dye effects, dye leaching or any other interference in the system. One way to increase the uniformity of the electrical field applied to the cells is to present one or more of the electrodes in close proximity to, or in contact with, the cells. However, this can affect the cells in deleterious ways leading to failure in the system. Some of the problems associated with close proximity or contact of the electrode(s) to the cells are caused by, for example, ohmic heating, oxidation and formation of bubbles on the electrode. The embodiments of the present invention as taught in Figures 8, 11, 24-28 and 32-35 are particularly preferred because they achieve a uniform electrical field across the cells without putting the electrodes in contact with or close proximity to the cells. Furthermore, the novel electrode design shown in Figure 36 achieves a uniform electrical field, by allowing close proximity of the electrode to the cells, without creating the problems of ohmic heating, oxidation, or bubbling of the cells.

It is believed that the subject EFS system embodiments produce substantially uniform fields, where the one or more electrical fields vary over an area of observation by no more than about 30% from the mean electrical field at any one time. Percentages are determined by measurements in two dimensions; or preferably, variation is calculated in three dimensions. In a more preferred embodiment, the one or more electrical fields vary over an area of observation by no more than about 15 % from the mean electrical field at any one time. In an even more preferred embodiment, the one or more electrical fields vary over an area of observation by no more than 10 % from the mean electrical field at any one time. In an optimal embodiment, the variation is no more than 5 % from the mean.

The similarity to a capacitor is obvious, but the low dielectric 3620 between the plates 3610 and 3630 reduces the amount of current required to initially charge the plates with only a miniscule current required to maintain the charge between the plates. An external electric field is generated that can be used to depolarize the cells. The external electric field density is reduced by a high dielectric between the plates as is used with an authentic capacitor and is maximal with a low dielectric such as teflon or mylar or no dielectric. The external field density is further enhanced by placing the plates very close together, but the optimal separation may be determined empirically.

Figure 36B shows an embodiment comprising a concurrent lead design. The concurrent lead comprises an internal wire 3655 and an external wire 3650. The internal wire passes through the top plate 3610 and dielectric plate 3620 and is attached or integral to the bottom plate 3630. The external wire is attached or integral to the top plate 3610. Those skilled in the art will recognize that the foregoing arrangement of the leads may be reversed. Figure 36C shows an embodiment comprising edge leads 3660 and 3665. Edge lead 3660 is attached or integral to top plate 3610 and edge lead 3665 is attached or integral to bottom plate 3630.

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Some of the embodiments of the subject invention include the following:

A method of characterizing the biological activity of a candidate compound comprising.

exposing one or more cells to said compound; repetitively exposing said one or more cells to one or more electric fíe1ds so as to effect a controlled change in transmembrane potential of said one or more cells; and monitoring, without using a patch clamp, changes in the transmembrane potential of said one or more cells.

The above method, where the monitoring comprises detecting fluorescence emission from an area of observation containing said one or more cells.

The above method, where the electric fíe1ds are biphasic.

The above method, additionally comprising limiting spatial variation in electric fíeld intensity so as to minimize irreversible cell electroporation.

The above method, where one or more electrical fíe1ds may cause an ion channel of interest to cycle between different voltage dependent states.

The above method, where the one or more electrical fíe1ds cause an ion channel of interest to open.

The above method, where the one or more electrical fields cause an ion channel of interest to be released from inactivation.

The above method, where the one or more cells comprise a voltage sensor selected from the group consisting of a FRET based voltage sensor, an electrochromic transmembrane potential dye, a transmembrane potential redistribution dye, an ion sensitive fluorescent or luminescent molecule and a radioactive ion.

The above method, where the one or more cells comprise a voltage regulated ion channel.

The above method, where the voltage regulated ion channel is selected from the group consisting of a potassium channel, a calcium channel, a chloride channel and a sodium channel.

The above method, where the electric field exhibits limited spatial variation in intensity in the area of observation of less than about 25% from. a mean intensity in that area.

The above method, where the one or more electrical fields varies over an area of observation by no more than about 15 % from the mean electrical field at any one time.

The above method, where the one or more electrical fields varies over an area of observation by no more than about 5 % from the mean electrical field at any one time.

The above method, where the one or more electrical fields comprises stimulation with either a square wave-form, a sinusoidal wave-form or a saw tooth wave-form.

The above method, where the one or more electrical fields have an amplitude within the range of about 1 0 V/cm to about 1 00 V/cm.

The above method, where the one or more electrical fields have an amplitude within the range of about 20 V/cm to about 80 V/cm.

The above method, where the one or more electrical fields are repeated at a frequency of stimulation that is greater than or equal to the reciprocal of the transmembrane time constant of said one or more cells.

The above method, where the one or more electrical fields are repeated at a frequency of stimulation within the range of zero to 1 kHz.

The above method, where the one or more electrical fields have a pulse duration within the range of about 100 microseconds to about 20 milliseconds.

The above method, where the transmembrane potential is developed across the plasma membrane of said one or more cells.

A method of assaying the biochemical activity of a compound against a target ion channel comprising.

selecting a cell line having a normal resting transmembrane potential corresponding to a selected voltage dependent state of said target ion channel; expressing said target

ion channel in a population of cells of said selected cell line; exposing said population of cells to said compound; repetitively exposing said population of cells to one or more electric ffe1ds so as to effect a controlled change in transmembrane potential of said one or more cells; and monitoring changes in the transmembrane potential of said one or more cells.

The above method, where the target ion channel is exogenously expressed in said cell line.

The above method, where the cell line is transfected with nucleic acid encoding said target ion channel.

The above method, where the cell line expresses insignificant levels of other ion channels.

The above method, where the cell line is selected from the group consisting of CUL,LTK(-), and CHO-M.

The above method, where the target ion channel is a sodium channel, and wherein said population of cells is selected from the group consisting of CHL cells, LTK(-) cells, and CHO-K1 cells.

The above method, where the target ion channel is a sodium channel, and wherein said population of cells is selected from the group consisting of HEK-293 cells, RBL cells, F11 cells, and HL5 cells.

The above method, where the target ion channel is a potassium channel, and wherein said population of cells is selected from the group consisting of CHL cells, LTK(-) cells, and CHO-K1cells.

The above method, where the target ion channel is a calcium channel, and wherein said population of cells is selected from the group consisting of CHL cells, LTK(-) cells, and CHO-K1 cells.

A method of assaying ion channel activity comprising.

exposing at least one cell to a plurality of electric field pulses so as to create a controlled change in transmembrane potential and so as to activate an ion channel of interest; and detecting ion channel activity by detecting one or more changes in transmembrane potential without using a patch clamp.

The above method, where the at least one cell comprises a voltage sensor selected from the group consisting of a FRET based voltage sensor, an electrochromic transmembrane potential dye, a transmembrane potential redistribution dye, an ion sensitive fluorescent or luminescent molecule and a radioactive ion.

The above method, where the voltage sensor comprises a FRET based voltage sensor.

The above method, where the ion channel of interest is a voltage regulated ion channel.

The above method, where the plurality of electric field pulses cause said ion channel of interest to cycle between different voltage dependent states.

The above method, where the at least one cell is an eukaryotic cell.

The above method, where the at least one cell is a non-excitable cell.

The above method, where the at least one cell is a prokaryotic cell.

The above method, where the at least one cell is a tissue culture cell.

The above method, where the at least one cell is a primary cell line.

The above method, where the at least one cell is part of an intact living organism.

A method of assaying ion channel activity comprising.

expressing a selected target ion channel in at least one cell; expressing a selected counter ion channel in said at least one cell; exposing said at least one cell to a plurality of electric field pulses so as to create a controlled change in transmembrane potential and so as to activate said counter ion channel; and monitoring the transmembrane potential of said at least one cell.

The above method, where a transmembrane potential change is detected when said ion channel of interest is blocked.

The above method, where the ion channel of interest comprises a ligand gated ion channel.

The above method, where the counter channel comprises a sodium channel.

A method of modifying the transmembrane potential of a cell comprising repetitively applying biphasic electric field pulses to said cell, wherein said pulses have a maximum amplitude of less than approximately 90 V/cm, wherein said pulses

are applied at a rate of at least about 1 per second, and wherein the total duration of each pulse is at least about 1 millisecond.

The above method, where the maximum amplitude is approximately 20 to 40 V/cm.

The above method, where the pulse duration is approximately 2 to 10 milliseconds per phase.

The above method, where the pulses are applied at a rate of approximately 20 to 100 pulses per second.

A method of characterizing the biological activity of a candidate compound comprising.

placing one or more cells into an area of observation in a sample well; exposing said one or more cells to said compound; repetitively exposing said one or more cells to a series of biphasic electric fields at a rate of approximately 20 to 100 pulses per second, wherein said electric fíe1ds exhibit limited spatial variation in intensity in the area of observation of less than about 25% from a mean intensity in that area, and wherein said electric fields produce a controlled change in transmembrane potential of said one or more cells; and monitoring changes in the transmembrane potential of said one or more cells by detecting fluorescence emission of a FRET based voltage sensor from, an area of observation containing said one or more cells.

The above method, where the one or more electrical fields cause an ion channel of interest to open.

The above method, where the one or more electrical fields cause an ion channel of interest to be released from inactivation.

The above method, where the one or more cells comprise a voltage regulated ion channel.

The above method, where the voltage regulated ion channel is selected from the group consisting of a potassium channel, a calcium channel, a chloride channel and a sodium channel.

The above method, where the one or more electrical fields likely vary over an area of observation by no more than about 15 % from the mean electrical field at any one time.

The above method, where the one or more electrical fields varies over an area of observation by no more than about 5 % from the mean electrical field at any one time.

The above method, where the one or more electrical fields are selected from a square wave-form, a sinusoidal wave-form or a saw tooth wave-form.

A high throughput screening system comprising.

a plurality of wells having a high transmittance portion through which cells present in said wells are optically observable in an area of observation; two electrodes in each of said plurality of wells; an optical detector configured to detect light emanating from said wells through said high transmittance portion; a power supply connected to said electrodes; wherein said power supply and said electrodes are configured to apply a series of electric fields to cells within said area of observation, said electric fields having a spatial variation of less than about 25% of a mean field intensity within said area of observation, said electric fields being effective to controllably alter the transmembrane potential of a portion of said cells; a data processing unit configured to interpret said light emanating from said wells, through said high transmittance portion as ion channel activity resulting from said transmembrane potential alterations.

The above high throughput screening system, where the pluarality of wells are located in a multiwell plate.

The above high throughput screening system, where the high transmittance portion is made from a material selected from the group consisting of glass, quartz, cycloolefin, Aclar, polypropylene, polyethylene and polystyrene.

The above high throughput screening system, where the high transmittance portion exhibits less fluorescence when excited with UV light in the range of 250 nm to 400 nm than polystyrene.

The above high throughput screening system, where the electrodes are located in a well of said plurality of wells.

The above high throughput screening system, where the electrodes are located in a bottom layer of said plurality of wells.

The above high throughput screening system, where the multiwell plate comprises up to 96 wells.

The above high throughput screening system, where the multiwell plate comprises greater than 96 wells.

The above high throughput screening system, where the multiwell plate comprises greater than 384 wells.

The above high throughput screening system, where the electrodes are made of a material selected from the group consisting of gold, platinum, palladium, chromium, molybdenum, iridium, tungsten, tantalum and titanium.

The above high throughput screening system, where the multiwell plate comprises optically opaque materials or pigments to reduce the transmission of light.

The above high throughput screening system, where the electrodes are separated by a gap within the range of about 1 to 4 mm.

The above high throughput screening system, where the electrodes are separated by a gap within the range of about 0. 1 to 1 mm.

1.0 The above high throughput screening system, where the electrodes are separated by a gap within the range of about 0.01 to 0.1 mm.

The above high throughput screening system, where the electrodes are charged to create an electrical field intensity of between 5 to 100 V/cm across said gap, and wherein the total charge transferred across the surface area of the electrically conductive material, in fluidic connection with the interior of the well is less than or equal to 100μC/mm2.

The above high throughput screening system, where the plurality of wells further comprise an insulator orientated and configured so as to create an area of observation within said well in which. the electrical field intensity varies by no more than 1 0 % from the mean electrical field intensity when said at least two strips of electrically conductive material are charged to create an electrical field intensity of between 5 to 100 V/cm across said gap, and. wherein the total charge transferred across the surface area of the electrically conductive material, in fluidic connection with the interior of the well is less than or equal to looptC/mm2 .

The above high throughput screening system, where the plurality of wells further comprise at least two satellite electrical conductors.

A high throughput screening system comprising.

sample wells; liquid handling stations for adding reagents and/or cells to said sample wells; and means for controlling the transmembrane potential of cells in said sample wells so as to selectively cause ion channel activity.

means for optically monitoring changes in said transmembrane potential.

The above high throughput screening system, where the means comprises electrodes configured to create an electric field having a spatial variation of less than about 25% of a mean field intensity within an area of observation.

The above high throughput screening system, where the means for controlling the transmembrane potential comprise an electrode array assembly.

The above high throughput screening system, where the electrode assembly array comprises 8 electrode assemblies.

The above high throughput screening system, where the electrode assembly array comprises 96 electrode assemblies.

The above -high throughput screening system, where the electrode assembly array comprises greater than 96 electrode assemblies.

The above high throughput screening system, where the system further comprises means for retractably moving said electrode assembly into and out of the wells of a multiwell plate.

The above high throughput screening system, where the means for controlling the transmembrane potential comprises electrical conductors with two substantially parallel planar surfaces.

The above high throughput screening system, where the electrical conductors are separated by a gap within the range of 1 to 4 mm.

The above high throughput screening system, where the electrical conductors are separated5 by a gap within the range of 0. 1 to 1 mm.

The above high throughput: screening system, where the electrical conductors further comprise a first insulator.

The above high throughput screening system, where the first insulator comprises two planar surfaces orientated perpendicular to said substantially parallel planar surfaces of said electrical conductors and substantially parallel with respect to each other.

The above high throughput: screening system, where the electrical conductors further comprise a second insulator attached to said at least two electrical conductors, wherein said second insulator is interposed in said gap between said at least two electrical conductors to define the depth of said aqueous solution between said at least two electrical conductors.

The above high throughput: screening system, where the first insulator is composed of allow fluorescence material, wherein. said low fluorescence material exhibits less fluorescence when excited with UV light in the range 250 nm to 400 nm than polystyrene of comparable size.

The above high throughput screening system, where the second insulator is composed of a low fluorescence material, wherein said low fluorescence material exhibits less fluorescence when excited with UV light in the range 250 nm to 400 nm than polystyrene of comparable size.

The above high throughput screening system, where the first insulator comprises an insulator selected from the group consisting of plastic, glass and ceramic.

The above high throughput screening system, where the plastic is selected from the group consisting of nylon, polystyrene, Teflon (tetrafluoroethylene), polypropylene, polyethylene,poly-vinyl chloride, and cycloolefín.

The above high throughput screening system, where the electrical conductors comprise a conductor selected from the group consisting of gold, platinum, titanium, tungsten, molybdenum, iridium, vandium, Nb, Ta, stainless steel and graphite.

The above high throughput screening system, where the electrical conductors comprise a surface treatment to reduce electrolysis.

The above high throughput screening system, where the surface treatment to reduce electrolysis comprises platinum black, gold black, iridium/iridium oxide, titanium/titanium nitride or polypyrrole films.

The above high throughput screening system, where the electrical field intensity varies by no more than 10 % from the mean electrical field intensity when said at least two electrical conductors are charged to create an electrical field intensity of between 5 to 100 V/cm across said gap, wherein the total charge transferred across the surface area of the electrical conductors in contact with said aqueous solution is less than or equal to 1 00 μC/mm2.

The above high throughput screening system, where the electrical field intensity varies by no more than 5% from the mean electrical field intensity when said at least two electrical conductors are charged to create an electrical field intensity of between 5 to 100 V/cm across said gap, wherein the total charge transferred across the surface area of the electrical conductors in contact with said aqueous solution is less than or equal to 100 μC/mm2.

A method of screening a plurality of drug candidate compounds against a target ion channel comprising.

expressing said target ion channel in a population of host cells; placing a plurality of said host cells into each of a plurality of sample wells; adding a candidate drug compound to at least: one of said plurality of sample wells; and modulating the transmembrane potential of host cells in said plurality of sample wells with a repetitive application of electric fíe1ds so as to set said transmembrane potential to a level corresponding to a pre-selected voltage dependent state of said target ion channel.

The above method, additionally comprising selecting a host: cell line having a normal resting transmembrane potential corresponding to a second pre-selected voltage dependent state of said target ion channel.

The above method, where the electric fíe1ds are biphasic.

The above method, where electric fields cause an ion channel of interest to cycle between different voltage dependent states.

The above method, where the electric fields cause an ion channel of interest to open.

The above method, where the electric fields cause an ion channel of interest to be released from inactivation.

The above method, where the one or more cells comprise a voltage sensor selected from the group consisting of a FRET based voltage sensor, an electrochromic transmembrane potential dye, a transmembrane potential redistribution dye, an ion sensitive fluorescent or luminescent molecule and a radioactive ion.

The above method, where the target ion channel is selected from the group consisting of a potassium channel, a calcium channel, a chloride channel and a sodium channel.

The above method, where the one or more electrical fields comprises stimulation with either a square wave-form, a sinusoidal wave-form or a saw tooth wave-form.

The above method, where the one or more electrical fields have an amplitude within the range of about 10 V/cm to about 100 V/cm.

The above method, where the one or more electrical fíe1ds have an amplitude within the range of about 20 V/cm to, about 80 V/cm.

An assay plate and electrode assembly comprising at least one sample well having electrodes placed therein, wherein said electrodes are positioned with respect to the bottom surface of the well to provide an electric field adjacent to said bottom surface that varies by less than about 10% from a mean field intensity over at least about 20% of the surface area of said bottom surface.

The above assembly, where the electrodes comprise plate electrodes extending down into said well such that bottom ends of said electrodes are adjacent to but not in contact with said bottom surface.

The above assembly, comprising two electrodes per sample well. The above assembly, comprising more than two electrodes per sample well.

The above assembly, where the electrodes are plated onto said bottom surface of said well. The above assembly, where the bottom surface comprises a high optical transmittance portion.

The above assembly, where the high transmittance portion is made from a material selected from the group consisting of glass, quartz, cyc1oolefin, Aclar, polypropylene, polyethylene and polystyrene.

The above assembly, where the high transmittance portion exhibits less fluorescence when excited with UV light in the range of 250 nm to 400 nm than polystyrene.

The above assembly, where the electrodes are located in a wall of said plurality of wells.

The above assembly, where the plate comprises up to 96 wells.

The above assembly, where the plate comprises greater than 96 wells.

The above assembly, where the plate comprises greater than 384 wells.

The above assembly, where the electrodes are made of a material selected from the group consisting of gold, platinum, palladium, chromium, molybdenum, iridium, tungsten, tantalum and titanium.

The above assembly, where the electrodes are separated by a gap within the range of about1 to 4 mm.

The above assembly, where the electrodes are separated by a gap within the range of about 0.1 to 1 mm.

The above assembly, where the electrodes are separated by a gap within the range of about 0.01 to 0.1 mm.

A bottom panel for a multi-well plate comprising.

at least one row of high transmittance regions with positions corresponding to well locations; a first: strip of conductive material extending along said row and overlapping a first portion of said well locations; and a second strip of conductive material extending along said row and overlapping a second portion of said well locations.

The above bottom panel, additionally comprising a first: electrical contact proximate to an end of said first strip and a second electrical contact proximate to an end of said second strip.

An assay apparatus comprising.

a sample well; a first pair of electrodes positioned within said sample well; at least one additional satellite electrode positioned within said sample well.

The above assay apparatus, where the at least one additional satellite electrode comprises second and third pairs of electrodes.

The above assay apparatus, where the satellite electrodes are charged to a potential less than that of the first pair of electrodes.

The above assay apparatus, where the electrodes are positioned with respect to the bottom surface of the well to provide an electric field adjacent to said bottom surface that varies by less than about 10% from a mean field intensity over at least about 20% of the surface area of said bottom surface.

The present invention is not to be limited in scope by the specific embodiments described herein. Indeed, various modifications of the invention in addition to those described herein will become apparent to those skilled in the art from the foregoing description. Such modifications are intended to fall within the scope of the appended claims.

Various publications are cited herein, the disclosures of which are incorporated by reference in their entireties. Furthermore, for general information, PCT Publication No. PCT/US01/21652 is incorporated herein in its entirety to the extent it is accurate and not inconsistent with the teachings herein. All patents, patent applications, publications, texts and references discussed or cited herein are understood to be incorporated by reference to the same extent as if each individual publication or patent application was specifically and individually set forth in its entirety. In addition, all references, patents, applications, and other documents cited in an Invention Disclosure Statement, Examiner's Summary of Cited References, or otherwise entered into the file history of this application are taken to be incorporated by reference into this specification for the benefit of later applications claiming

priority to this application. Finally, all terms not specifically defined are first taken to have the meaning given through usage in this disclosure, and if no such meaning is inferable, their normal meaning.

WHAT IS CLAIMED IS:

1. A method for identifying modulators of the activity of a voltage-gated ion channel comprising:

(a) altering the transmembrane potential of at least a portion of the membrane of a cell expressing the voltage-gated ion channel by applying a voltage to the cells through extracellular electrodes while monitoring ion flow through the voltage-gated ion channel;

(b) exposing the cell in step (a) to a substance and monitoring ion flow through the voltage-gated ion channel;

(c) comparing the ion flow through the voltage-gated ion channel in step (a) and step (b);

where a difference in the ion flow through the voltage-gated ion channel in step (a) and step (b) indicates that the substance is a modulator of the voltage-gated ion channel.

2. A method for identifying modulators of the activity of a voltage-gated ion channel comprising:

(a) dividing a plurality of cells expressing the voltage-gated ion channel into a control portion and a test portion;

(b) altering the transmembrane potential of the control portion of cells by applying a voltage to the cells through extracellular electrodes while monitoring ion flow through the voltage-gated ion channel;

(c) altering the transmembrane potential of the test portion of cells by applying the voltage to the cells through extracellular electrodes in the presence of a substance while monitoring ion flow through the voltage-gated ion channel;

(d) comparing the ion flow through the voltage-gated ion channel in step (b) and step (c);

where a difference in the ion flow through the voltage-gated ion channel in step (b) and step (c) indicates that the substance is modulator of the voltage-gated ion channel.

3. A method of identifying activators of a voltage-gated ion channel comprising:

(a) providing a substrate upon which are living eukaryotic cells that express a plurality of the voltage-gated ion channels in their plasma membranes;

(b) providing positive and negative electrodes positioned either on or near the substrate such that when a preselected voltage is applied through the positive and negative electrodes the transmembrane electrical potential of the cells is altered such that at least a portion of the voltage-gated ion channels are closed;

(c) applying the preselected voltage through the positive and negative electrodes;

(d) determining a control value for the flow of ions through the voltage-gated ion channels of the cells in step (c);

(e) exposing the cells of step (c) to a substance for a period sufficient and under conditions such that a detectable number of the portion of the voltage-gated ion channels that are closed become open and allow ion flow through the detectable number of voltage-gated ion channels if the substance is an activator of the voltage-gated ion channels;

(f) determining a test value for the flow of ions through the voltage-gated ion channels of the cells of step (e);

(g) comparing the control value to the test value;

where if the control value is less than the test value, then the substance is an activator of the voltage-gated ion channel.

4. The method of claim 3 where the substrate is glass or a multiwell tissue culture plate and is not silicon or a field effect transistor.

5. The method of claim 4 where the substrate contains wells in which the cells are present.

6. The method of claim 5 where the number of wells is 12, 24, 96, 384, 1,536, or 3,456.

7. The method of claim 5 where the wells are virtual wells.

8. The method of claim 3 where at least 50,000 substances are tested in a 24 hour period.

9. The method of claim 3 where the voltage-gated ion channel is a voltage-gated sodium channel, a voltage-gated potassium channel, or a voltage-gated calcium channel.

10. The method of claim 9 where the voltage-gated ion channel is a voltage-gated sodium channel.

11. The method of claim 9 where the voltage-gated ion channel is a voltage-gated potassium channel.

12. The method of claim 9 where the voltage-gated ion channel is a voltage-gated calcium channel.

13. The method of claim 3 where the cells are selected from the group consisting of: L cells L-M(TK$^-$) (ATCC CCL 1.3), L cells L-M (ATCC CCL 1.2), HEK293 (ATCC CRL 1573), Raji (ATCC CCL 86), CV-1 (ATCC CCL 70), COS-1 (ATCC CRL 1650), COS-7 (ATCC CRL 1651), CHO-K1 (ATCC CCL 61), 3T3 (ATCC CCL 92), NIH/3T3 (ATCC CRL 1658), HeLa (ATCC CCL 2), C127I (ATCC CRL 1616), BS-C-1 (ATCC CCL 26), MRC-5 (ATCC CCL 171), CPAE (ATCC CCL 209), Saos-2 (ATCC HTB-85), ARPE-19 human retinal pigment epithelium (ATCC CRL-2302), GH3 cells, and primary cardiac myocytes.

14. The method of claim 13 where the cells are HEK293 (ATCC CRL 1573), GH3 cells, or primary cardiac myocytes.

15. The method of claim 3 where the cells contain a fluorescent indicator compound.

16. The method of claim 15 where the fluorescent indicator compound is selected from the group consisting of: fluo-3, fura-2, fluo-4, fluo-5, calcium green-1, Oregon green, 488 BAPTA, SNARF-1, and indo-1.

17. The method of claim 3 where the positive and negative electrodes are interdigitating.

18. The method of claim 3 where the substrate is a multiwell tissue culture plate having a plurality of wells that contain one positive and one negative electrode.

19. The method of claim 3 where the substrate is a multiwell tissue culture plate having a plurality of wells where one of the positive or negative electrodes forms the bottom of the wells and the other of the positive or negative electrode enters the wells from above.

20. The method of claim 3 where the substrate is a multiwell tissue culture plate having a plurality of virtual wells.

21. The method of claim 5 where each well contains from 10^3 to 10^7 cells and the cells contain a fluorescent indicator compound or a fluorescent voltage sensing dye.

22. The method of claim 3 where the cells do not naturally express the voltage-gated ion channel but have been transfected with an expression vector that encodes the voltage-gated ion channel.

23. A method of identifying inhibitors of a voltage-gated ion channel comprising:

(a) providing a substrate upon which are living eukaryotic cells that express a plurality of the voltage-gated ion channels in their plasma membranes;

(b) providing positive and negative electrodes positioned either on or near the substrate such that when a preselected voltage is applied through the positive and negative electrodes the transmembrane electrical potential of the cells is altered such that at least a portion of the voltage-gated ion channels are open;

(c) applying the preselected voltage through the positive and negative electrodes;

(d) determining a control value for the flow of ions through the voltage-gated ion channels of the cells in step (c);

(e) exposing the cells of step (c) to a substance for a period sufficient and under conditions such that a detectable number of the portion of the voltage-gated ion channels that are open become closed and restrict ion flow through the detectable number of voltage-gated ion channels if the substance is an inhibitor of the voltage-gated ion channels;

(f) determining a test value for the flow of ions through the voltage-gated ion channels of the cells of step (e);

(g) comparing the control value to the test value;

where if the control value is greater than the test value, then the substance is an inhibitor of the voltage-gated ion channel.

24. The method of claim 23 where the substrate is glass or a multiwell tissue culture plate and is not silicon or a field effect transistor.

25. The method of claim 24 where the substrate contains wells in which the cells are present.

26. The method of claim 25 where the number of wells is 12, 24, 96, 384, 1,536, or 3,456.

27. The method of claim 26 where the wells are virtual wells.

28. The method of claim 23 where at least 50,000 substances are tested in a 24 hour period.

29. The method of claim 23 where the voltage-gated ion channel is a voltage-gated sodium channel, a voltage-gated potassium channel, or a voltage-gated calcium channel.

30. The method of claim 29 where the voltage-gated ion channel is a voltage-gated sodium channel.

31. The method of claim 29 where the voltage-gated ion channel is a voltage-gated potassium channel.

32. The method of claim 29 where the voltage-gated ion channel is a voltage-gated calcium channel.

33. The method of claim 23 where the cells are selected from the group consisting of: L cells L-M(TK$^-$) (ATCC CCL 1.3), L cells L-M (ATCC CCL 1.2), HEK293 (ATCC CRL 1573), Raji (ATCC CCL 86), CV-1 (ATCC CCL 70), COS-1 (ATCC CRL 1650), COS-7 (ATCC CRL 1651), CHO-K1 (ATCC CCL 61), 3T3 (ATCC CCL 92), NIH/3T3 (ATCC CRL 1658), HeLa (ATCC CCL 2), C127I (ATCC CRL 1616), BS-C-1 (ATCC CCL 26), MRC-5 (ATCC CCL 171), CPAE (ATCC CCL 209), Saos-2 (ATCC HTB-85), ARPE-19 human retinal pigment epithelium (ATCC CRL-2302), GH3 cells, and primary cardiac myocytes.

34. The method of claim 33 where the cells are HEK293 (ATCC CRL 1573), GH3 cells, or primary cardiac myocytes.

35. The method of claim 23 where the cells contain a fluorescent indicator compound.

36. The method of claim 35 where the fluorescent indicator compound is selected from the group consisting of: fluo-3, fura-2, fluo-4, fluo-5, calcium green-1, Oregon green, 488 BAPTA, SNARF-1, and indo-1.

37. The method of claim 23 where the positive and negative electrodes are interdigitating.

38. The method of claim 23 where the substrate is a multiwell tissue culture plate having a plurality of wells that contain one positive and one negative electrode.

39. The method of claim 23 where the substrate is a multiwell tissue culture plate having a plurality of wells where one of the positive or negative

electrodes forms the bottom of the wells and the other of the positive or negative electrodes enters the wells from above.

40. The method of claim 23 where the substrate is a multiwell tissue culture plate having a plurality of virtual wells.

41. The method of claim 25 where each well contains from 10^3 to 10^7 cells and the cells contain a fluorescent indicator compound or a fluorescent voltage sensing dye.

42. The method of claim 23 where the cells do not naturally express the voltage-gated ion channel but have been transfected with an expression vector that encodes the voltage-gated ion channel.

43, A method of identifying activators of a voltage-gated ion channel comprising:

(a) providing a substrate upon which are living eukaryotic cells that express a plurality of the voltage-gated ion channels in their plasma membranes;

(b) providing positive and negative electrodes positioned either on or near the substrate such that when a preselected voltage is applied through the positive and negative electrodes the transmembrane electrical potential of the cells is altered such that at least a portion of the voltage-gated ion channels are closed;

(c) applying the preselected voltage through the positive and negative electrodes to a control sample of the cells;

(d) determining a control value for the flow of ions through the voltage-gated ion channels of the control sample of the cells in step (c);

(e) applying the preselected voltage through the positive and negative electrodes to a test sample of the cells while exposing the test sample of the cells to a substance for a period sufficient and under conditions such that a detectable number of the portion of the voltage-gated ion channels that are closed in the test sample become open and allow ion flow through the detectable number of voltage-gated ion channels if the substance is an activator of the voltage-gated ion channels;

(f) determining a test value for the flow of ions through the voltage-gated ion channels of the test sample of cells of step (e);

(g) comparing the control value to the test value;

where if the control value is less than the test value, then the substance is an activator of the voltage-gated ion channel.

44. A method of identifying inhibitors of a voltage-gated ion channel comprising:

(a) providing a substrate upon which are living eukaryotic cells that express a plurality of the voltage-gated ion channels in their plasma membranes;

(b) providing positive and negative electrodes positioned either on or near the substrate such that when a preselected voltage is applied through the positive and negative electrodes the transmembrane electrical potential of the cells is altered such that at least a portion of the voltage-gated ion channels are open;

(c) applying the preselected voltage through the positive and negative electrodes to a control sample of the cells;

(d) determining a control value for the flow of ions through the voltage-gated ion channels of the control sample of the cells in step (c);

(e) applying the preselected voltage through the positive and negative electrodes to a test sample of the cells while exposing the test sample of the cells to a substance for a period sufficient and under conditions such that a detectable number of the portion of the voltage-gated ion channels that are open in the test sample become closed and restrict ion flow through the detectable number of voltage-gated ion channels if the substance is an inhibitor of the voltage-gated ion channels;

(f) determining a test value for the flow of ions through the voltage-gated ion channels of the test sample of cells of step (e);

(g) comparing the control value to the test value;

where if the control value is greater than the test value, then the substance is an inhibitor of the voltage-gated ion channel.

45. An apparatus for use in identifying activators or inhibitors of voltage-gated ion channels comprising:

a substrate having an upper surface upon which are present at least 10^3 living eukaryotic cells which have a voltage-gated ion channel of interest in their plasma membranes;

a plurality of positive electrodes and a plurality of negative electrodes positioned either on or near the substrate such that when a voltage is applied through the positive and negative electrodes the transmembrane potential of the cells is controlled;

at least one substance that is suspected of being an activator or an inhibitor of the voltage-gated ion channel;

where the cells contain a fluorescent indicator compound.

46. A multiwell tissue culture plate having:

a plurality of wells in which are present at least 10^3 living eukaryotic cells per well of the plurality which cells have a voltage-gated ion channel of interest in their plasma membranes;

a plurality of positive electrodes and a plurality of negative electrodes positioned such that when a preselected voltage is applied through the positive and negative electrodes, the transmembrane potential of the cells is altered;

at least one substance that is suspected of being an activator or an inhibitor of the voltage-gated ion channel in at least one of the plurality of the wells;

where the cells contain a fluorescent indicator compound or a voltage sensitive membrane dye.

47. A multiwell tissue culture plate where a plurality of the wells of the plate contain a pair of electrodes disposed such that when a preselected voltage is applied across the electrodes the transmembrane potential of cells within the wells is altered.

48. The multiwell tissue culture plate of claim 47 where the multiwell tissue culture plate contains one of the pair of electrodes on the bottom of the wells and the other of the pair of electrodes on the side of the wells.

49. The multiwell tissue culture plate of claim 47 where the multiwell tissue culture plate contains both of the pair of electrodes on the bottom of the wells.

50. The multiwell tissue culture plate of claim 47 where one of the pair of electrodes is a layer of conductive material that forms the bottom of the wells and the other of the pair of electrodes enters the wells from above.

51. The multiwell tissue culture plate of claim 47 where both of the pair of electrodes are embedded in an insulator and enter the wells from above.

52. The multiwell tissue culture plate of claim 50 where the electrode that enters the wells from above has a central conductive material portion that is surrounded by an insulator.

53. The multiwell tissue culture plate of claim 47 where the pairs of electrodes form an alternating pattern of positive and negative electrodes in the wells.

54. The multiwell tissue culture plate of claim 50 where the layer of conductive material that forms the bottom of the wells is a layer of indium tin oxide that overlays a glass substrate.

55. The multiwell tissue culture plate of claim 54 where the layer of conductive material and the glass substrate are transparent.

56. The multiwell tissue culture plate of claim 47 where a plurality of the wells of the plate contain interdigitating electrodes.

57. A multiwell tissue culture plate where:

the bottom of the wells is a filter membrane upon which cells can be grown;

the wells are located in a trough suitable for containing a fluid;

the trough contains a first electrode;

a second electrode enters the wells from above;

where the first and second electrodes are so disposed that when a preselected voltage is applied across the electrodes the transmembrane potential of cells within the wells is altered.

58. A combination of the multiwell tissue culture plate, as according to claims 46 to 57, and a fluorescence imager where the multiwell tissue culture plate and the fluorescence imager are positioned relative to one another such that the fluorescence imager can obtain fluorescence readings from the wells of the multiwell tissue culture plate.

59. A combination of a top substrate and a bottom substrate where the top and bottom substrates each contain:
a plurality of virtual wells; and
a layer of conductive material that forms the bottoms of the virtual wells;
where the layers of conductive material in the top and bottom substrates are connected to a pulse generator such that the layers of conductive material function as electrodes such that when a preselected voltage is applied across the electrodes the transmembrane potential of cells within the virtual wells is altered.

60. A substrate having square or rectangular wells formed by a plurality of generally parallel positive and negative electrodes and a plurality of spacers arranged generally at right angles to the electrodes, where:
one wall of the wells is formed by a positive electrode and the opposite wall of the well is formed by a negative electrode;
the spacers form the walls of the wells that are at right angles to the walls formed by the electrodes;
where the electrodes are so disposed that when a preselected voltage is applied across the electrodes the transmembrane potential of cells within the wells is altered.

61. A system for applying electrical field stimulation to cells, said system comprising:

a multiwell tissue culture plate, wherein the bottom of the wells is comprised of an optically transparent filter membrane upon which cells can be grown;

a trough suitable for containing fluid and configured such that said multiwell tissue culture plate may sit therein;

at least one first electrode disposed in said trough; and

an electrode head comprising a plurality of second electrodes in an amount corresponding to the number of wells in said multiwell tissue culture plate, wherein said electrode head and said plurality of said second electrodes are configured such that said plurality of electrodes are disposed in the wells of the multiwell tissue culture plate upon positioning said electrode head onto said multiwell tissue culture plate;

wherein said at least one first electrode and said plurality of said second electrodes are so disposed that when a preselected voltage is applied across the electrodes the transmembrane potential of cells within the wells is altered.

62. The system of claim 61, further comprising a waveform generator that is in electrical communication with said at least one first electrode or said plurality of second electrodes, or both, whereby electric pulse signals are generated by said waveform generator.

63. The system of claim 62 further comprising a computer electrically connected to said waveform generator, said computer comprising software for coordinating said pulse signals produced by said waveform generator.

64. The system of claim 62, wherein said waveform generator generates a binary value that represents the address of the well to be excited by said pulse signals.

65. The system of claim 62, further comprising electrical relays upstream of said plurality of second electrodes.

66. The system of said 65 further comprising a microcontroller in electrical communication with said waveform generator and said electrical relays, so disposed such that upon receiving a trigger pulse and a particular binary value from said waveform generator, said microcontroller switches on the appropriate relay

thereby directing a pulse to the particular electrode corresponding to said particular binary value.

67. The system of claim 61 wherein said trough comprises one first electrode.

68. A system for applying electrical field stimulation to cells, said system comprising:

a multiwell tissue culture plate, wherein the bottom of the wells is comprised of a optically transparent filter membrane upon which cells can be grown;

a tray comprising a plurality of individual troughs suitable for containing fluid; wherein the number of said plurality of troughs corresponds to the amount of wells comprised in said multiwell tissue culture plate; wherein said plurality of troughs are so disposed to individually contain each well of said multiwell tissue culture plate; and wherein said plurality of troughs may be accessed by a port defined in said multiwell tissue culture plate and disposed laterally to each well;

a conductive electrode plate configured to be mounted above said multiwell tissue culture plate; wherein said electrode plate comprises a plurality of apertures configured to allow the wells of the multiwell tissue plate to pass through said conductive electrode plate; wherein said electrode plate comprises a plurality of conductive pins integral or attached to said conductive electrode plate; and wherein individual pins of said plurality of conductive pins pass through said port to be disposed in individual troughs upon mounting said electrode plate on top of said multiwell tissue culture plate; and

an electrode head comprising a plurality of second electrodes in an amount corresponding to the number of wells in said multiwell tissue culture plate, wherein said electrode head and said plurality of said second electrodes are configured such that said plurality of electrodes are disposed in the wells of the multiwell tissue

culture plate upon positioning said electrode head onto said conductive electrode plate;

wherein said at least one first electrode and said plurality of said second electrodes are so disposed that when a preselected voltage is applied across the electrodes the transmembrane potential of cells within the wells is altered.

69. A novel electrode comprising a dielectric disc comprised of a dielectric material; a first conductive disc disposed on one side of said dielectric disc and a second conductive disc disposed on the other side of said dielectric disc.

70. The electrode of claim 69 further comprising a concentric lead, wherein said concentric lead comprises at least one internal lead and at least one external lead whereby said internal lead passes through said first disc and said dielectric disc and is electrically connected to said second disc.

71. The electrode of claim 69 further comprising a first lead electrically connected to said first disc and a second lead electrically connected to said second disc.

72. The electrode of claim 69, wherein when a preselected voltage is applied across said first conductive disc and said second conductive disc to establish and electrical field.

73. The electrode of claim 72, wherein said electrode is able to provide a substantially uniform electrical field, while diminishing ohmic heating to a level such that said electrode may be brought into close proximity to cells to be manipulated.

74. The electrode of claim 73, wherein said electrode may be put in proximity with said cells at a distance of 10mm between said electrode and said cells to a distance closer to said cells without said electrode contacting said cells.

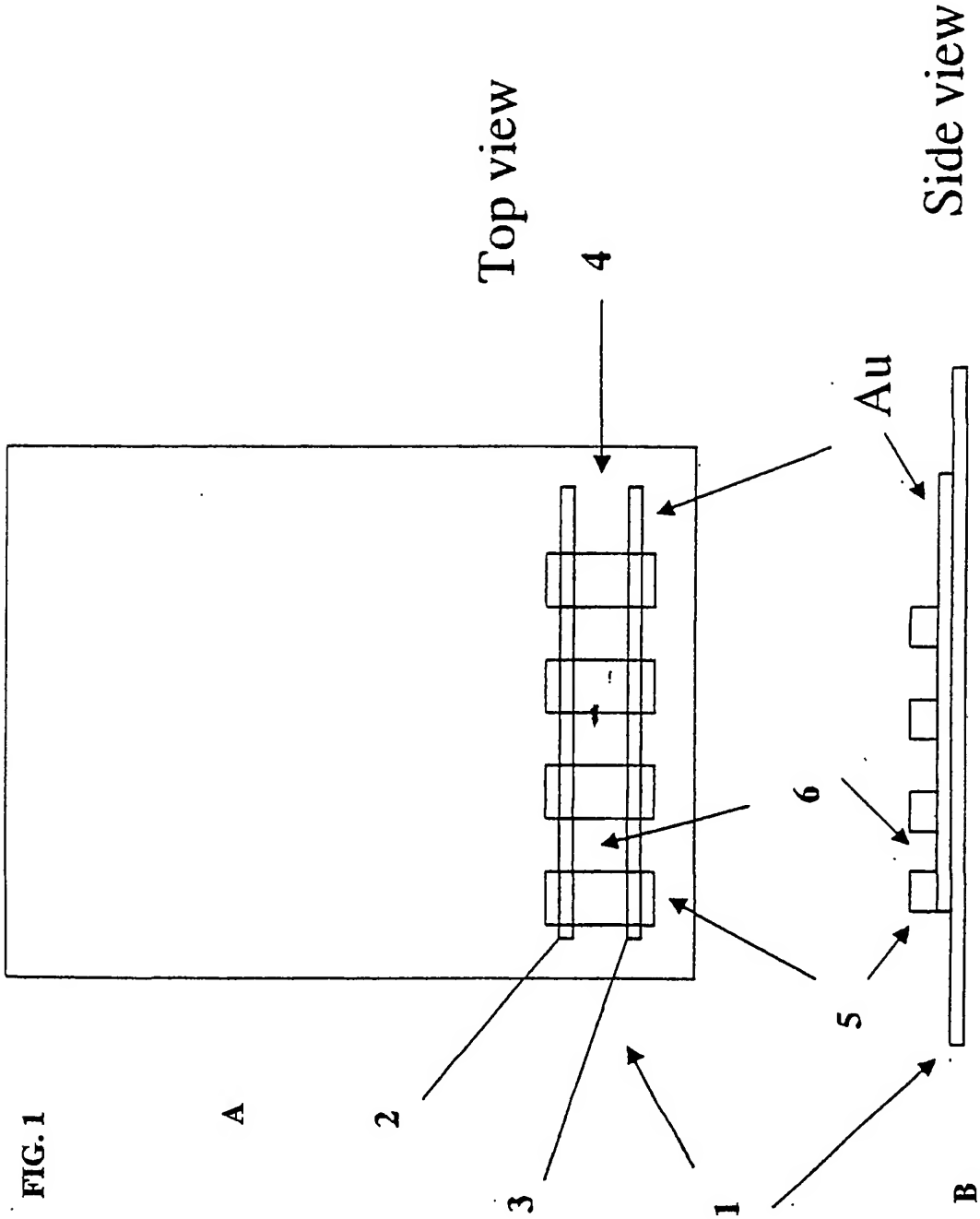


FIG. 1

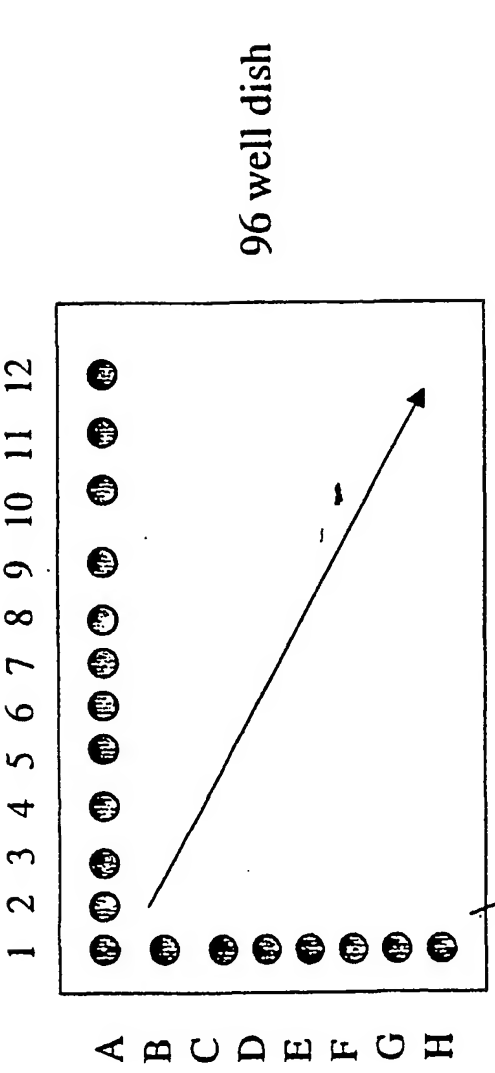

96 well dish
1 2 3 4 5 6 7 8 9 10 11 12
A B C D E F G H
One well not to scale

FIG. 2A

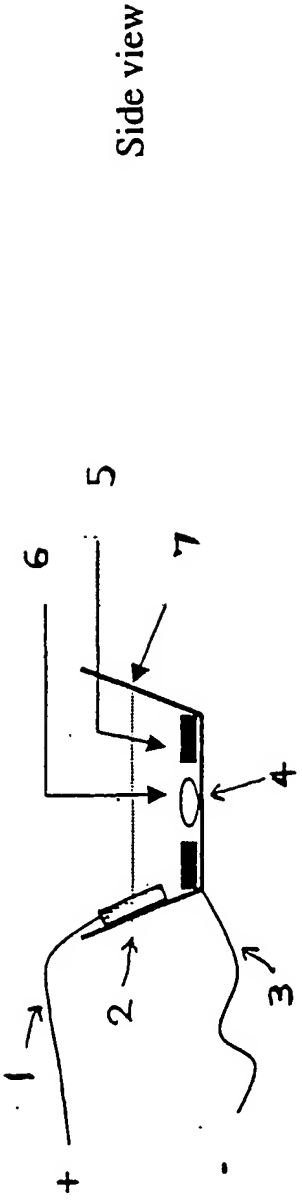

Side view
1
2
3
4
5
6
7
+
-

FIG. 2B

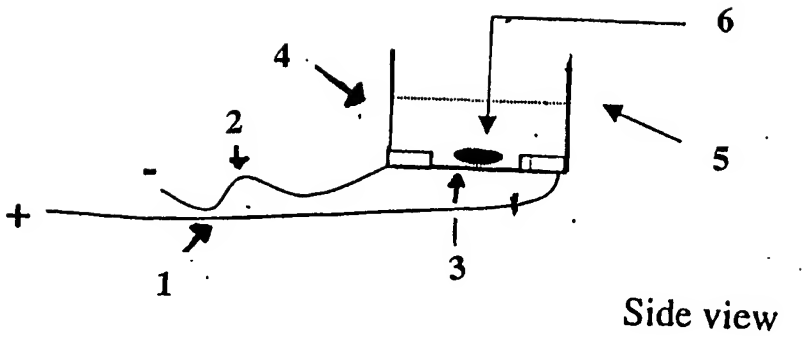


FIG. 2C

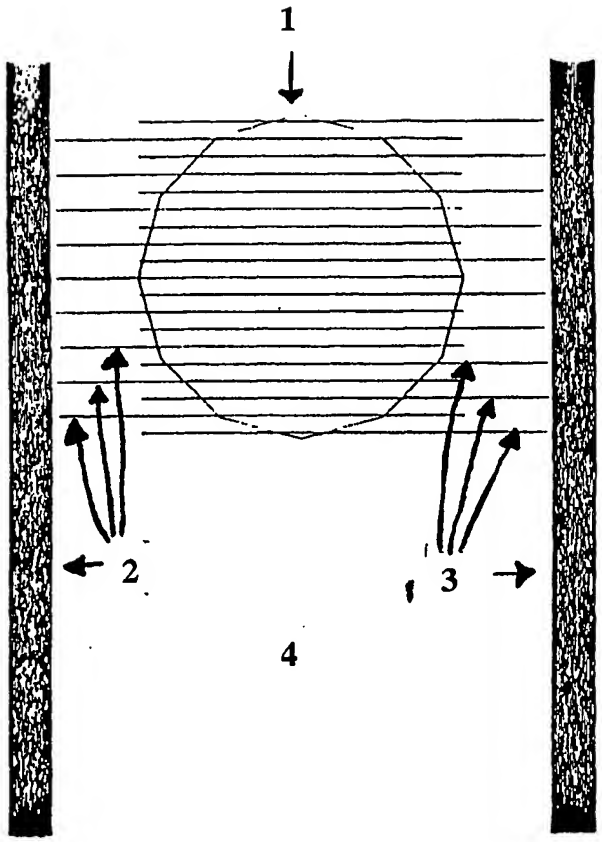


FIG. 3

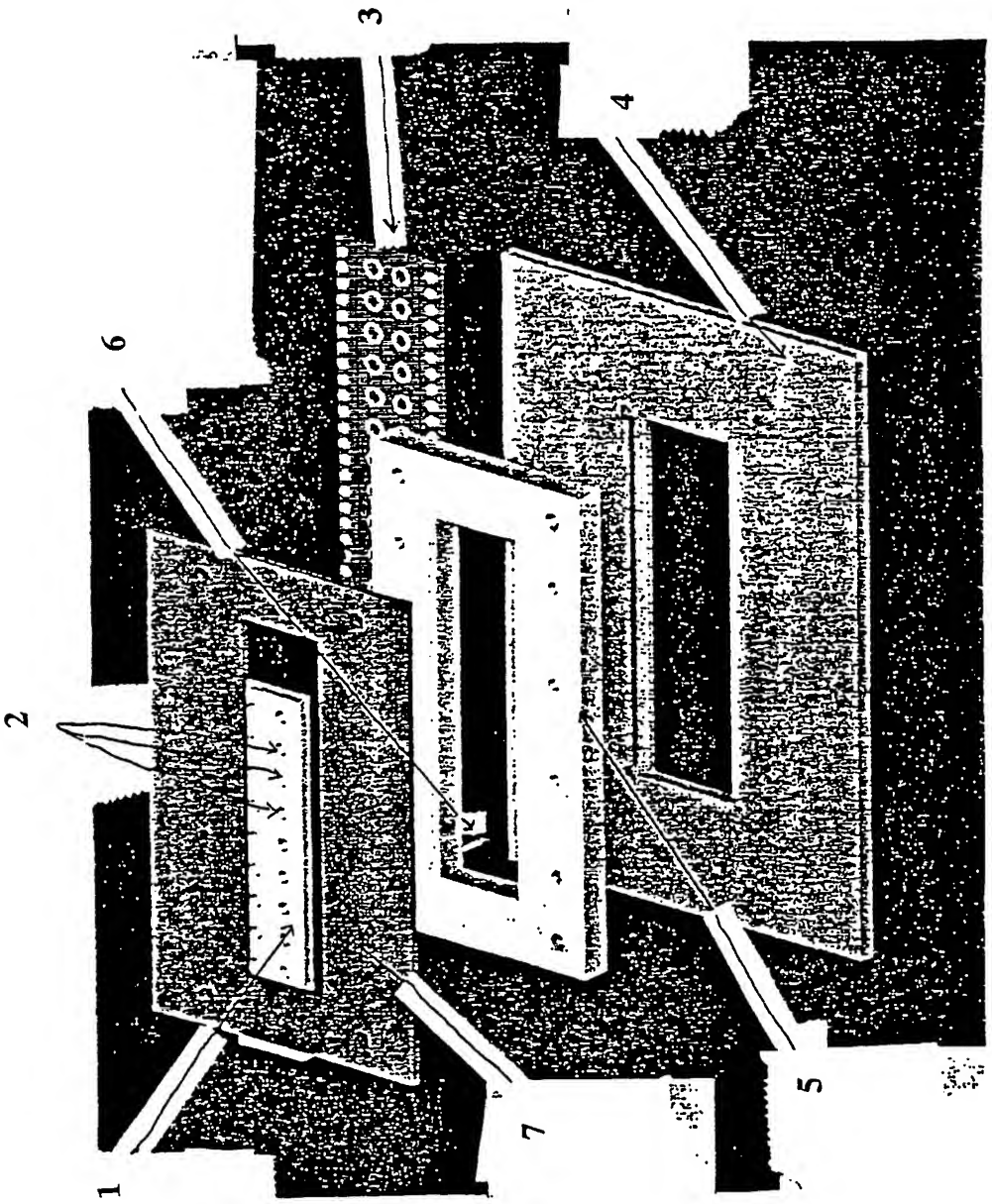


FIG. 4A

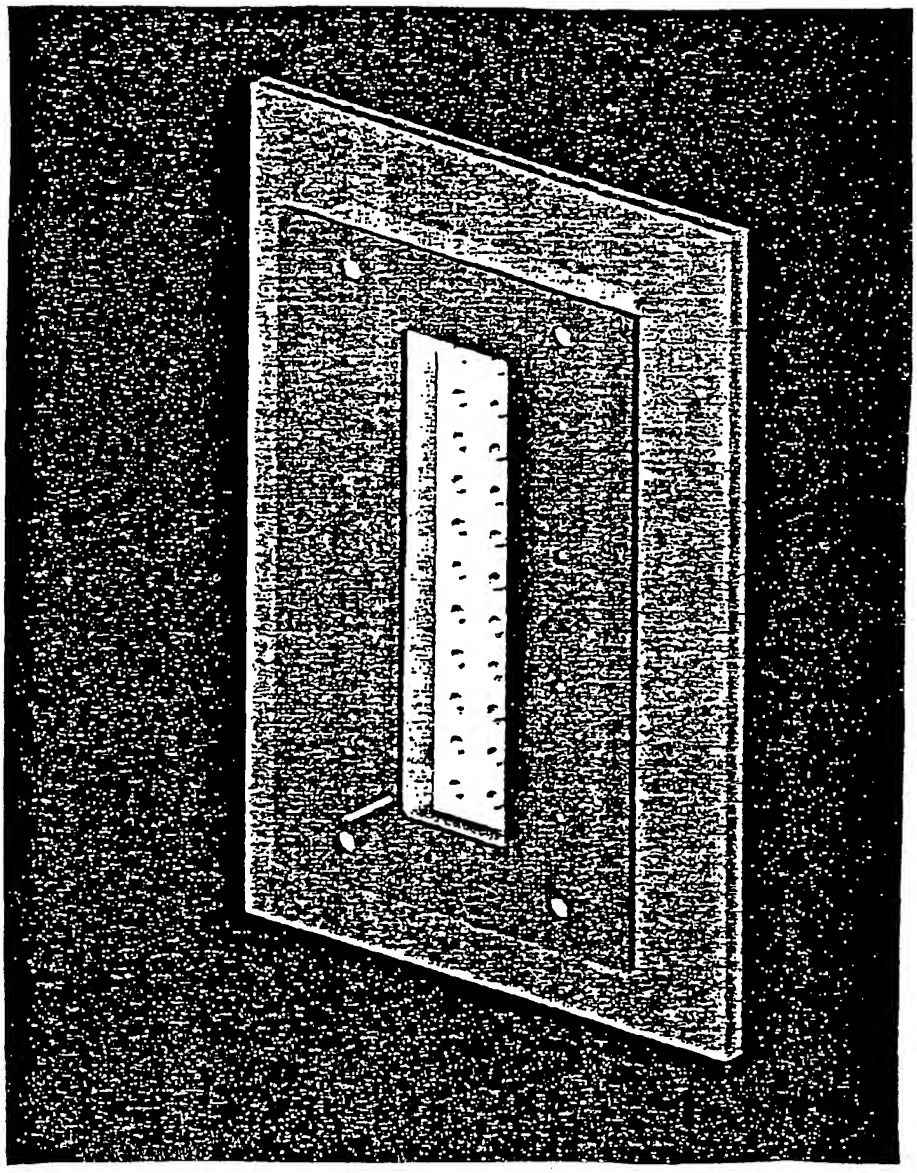


FIG. 4B

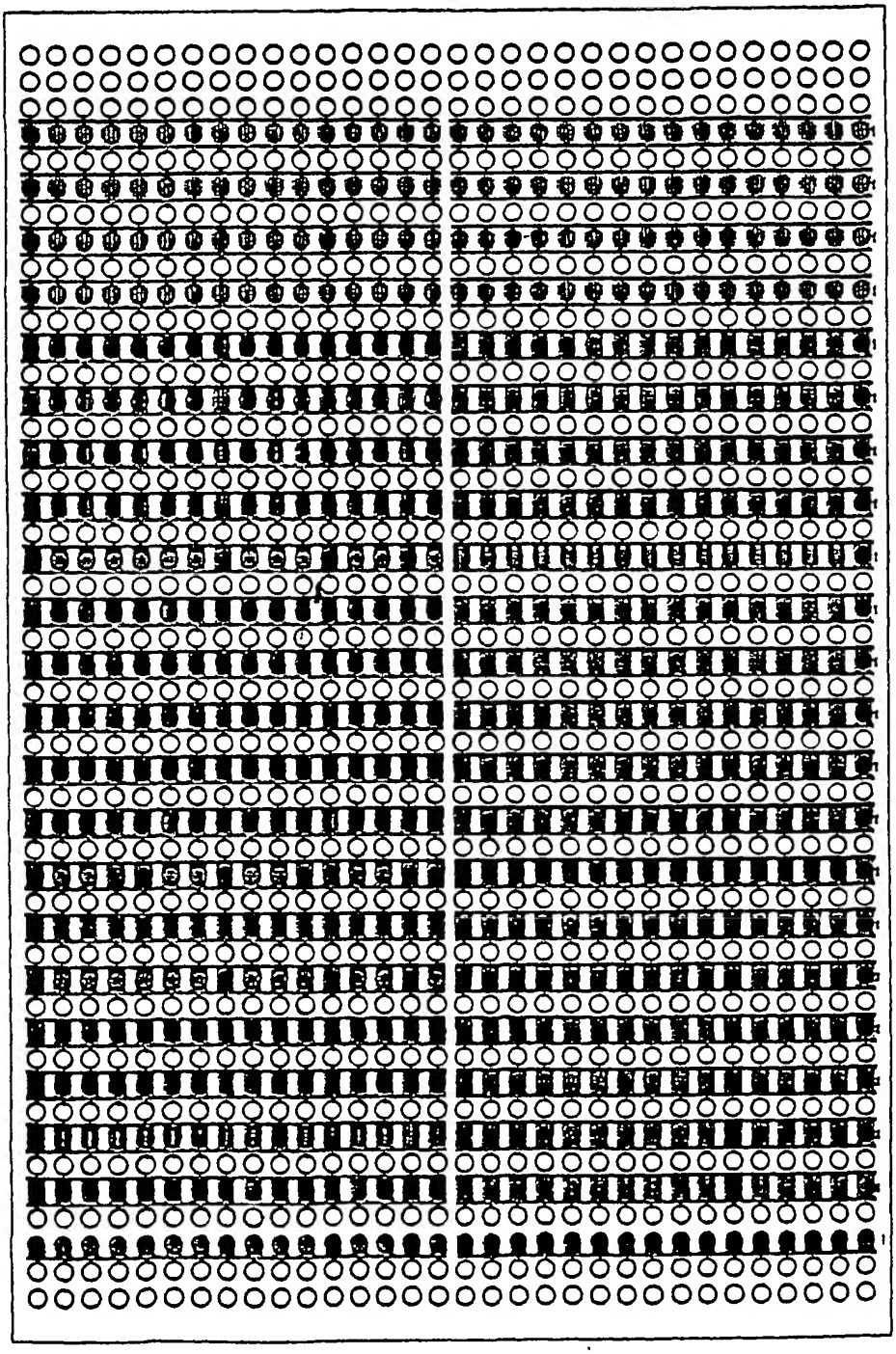


FIG. 5

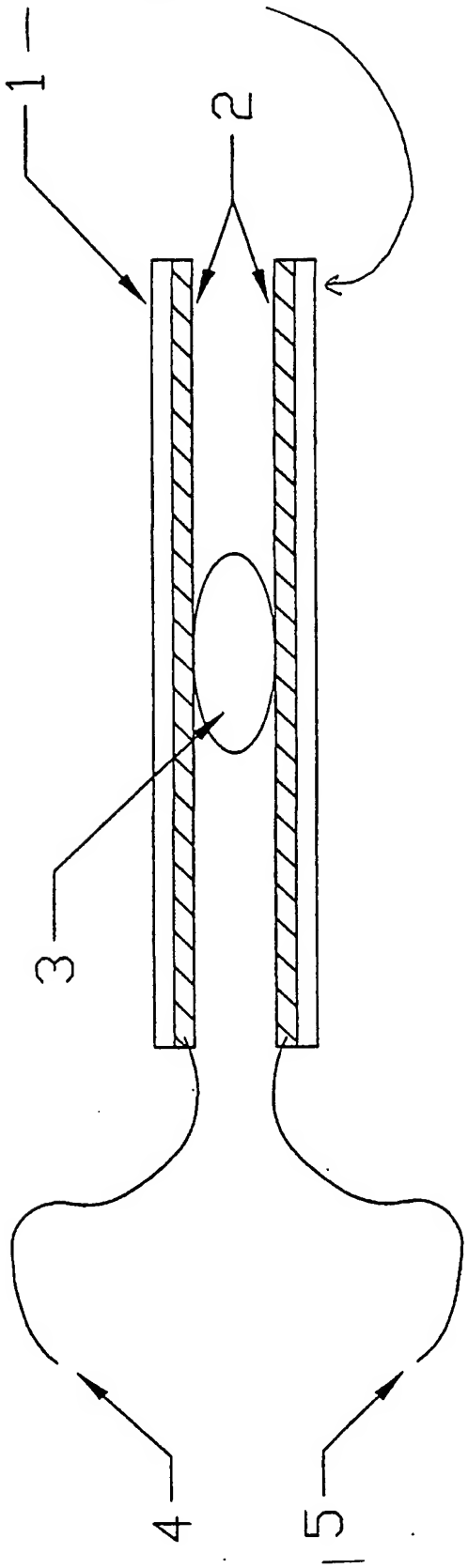


FIG. 6

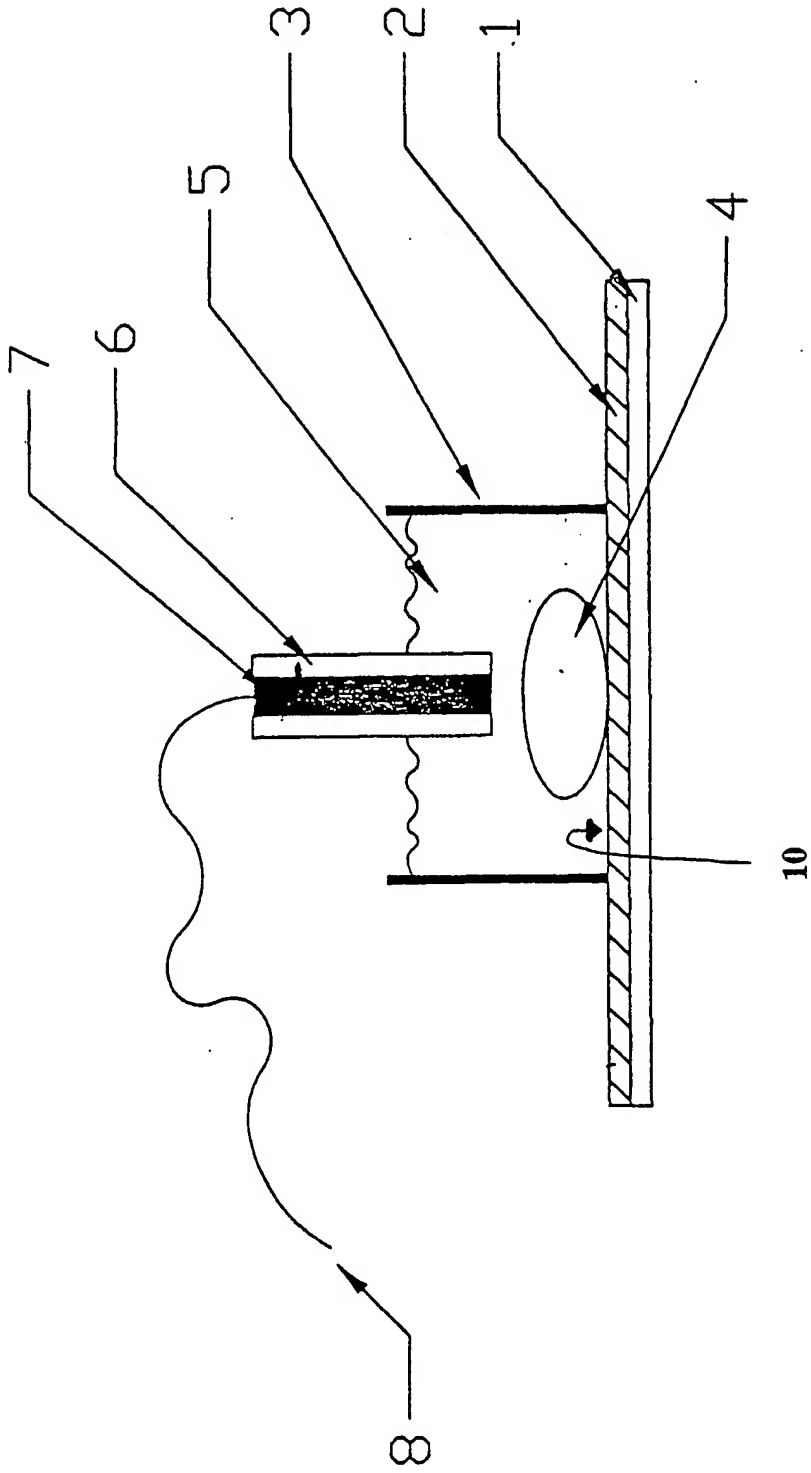


FIG. 7

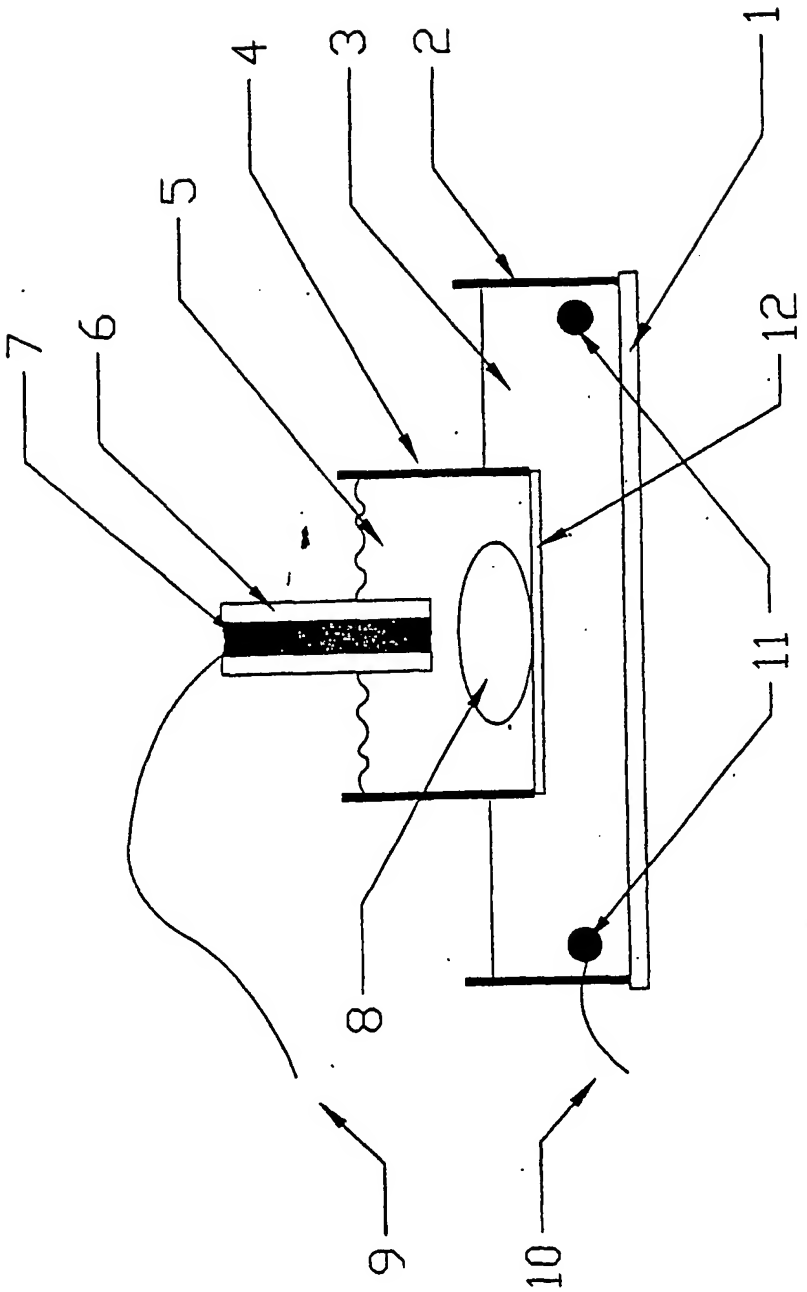


FIG. 8

FIG. 9A

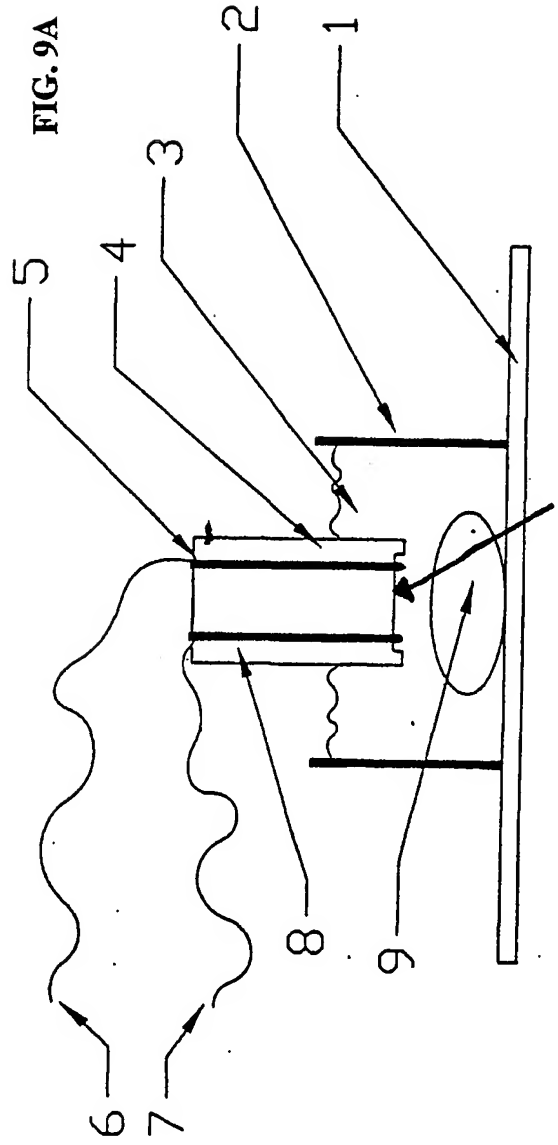
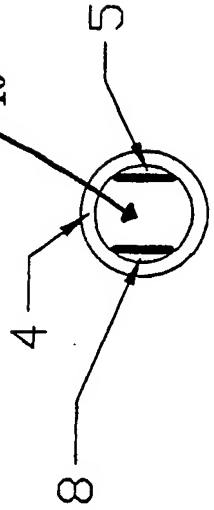


FIG. 9B



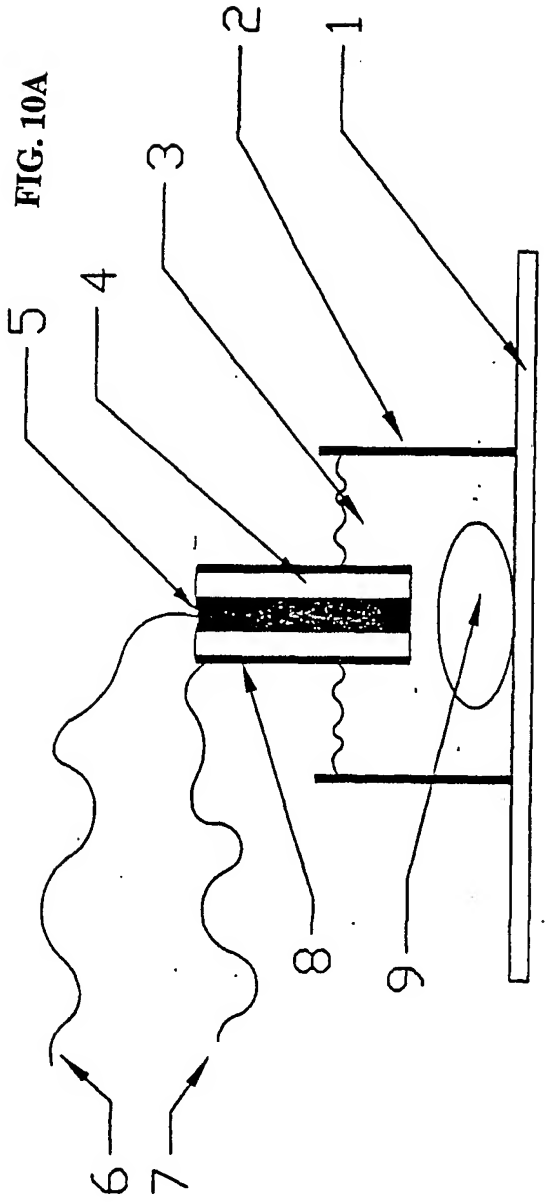


FIG. 10A

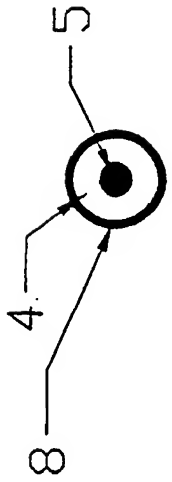


FIG. 10B

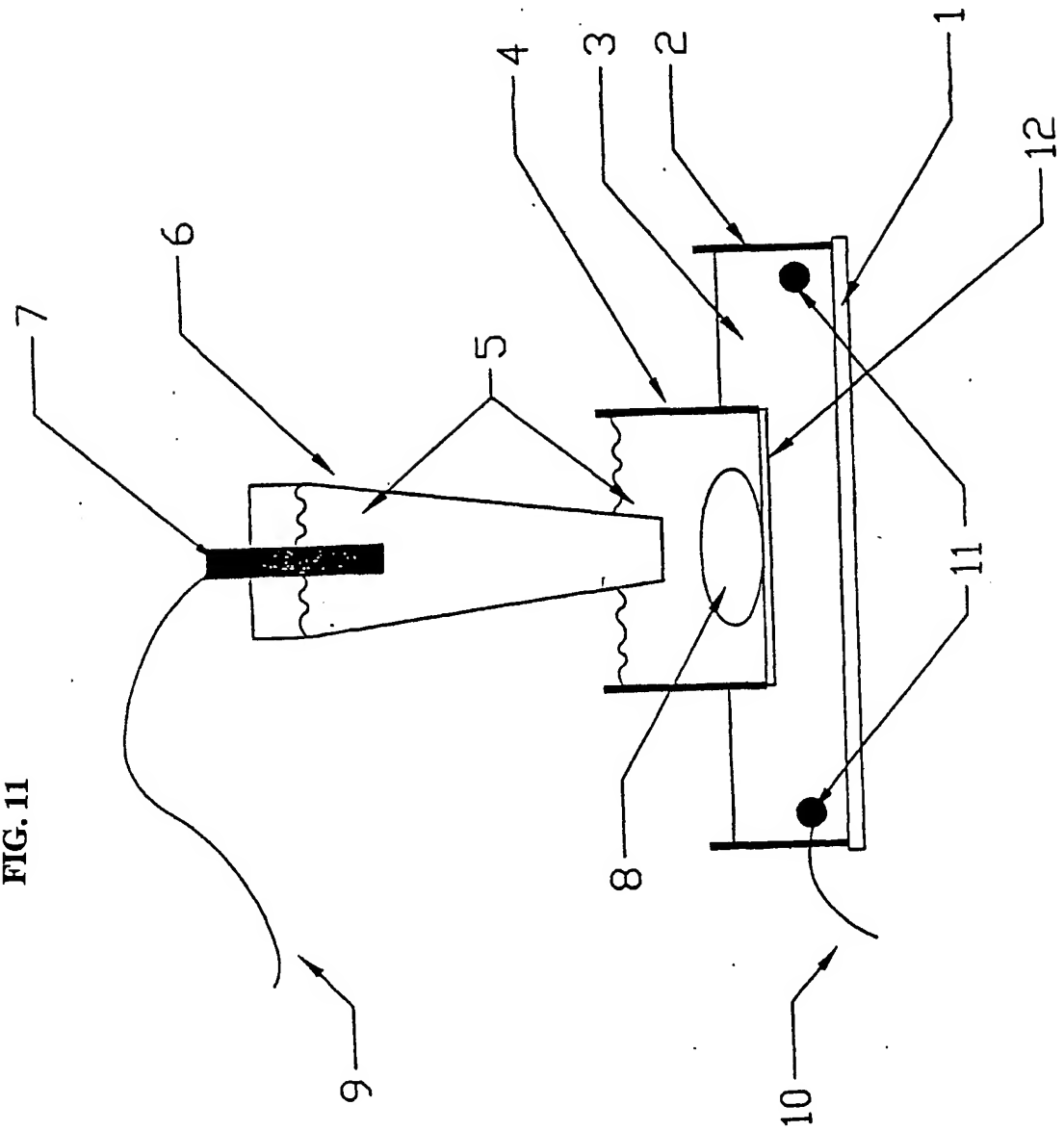


FIG. 11

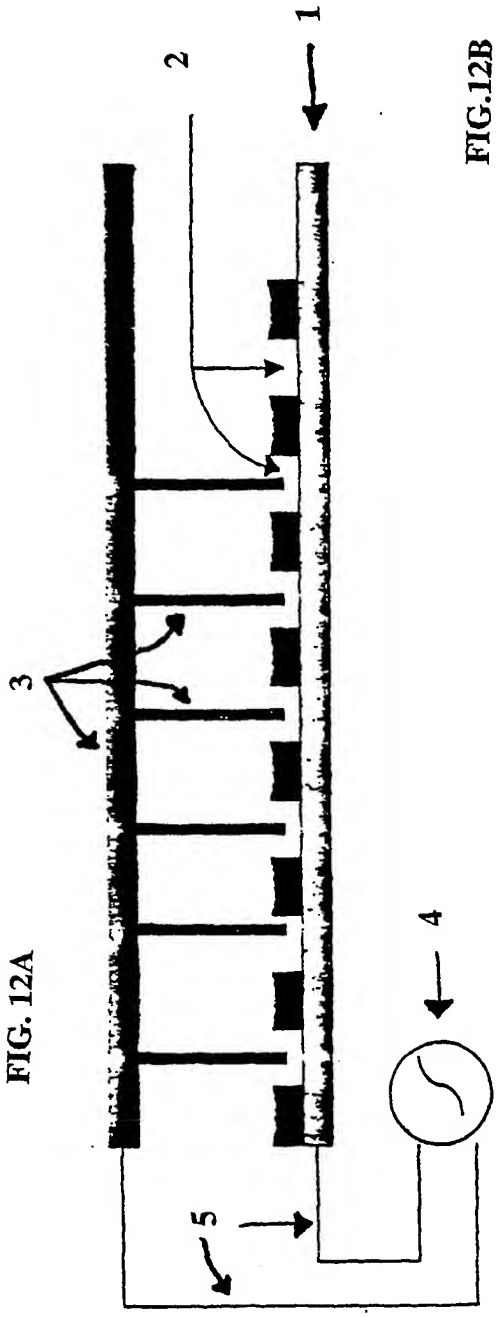


FIG. 12A

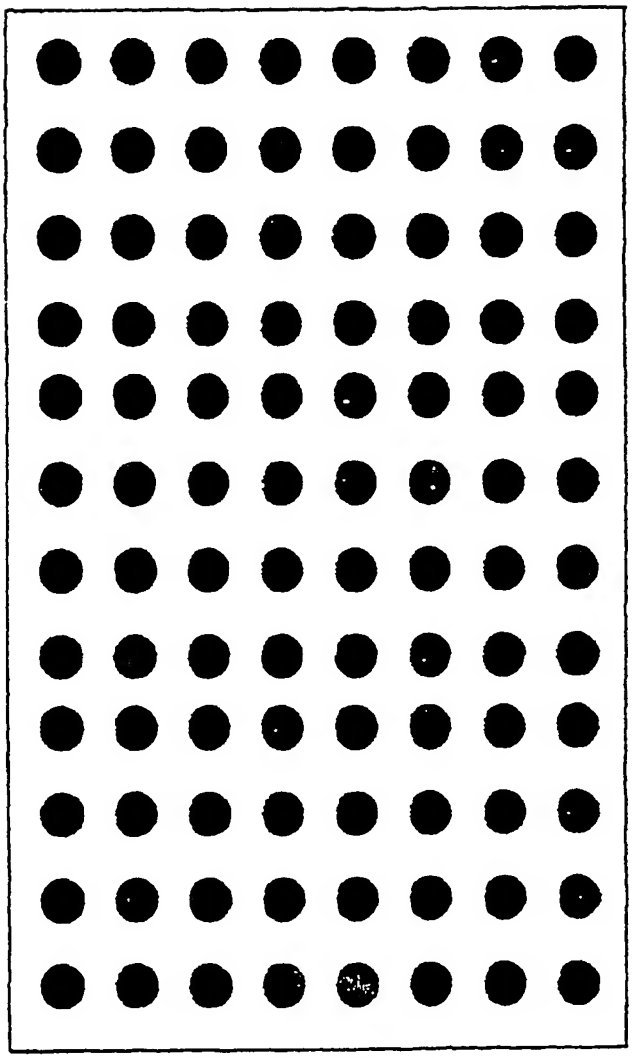


FIG.12B

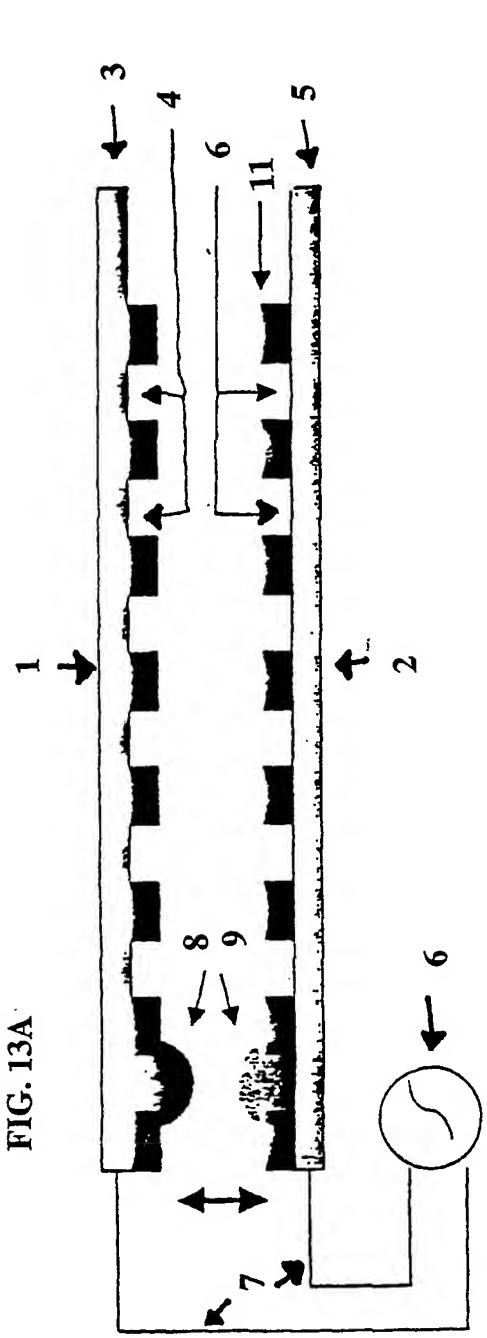


FIG. 13A

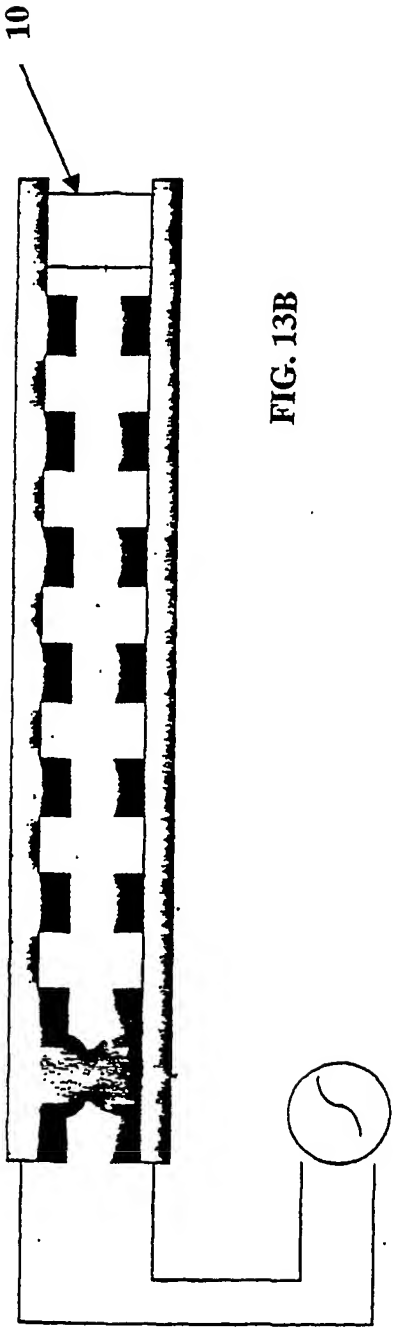


FIG. 13B

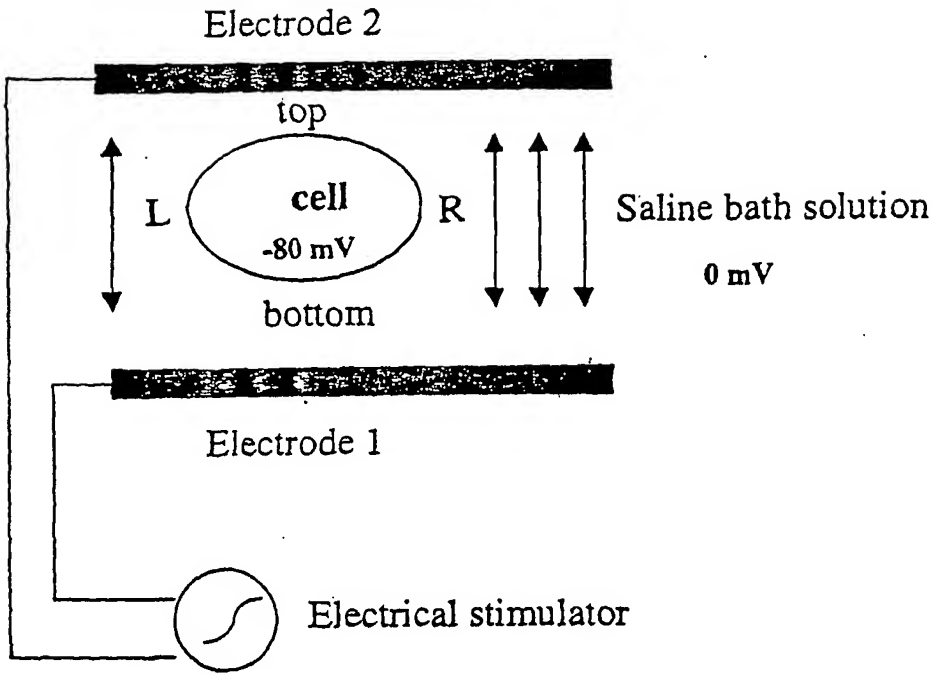

Electrode 2
top
L
cell
-80 mV
R
Saline bath solution
0 mV
bottom
Electrode 1
Electrical stimulator

FIG. 14

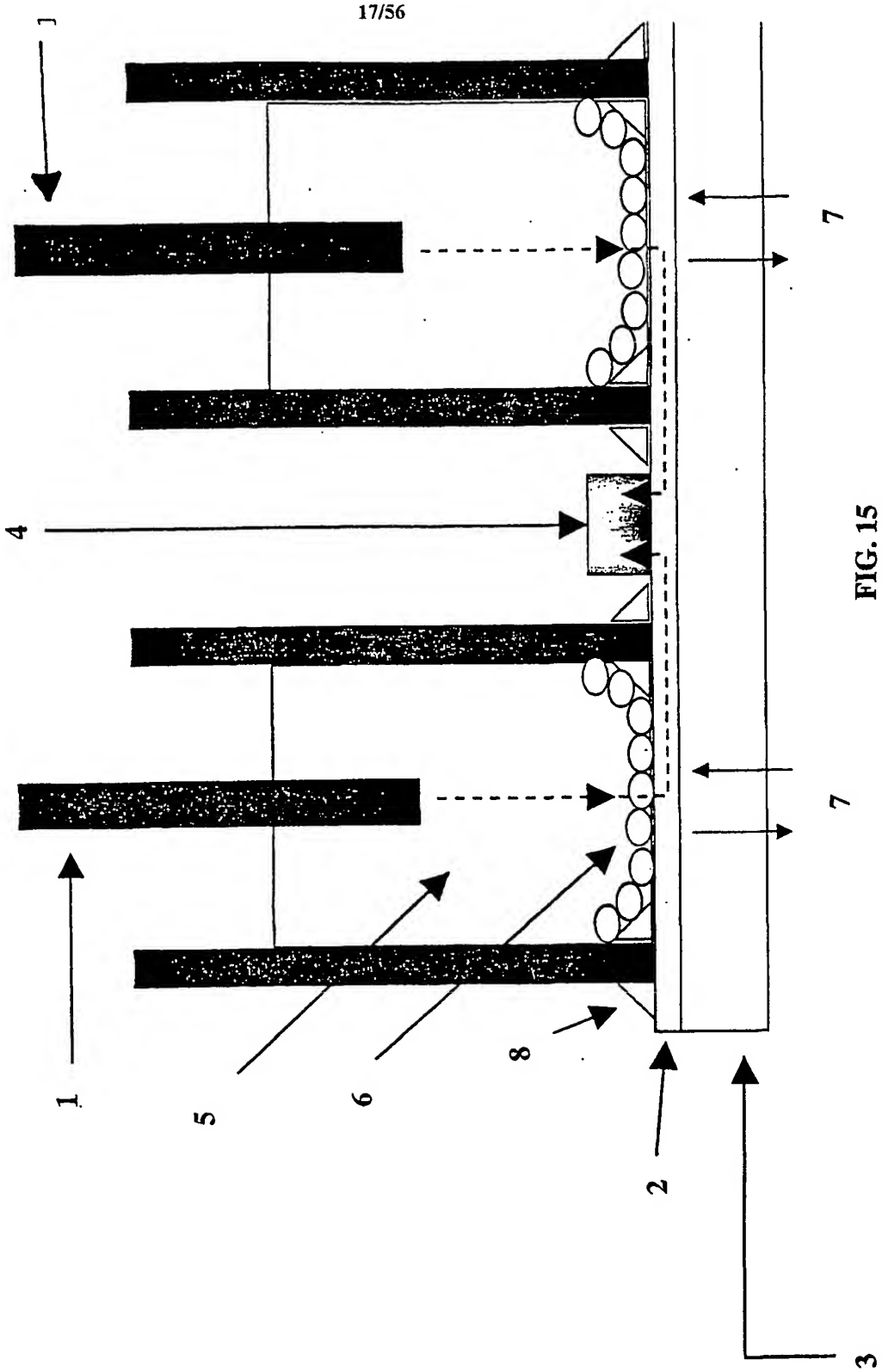


FIG. 15

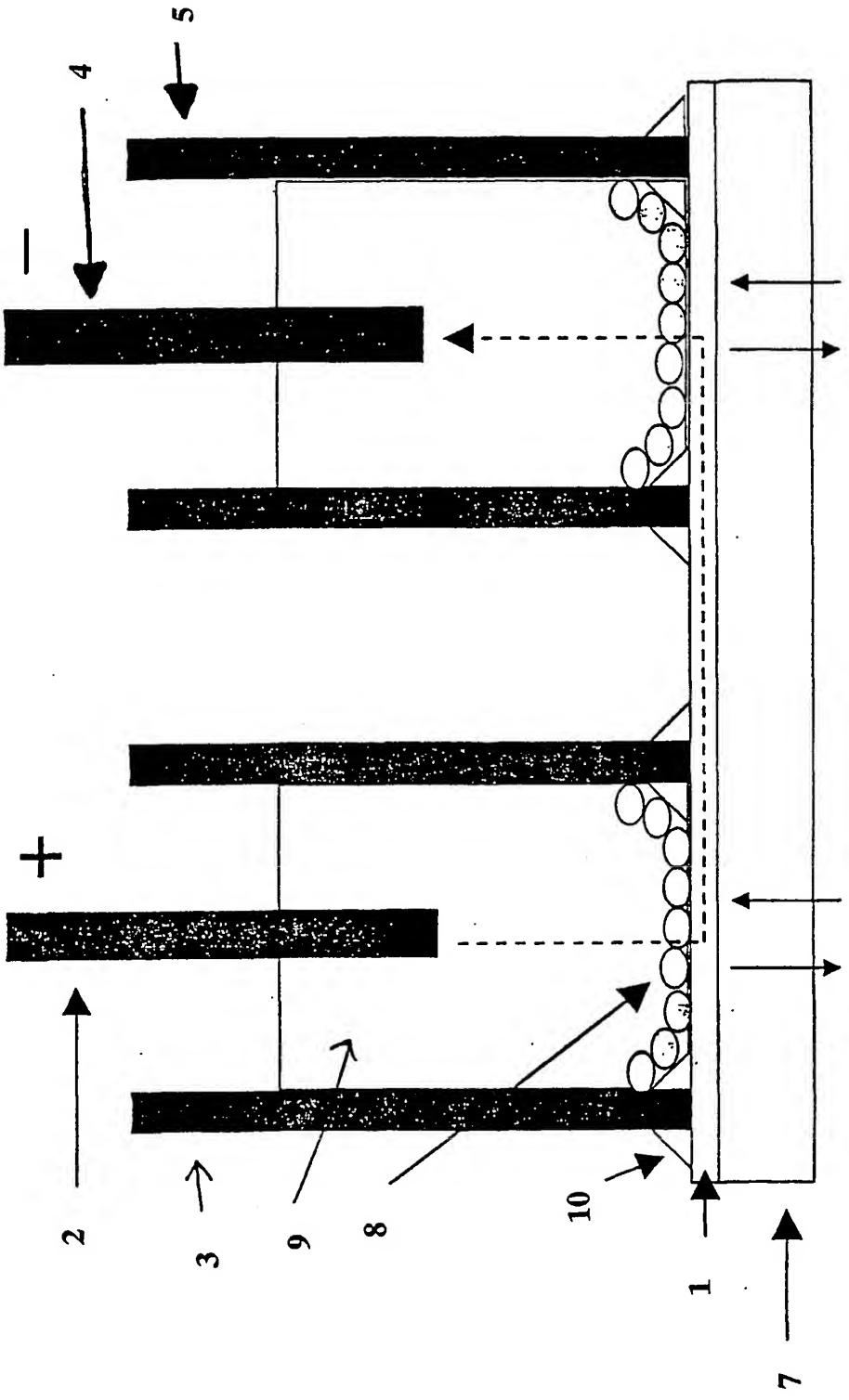


FIG. 16A

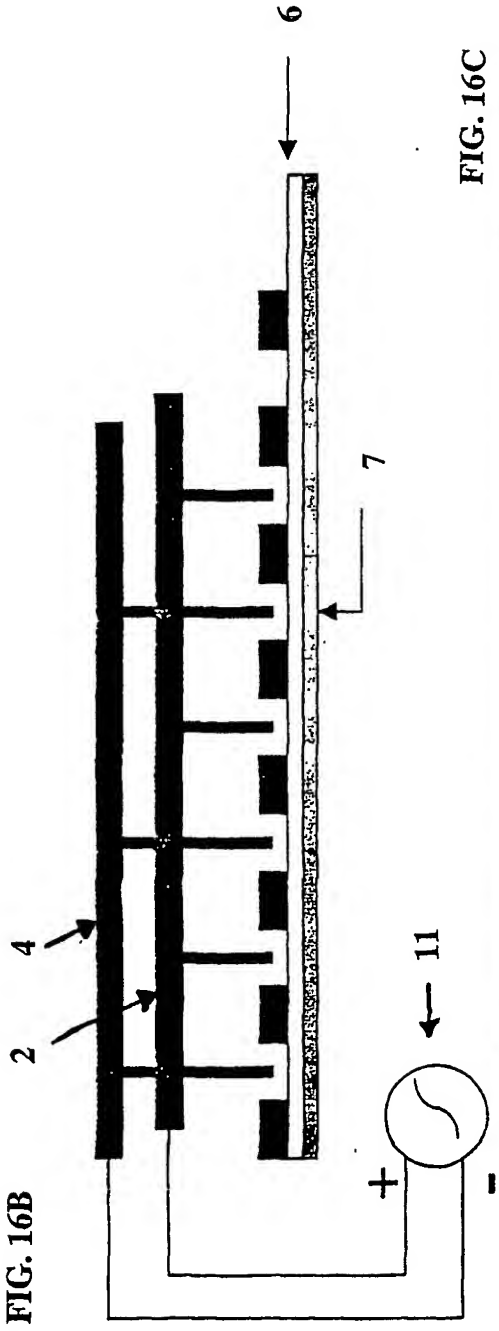


FIG. 16B

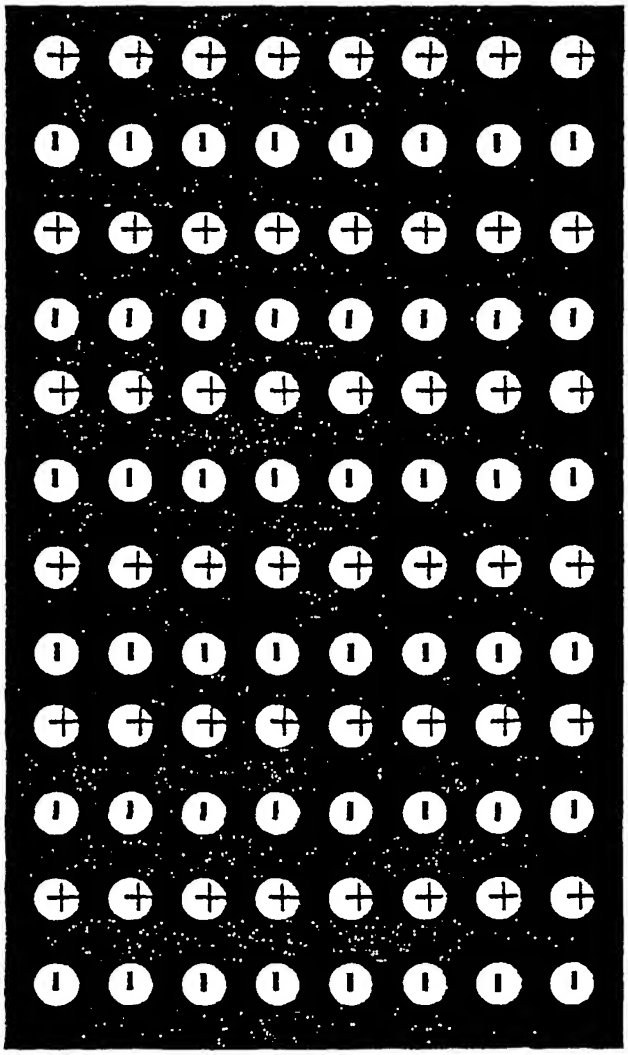


FIG. 16C

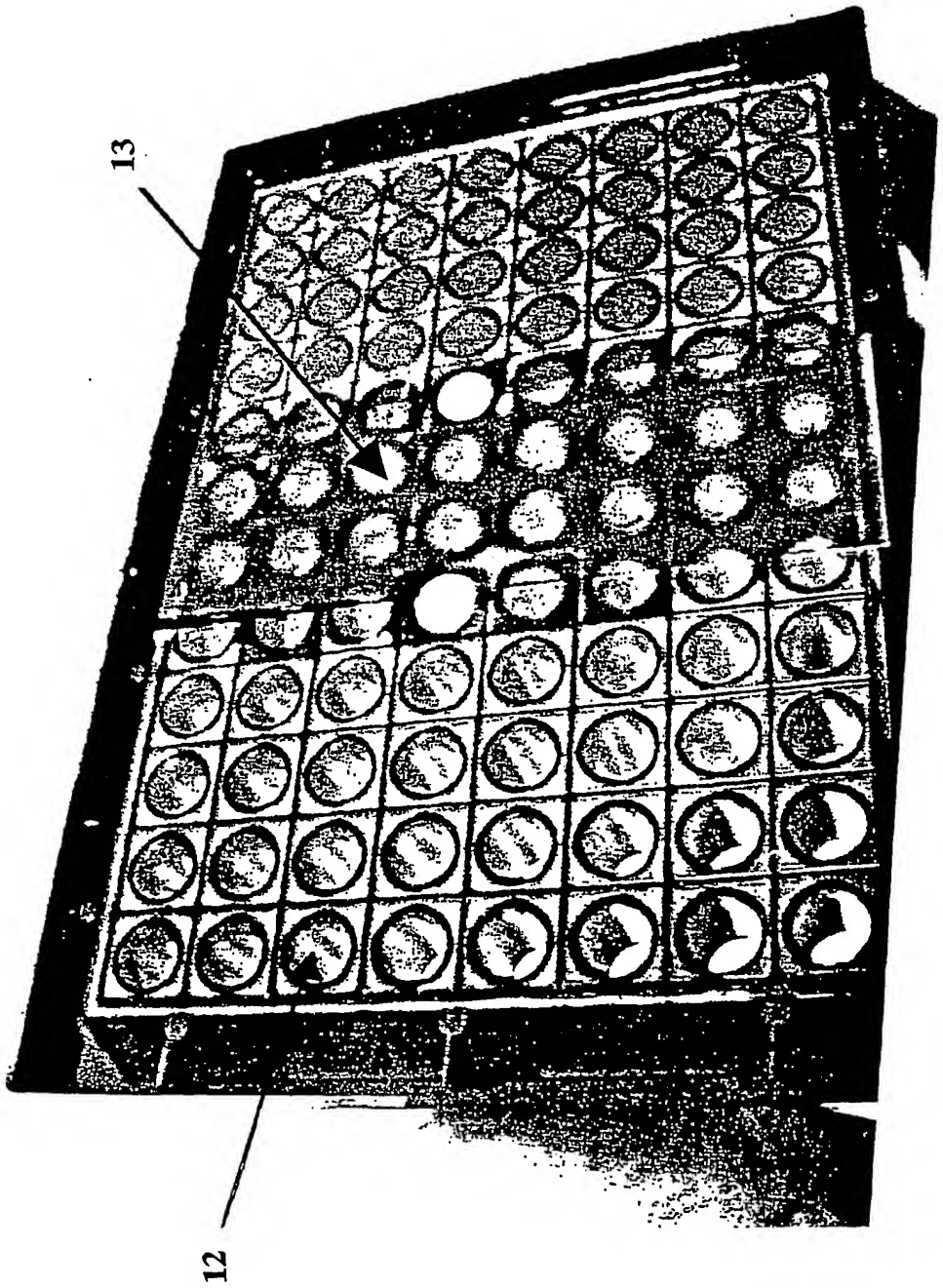


FIG. 16D

FIG. 17

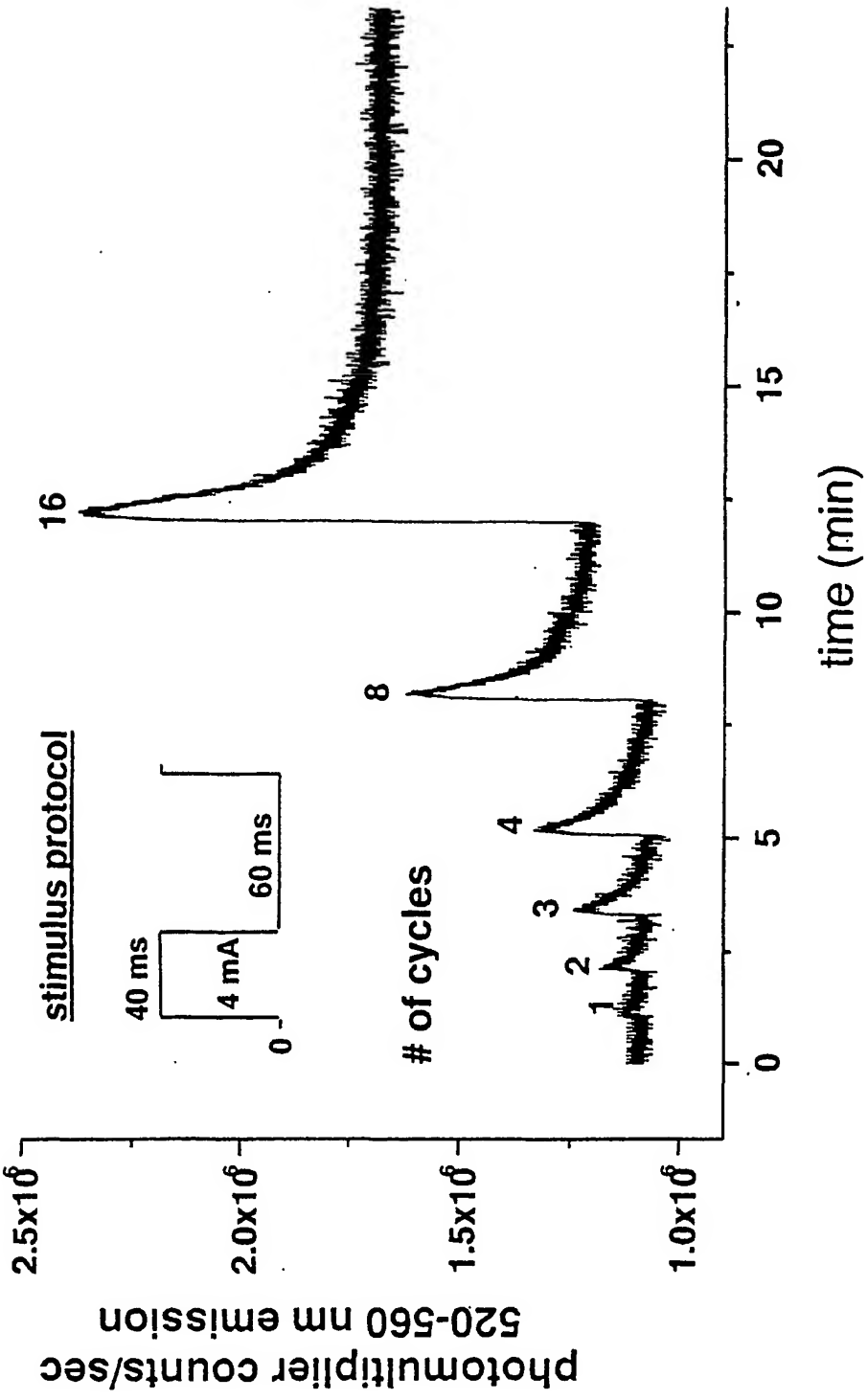

photomultiplier counts/sec
520-560 nm emission
2.5x10^6
2.0x10^6
1.5x10^6
1.0x10^6
0
5
10
15
20
time (min)
stimulus protocol
40 ms
4 mA
60 ms
0
# of cycles
1
2
3
4
8
16

FIGURE 18A

1 atggaattcc ccattggatc cctcgaaact aacaacttcc gtcgctttac tccggagtca
61 ctggtggaga tagagaagca aattgctgcc aagcagggaa caaagaaagc cagagagaag
121 catagggagc agaaggacca agaagagaag cctcggcccc agctggactt gaaagcctgc
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241 gatctagatc cgttctacag cacacaccgg acatttatgg tgctgaacaa agggaggacc
301 atttcccggt ttagtgccac tcgggccctg tggctattca gtcctttcaa cctgatcaga
361 agaacggcca tcaaagtgtc tgtccactcg tggttcagtt tatttattac ggtcactatt
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481 actgtcattt acacctttga agccttgata aagatactgg caagaggatt ttgtctaaat
541 gagttcacgt acctgagaga tccttggaac tggctggatt ttagcgtcat taccctggca
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1261 gccctcgaga tgctccggaa ggagcaggag gtgctagcag cactagggat tgacacaacc
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1441 ccttacaacc agcgcaggat gtcttttcta ggcctcgcct ctggaaaacg ccgggctagt
1501 catggcagtg tgttccattt ccggtcccct ggccgagata tctcactccc tgagggagtc
1561 acagatgatg gagtctttcc tggagaccac gaaagccatc ggggctctct gctgctgggt
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1681 gactccaggc atggagaaga tgaacaccaa ccgccgccca ctagtgagct tgcccctgga
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1801 ttagatgaac ctttccgggc ccaaagggca atgagtgttg tcagtatcat aacctccgtc
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2341 ggaaactcag tgggggcact ggggaacctc accatcatcc tggccatcat tgtctttgtc
2401 tttgctctgg ttggcaagca gctcctaggg gaaaactacc gtaacaaccg aaaaaatatc
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FIG. 18B

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3001 gagggtgaat ctgatcttga tgacttggag gatgatggtg gggaagatgc tcagagcttc
3061 cagcaggaag tgatccccaa aggacagcag gagcagctgc agcaagtcga gaggtgtggg
3121 gaccacctga cacccaggag cccaggcact ggaacatctt ctgaggacct ggctccatcc
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3241 acaagctcct ctgagggcag cacggtggac tgcctagatc ctgaggaaat cctgaggaag
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4141 gaggtcaaca tgcaacccaa gtgggaggac aacgtgtaca tgtatttgta ctttgtcatc
4201 ttcatcattt ttggaggctt cttcacactg aatctctttg ttggggtcat aattgacaac
4261 ttcaatcaac agaaaaaaaa gttaggggggc caggacatct tcatgacaga ggagcagaag
4321 aaatactaca atgccatgaa gaagttgggc tccaagaagc cccagaagcc catcccacgg
4381 cccctgaaca agttccaggg ttttgtcttt gacatcgtga ccagacaagc ttttgacatc
4441 accatcatgg tcctcatctg cctcaacatg atcaccatga tggtggagac tgatgaccaa
4501 agtgaagaaa agacgaaaat tctgggcaaa atcaaccagt tctttgtggc cgtcttcaca
4561 ggcgaatgtg tcatgaagat gttcgctttg aggcagtact acttcacaaa tggctggaat
4621 gtgtttgact tcattgtggt ggttctctcc attgcgagcc tgatttttc tgcaattctt
4681 aagtcacttc aaagttactt ctccccaacg ctcttcagag tcatccgcct ggcccgaatt
4741 ggccgcatcc tcagactgat ccgagcggcc aaggggatcc gcacactgct ctttgccctc
4801 atgatgtccc tgcctgccct cttcaacatc gggctgttgc tattccttgt catgttcatc
4861 tactccatct tcggtatgtc cagctttccc catgtgaggt gggaggctgg catcgacgac
4921 atgttcaact tccagacctt cgccaacagc atgctgtgcc tcttccagat taccacgtcg
4981 gccggctggg atggcctcct cagccccatc ctcaacacag ggccccccta ctgtgacccc
5041 aatctgccca acagcaatgg caccagaggg gactgtggga gcccagccgt aggcatcatc
5101 ttcttcacca cctacatcat catctccttc ctcatcgtgg tcaacatgta cattgcagtg
5161 attctggaga acttcaatgt ggccacggag gagagcactg agcctctgag tgaggacgac
5221 tttgacatgt tctatgagac ctgggagaag tttgacccag aggccactca gtttattacc
5281 ttttctgctc tctcggactt tgcagacact ctctctggtc ccctgagaat cccaaaaccc
5341 aatcgaaata tactgatcca gatggacctg cctttggtcc ctggagataa gatccactgc
5401 ttggacatcc tttttgcttt caccaagaat gtcctaggag aatccgggga gttggattct
5461 ctgaaggcaa atatggagga gaagtttatg gcaactaatc tttcaaaatc atcctatgaa
5521 ccaatagcaa ccactctccg atggaagcaa gaagacattt cagccactgt cattcaaaag
5581 gcctatcgga gctatgtgct gcaccgctcc atggcactct ctaacacccc atgtgtgccc
5641 agagctgagg aggaggctgc atcactccca gatgaaggtt ttgttgcatt cacagcaaat
5701 gaaaattgtg tactcccaga caaatctgaa actgcttctg ccacatcatt cccaccgtcc

FIG. 18C

MEFPIGSLETNNFRRFTPESLVEIEKQIAAKQGTKKAREKHREQ
KDQEEKPRPQLDLKACNQLPKFYGELPAELIGEPLEDLDPFYSTHRTFMVLNKGRTIS
RFSATRALWLFSPFNLIRRTAIKVSVHSWFSLFITVTILVNCVCMTRTDLPEKIEYVF
TVIYTFEALIKILARGFCLNEFTYLRDPWNWLDFSVITLAYVGTAIDLRGISGLRTFR
VLRALKTVSVIPGLKVIVGALIHSVKKLADVTILTIFCLSVFALVGLQLFKGNLKNKC
VKNDMAVNETTNYSSHRKPDIYINKRGTSDPLLCGNGSDSGHCPDGYICLKTSDNPDF
NYTSFDSFAWAFLSLFRLMTQDSWERLYQQTLRTSGKIYMIFFVLVIFLGSFYLVNLI
LAVVTMAYEEQNQATTDEIEAKEKKFQEALEMLRKEQEVLAALGIDTTSLHSHNGSPL
TSKNASERRHRIKPRVSEGSTEDNKSPRSDPYNQRRMSFLGLASGKRRASHGSVFHFR
SPGRDISLPEGVTDDGVFPGDHESHRGSLLLGGGAGQQGPLPRSPLPQPSNPDSRHGE
DEHQPPPTSELAPGAVDVSAFDAGQKKTFLSAEYLDEPFRAQRAMSVVSIITSVLEEL
EESEQKCPPCLTSLSQKYLIWDCCPMWVKLKTILFGLVTDPFAELTITLCIVVNTIFM
AMEHHGMSPTFEAMLQIGNIVFTIFFTAEMVFKIIAFDPYYYFQKKWNIFDCIIVTVS
LLELGVAKKGSLSVLRSFRLLRVFKLAKSWPTLNTLIKIIGNSVGALGNLTIILAIIV
FVFALVGKQLLGENYRNNRKNISAPHEDWPRWHMHDFFHSFLIVFRILCGEWIENMWA
CMEVGQKSICLILFLTVMVLGNLVVLNLFIALLLNSFSADNLTAPEDDGEVNNLQVAL
ARIQVFGHRTKQALCSFFSRSCPFPQPKAEPELVVKLPLSSSKAENHIAANTARGSSG
GLQAPRGPRDEHSDFIANPTVWVSVPIAEGESDLDDLEDDGGEDAQSFQQEVIPKGQQ
EQLQQVERCGDHLTPRSPGTGTSSEDLAPSLGETWKDESVPQAPAEGVDDTSSSEGST
VDCLDPEEILRKIPELADDLEEPDDCFTEGCIRHCPCCKLDTTKSPWDVGWQVRKTCY
RIVEHSWFESFIIFMILLSSGSLAFEDYYLDQKPTVKALLEYTDRVFTFIFVFEMLLK
WVAYGFKKYFTNAWCWLDFLIVNISLISLTAKILEYSEVAPIKALRTLRALRPLRALS
RFEGMRVVVDALVGAIPSIMNVLLVCLIFWLIFSIMGVNLFAGKFWRCINYTDGEFSL
VPLSIVNNKSDCKIQNSTGSFFWVNVKVNFDNVAMGYLALLQVATFKGWMDIMYAAVD
SREVNMQPKWEDNVYMYLYFVIFIIFGGFFTLNLFVGVIIDNFNQQKKKLGGQDIFMT
EEQKKYYNAMKKLGSKKPQKPIPRPLNKFQGFVFDIVTRQAFDITIMVLICLNMITMM
VETDDQSEEKTKILGKINQFFVAVFTGECVMKMFALRQYYFTNGWNVFDFIVVVLSIA
SLIFSAILKSLQSYFSPTLFRVIRLARIGRILRLIRAAKGIRTLLFALMMSLPALFNI
GLLLFLVMFIYSIFGMSSFPHVRWEAGIDDMFNFQTFANSMLCLFQITTSAGWDGLLS
PILNTGPPYCDPNLPNSNGTRGDCGSPAVGIIFFTTYIIISFLIVVNMYIAVILENFN
VATEESTEPLSEDDFDMFYETWEKFDPEATQFITFSALSDFADTLSGPLRIPKPNRNI
LIQMDLPLVPGDKIHCLDILFAFTKNVLGESGELDSLKANMEEKFMATNLSKSSYEPI
ATTLRWKQEDISATVIQKAYRSYVLHRSMALSNTPCVPRAEEEAASLPDEGFVAFTAN
ENCVLPDKSETASATSFPPSYESVTRGLSDRVNMRTSSSIQNEDEATSMELIAPGP

FIGURE 19A

1 cgaggccgcc gccgtcgcct ccgccgggcg agccggagcc ggagtcgagc cgcggccggg
61 agccgggcgg gctggggacg cgggccgggg gcggaggcgc tgggggccgg ggccgggggcc
121 gggggcggag gcgctggggg ccggggccgg ggccgggcgc cgagcggggt ccgcggtgac
181 cgcgccgccc gggcgatgcc cgcgggggacg ccgccggcca gcagagcgag gtgctgccgg
241 ccgccaccat gaccgagggc gcacgggccg ccgacgaggt ccgggtgccc ctgggcgcgc
301 cgccccctgg ccctgcggcg ttggtggggg cgtccccgga gagccccggg gcgccgggac
361 gcgaggcgga gcgggggtcc gagctcggcg tgtcaccctc cgagagcccg gcggccgagc
421 gcggcgcgga gctgggtgcc gacgaggagc agcgcgtccc gtacccggcc ttggcggcca
481 cggtcttctt ctgcctcggt cagaccacgc ggccgcgcag ctggtgcctc cggctggtct
541 gcaacccatg gttcgagcac gtgagcatgc tggtaatcat gctcaactgc gtgaccctgg
601 gcatgttccg gccctgtgag gacgttgagt gcggctccga gcgctgcaac atcctggagg
661 cctttgacgc cttcattttc gccttttttg cggtggagat ggtcatcaag atggtggcct
721 tggggctgtt cgggcagaag tgttacctgg gtgacacgtg gaacaggctg gatttcttca
781 tcgtcgtggc gggcatgatg gagtactcgt tggacggaca caacgtgagc ctctcggcta
841 tcaggaccgt gcgggtgctg cggcccctcc gcgccatcaa ccgcgtgcct agcatgcgga
901 tcctggtcac tctgctgctg gatacgctgc ccatgctcgg gaacgtcctt ctgctgtgct
961 tcttcgtctt cttcattttc ggcatcgttg gcgtccagct ctgggctggc ctcctgcgga
1021 accgctgctt cctggacagt gcctttgtca ggaacaacaa cctgaccttc ctgcggccgt
1081 actaccagac ggaggagggc gaggagaacc cgttcatctg ctcctcacgc cgagacaacg
1141 gcatgcagaa gtgctcgcac atccccggcc gccgcgagct gcgcatgccc tgcaccctgg
1201 gctgggaggc ctacacgcag ccgcaggccg agggggtggg cgctgcacgc aacgcctgca
1261 tcaactggaa ccagtactac aacgtgtgcc gctcgggtga ctccaacccc cacaacggtg
1321 ccatcaactt cgacaacatc ggctacgcct ggattgccat cttccaggtg atcacgctgg
1381 aaggctgggt ggacatcatg tactacgtca tggacgccca ctcattctac aacttcatct
1441 atttcatcct gctcatcatc gtgggctcct tcttcatgat caacctgtgc ctggtggtga
1501 ttgccacgca gttctcggag acgaagcagc gggagagtca gctgatgcgg gagcagcggg
1561 cacgccacct gtccaacgac agcacgctgg ccagcttctc cgagcctggc agctgctacg
1621 aagagctgct gaagtacgtg ggccacatat tccgcaaggt caagcggcgc agcttgcgcc
1681 tctacgcccg ctggcagagc cgctggcgca agaaggtgga ccccagtgct gtgcaaggcc
1741 agggtcccgg gcaccgccag cgccgggcag gcaggcacac agcctcggtg caccacctgg
1801 tctaccacca ccatcaccac caccaccacc actaccattt cagccatggc agcccccgca
1861 ggcccggccc cgagccaggc gcctgcgaca ccaggctggt ccgagctggc gcgccccccct
1921 cgccaccttc cccaggccgc ggacccccccg acgcagagtc tgtgcacagc atctaccatg
1981 ccgactgcca catagagggg ccgcaggaga gggcccgggt ggcacatgcc gcagccactg
2041 ccgctgccag cctcaggctg gccacagggc tgggcaccat gaactacccc acgatcctgc
2101 cctcaggggt gggcagcggc aaaggcagca ccagccccgg acccaagggg aagtgggccg
2161 gtggaccgcc aggcaccggg gggcacggcc cgttgagctt gaacagccct gatccctacg
2221 agaagatccc gcatgtggtc ggggagcatg gactgggcca ggcccctggc catctgtcgg
2281 gcctcagtgt gccctgcccc ctgcccagcc ccccagcggg cacactgacc tgtgagctga
2341 agagctgccc gtactgcacc cgtgccctgg aggacccgga gggtgagctc agcggctcgg
2401 aaagtggaga ctcagatggc cgtggcgtct atgaattcac gcaggacgtc cggcacggtg
2461 accgctggga ccccacgcga ccaccccgtg cgacggacac accaggccca ggcccaggca
2521 gcccccagcg gcgggcacag cagagggcag ccccgggcga gccaggctgg atgggccgcc
2581 tctgggttac cttcagcggc aagctgcgcc gcatcgtgga cagcaagtac ttcagccgtg
2641 gcatcatgat ggccatcctt gtcaacacgc tgagcatggg cgtggagtac catgagcagc
2701 ccgaggagct gactaatgct ctggagatca gcaacatcgt gttcaccagc atgtttgccc
2761 tggagatgct gctgaagctg ctggcctgcg gccctctggg ctacatccgg aacccgtaca
2821 acatcttcga cggcatcatc gtggtcatca gcgtctggga gatcgtgggg caggcggacg

FIG. 19B

2881 gtggcttgtc tgtgctgcgc accttccggc tgctgcgtgt gctgaagctg gtgcgctttc
2941 tgccagccct gcggcgccag ctcgtggtgc tggtgaagac catggacaac gtggctacct
3001 tctgcacgct gctcatgctc ttcattttca tcttcagcat cctgggcatg caccttttcg
3061 gctgcaagtt cagcctgaag acagacaccg gagacaccgt gcctgacagg aagaacttcg
3121 actccctgct gtgggccatc gtcaccgtgt tccagatcct gacccaggag gactggaacg
3181 tggtcctgta caacggcatg gcctccacct cctcctgggc cgccctctac ttcgtggccc
3241 tcatgacctt cggcaactat gtgctcttca acctgctggt ggccatcctc gtggaggggct
3301 tccaggcgga gggcgatgcc aacagatccg acacggacga ggacaagacg tcggtccact
3361 tcgaggagga cttccacaag ctcagagaac tccagaccac agagctgaag atgtgttccc
3421 tggccgtgac ccccaacggg cacctggagg gacgaggcag cctgtccccc cccctcatca
3481 tgtgcacagc tgccacgccc atgcctaccc ccaagagctc accattcctg gatgcagccc
3541 ccagcctccc agactctcgg cgtggcagca gcagctccgg ggacccgcca ctgggagacc
3601 agaagcctcc ggccagcctc cgaagttctc cctgtgcccc ctggggcccc agtggcgcct
3661 ggagcagccg gcgctccagc tggagcagcc tgggccgtgc ccccagcctc aagcgccgcg
3721 gccagtgtgg ggaacgtgag tccctgctgt ctggcgaggg caagggcagc accgacgacg
3781 aagctgagga cggcagggcc gcgcccgggc cccgtgccac cccactgcgg cgggccgagt
3841 ccctggaccc acggcccctg cggccggccg ccctcccgcc taccaagtgc cgcgatcgcg
3901 acgggcaggt ggtggccctg cccagcgact tcttcctgcg catcgacagc caccgtgagg
3961 atgcagccga gcttgacgac gactcggagg acagctgctg cctccgcctg cataaagtgc
4021 tggagcccta caagccccag tggtgccgga gccgcgaggc ctgggccctc tacctcttct
4081 ccccacagaa ccggttccgc gtctcctgcc agaaggtcat cacacacaag atgtttgatc
4141 acgtggtcct cgtcttcatc ttcctcaact gcgtcaccat cgccctggag aggcctgaca
4201 ttgaccccgg cagcaccgag cgggtcttcc tcagcgtctc caattacatc ttcacggcca
4261 tcttcgtggc ggagatgatg gtgaaggtgg tggccctggg gctgctgtcc ggcgagcacg
4321 cctacctgca gagcagctgg aacctgctgg atgggctgct ggtgctggtg tccctggtgg
4381 acattgtcgt ggccatggcc tcggctggtg gcgccaagat cctgggtgtt ctgcgcgtgc
4441 tgcgtctgct gcggaccctg cggcctctaa gggtcatcag ccgggccccg ggcctcaagc
4501 tggtggtgga gacgctgata tcgtcgctca ggcccattgg gaacatcgtc ctcatctgct
4561 gcgccttctt catcattttt ggcatcttgg gtgtgcagct cttcaaaggg aagttctact
4621 actgcgaggg ccccgacacc aggaacatct ccaccaaggc acagtgccgg gccgcccact
4681 accgctgggt gcgacgcaag tacaacttcg acaacctggg ccaggccctg atgtcgctgt
4741 tcgtgctgtc atccaaggat ggatgggtga acatcatgta cgacgggctg gatgccgtgg
4801 gtgtcgacca gcagcctgtg cagaaccaca acccctggat gctgctgtac ttcatctcct
4861 tcctgctcat cgtcagcttc ttcgtgctca acatgttcgt gggcgtcgtg gtcgagaact
4921 tccacaagtg ccggcagcac caggaggcgg aggaggcgcg gcggcgagag gagaagcggc
4981 tgcggcgcct agagaggagg cgcaggagca ctttccccag cccagaggcc cagcgccggc
5041 cctactatgc cgactactcg cccacgcgcc gctccattca ctcgctgtgc accagccact
5101 atctcgacct cttcatcacc ttcatcatct gtgtcaacgt catcaccatg tccatggagc
5161 actataacca acccaagtcg ctggacgagg ccctcaagta ctgcaactac gtcttcacca
5221 tcgtgtttgt cttcgaggct gcactgaagc tggtagcatt tgggttccgt cggttcttca
5281 aggacaggtg gaaccagctg gacctggcca tcgtgctgct gtcactcatg ggcatcacgc
5341 tggaggagat agagatgagc gccgcgctgc ccatcaaccc caccatcatc cgcatcatgc
5401 gcgtgcttcg cattgcccgt gtgctgaagc tgctgaagat ggctacgggc atgcgcgccc
5461 tgctggacac tgtggtgcaa gctctccccc aggtggggaa cctgggcctt cttttcatgc
5521 tcctgttttt tatctatgct gcgctgggag tggagctgtt cgggaggctg gagtgcagtg
5581 aagacaaccc ctgcgagggc ctgagcaggc acgccacctt cagcaacttc ggcatggcct
5641 tcctcacgct gttccgcgtg tccacggggg acaactggaa cgggatcatg aaggacacgc

FIG. 19C

5761 tctacttcgt gaccttcgtg ctggtggccc agttcgtgct ggtgaacgtg gtggtggccg
5821 tgctcatgaa gcacctggag gagagcaaca aggaggcacg ggaggatgcg gagctggacg
5881 ccgagatcga gctggagatg gcgcagggcc ccgggagtgc acgccgggtg gacgcggaca
5941 ggcctccctt gccccaggag agtccgggcg ccagggatgc cccaaacctg gttgcacgca
6001 aggtgtccgt gtccaggatg ctctcgctgc ccaacgacag ctacatgttc aggcccgtgg
6061 tgcctgcctc ggcgccccac ccccgcccgc tgcaggaggt ggagatggag acctatgggg
6121 ccggcacccc cttgggctcc gttgcctctg tgcactctcc gcccgcagag tcctgtgcct
6181 ccctccagat cccactggct gtgtcgtccc cagccaggag cggcgagccc ctccacgccc
6241 tgtcccctcg gggcacagcc cgctccccca gtctcagccg gctgctctgc agacaggagg
6301 ctgtgcacac cgattccttg gaagggaaga ttgacagccc tagggacacc ctggatcctg
6361 cagagcctgg tgagaaaacc ccggtgaggc cggtgaccca ggggggctcc ctgcagtccc
6421 caccacgctc cccacggccc gccagcgtcc gcactcgtaa gcataccttc ggacagcact
6481 gcgtctccag ccggccggcg gccccaggcg gagaggaggc cgaggcctcg gacccagccg
6541 acgaggaggt cagccacatc accagctccg cctgcccctg gcagcccaca gccgagcccc
6601 atggccccga agcctctccg gtggccggcg gcgagcggga cctgcgcagg ctctacagcg
6661 tggacgctca gggcttcctg gacaagccgg gccgggcaga cgagcagtgg cggccctcgg
6721 cggagctggg cagcggggag cctggggagg cgaaggcctg gggccctgag gccgagcccg
6781 ctctgggtgc gcgcagaaag aagaagatga gccccccctg catctcggtg gaacccccctg
6841 cggaggacga gggctctgcg cggccctccg cggcagaggg cggcagcacc acactgaggc
6901 gcaggacccc gtcctgtgag gccacgcctc acagggactc cctggagccc acagagggct
6961 caggcgccgg gggggaccct gcagccaagg gggagcgctg gggccaggcc tcctgccggg
7021 ctgagcacct gaccgtcccc agctttgcct ttgagccgct ggacctcggg gtccccagtg
7081 gagacccttt cttggacggt agccacagtg tgaccccaga atccagagct tcctcttcag
7141 gggccatagt gcccctggaa cccccagaat cagagcctcc catgcccgtc ggtgacccca
7201 cagagaagag gcgggggctg tacctcacag tcccccagtg tcctctggag aaaccagggt
7261 ccccctcagc cacccctgcc ccaggggtg gtgcagatga ccccgtgtag ctcggggctt
7321 ggtgccgccc acggctttgg ccctggggtc tgggggcccc gctggggtgg aggcccaggc
7381 agaaccctgc atggaccctg acttgggtcc cgtcgtgagc agaaaggccc ggggaggatg
7441 acggcccagg ccctggttct ctgcccagcg aagcaggagt agctgccggg ccccacgagc
7501 ctccatccgt tctggttcgg gtttctccga gttttgctac cagccgaggc tgtgcgggca
7561 actgggtcag cctcccgtca ggagagaagc cgcgtctgtg ggacgaagac cgggcacccg
7621 ccagagaggg gaaggtacca ggttgcgtcc tttcaggccc cgcgttgtta caggacactc
7681 gctgggggcc ctgtgccctt gccggcggca ggttgcagcc accgcggccc aatgtcacct
7741 tcactcacag tctgagttct tgtccgcctg tcacgccctc accaccctcc ccttccagcc
7801 accacccttt ccgttccgct cgggccttcc cagaagcgtc ctgtgactct gggagaggtg
7861 acacctcact aaggggccga ccccatggag taacgcgc

FIG. 19D

MTEGARAADEVRVPLGAPPPGPAALVGASPESPGAPGREAERGS
ELGVSPSESPAAERGAELGADEEQRVPYPALAATVFFCLGQTTRPRSWCLRLVCNPWF
EHVSMLVIMLNCVTLGMFRPCEDVECGSERCNILEAFDAFIFAFFAVEMVIKMVALGL
FGQKCYLGDTWNRLDFFIVVAGMMEYSLDGHNVSLSAIRTVRVLRPLRAINRVPSMRI
LVTLLLDTLPMLGNVLLLCFFVFFIFGIVGVQLWAGLLRNRCFLDSAFVRNNNLTFLR
PYYQTEEGEENPFICSSRRDNGMQKCSHIPGRRELRMPCTLGWEAYTQPQAEGVGAAR
NACINWNQYYNVCRSGDSNPHNGAINFDNIGYAWIAIFQVITLEGWVDIMYYVMDAHS
FYNFIYFILLIIVGSFFMINLCLVVIATQFSETKQRESQLMREQRARHLSNDSTLASF
SEPGSCYEELLKYVGHIFRKVKRRSLRLYARWQSRWRKKVDPSAVQGQGPGHRQRRAG
RHTASVHHLVYHHHHHHHHHHYHFSHGSPRRPGPEPGACDTRLVRAGAPPSPPSPGRGP
PDAESVHSIYHADCHIEGPQERARVAHAAATAAASLRLATGLGTMNYPTILPSGVGSG
KGSTSPGPKGKWAGGPPGTGGHGPLSLNSPDPYEKIPHVVGEHGLGQAPGHLSGLSVP
CPLPSPPAGTLTCELKSCPYCTRALEDPEGELSGSESGDSDGRGVYEFTQDVRHGDRW
DPTRPPRATDTPGPGPGSPQRRAQQRAAPGEPGWMGRLWVTFSGKLRRIVDSKYFSRG
IMMAILVNTLSMGVEYHEQPEELTNALEISNIVFTSMFALEMLLKLLACGPLGYIRNP
YNIFDGIIVVISVWEIVGQADGGLSVLRTFRLLRVLKLVRFLPALRRQLVVLVKTMDN
VATFCTLLMLFIFIFSILGMHLFGCKFSLKTDTGDTVPDRKNFDSLLWAIVTVFQILT
QEDWNVVLYNGMASTSSWAALYFVALMTFGNYVLFNLLVAILVEGFQAEGDANRSDTD
EDKTSVHFEEDFHKLRELQTTELKMCSLAVTPNGHLEGRGSLSPPLIMCTAATPMPTP
KSSPFLDAAPSLPDSRRGSSSSGDPPLGDQKPPASLRSSPCAPWGPSGAWSSRRSSWS
SLGRAPSLKRRGQCGERESLLSGEGKGSTDDEAEDGRAAPGPRATPLRRAESLDPRPL
RPAALPPTKCRDRDGQVVALPSDFFLRIDSHREDAAELDDDSEDSCCLRLHKVLEPYK
PQWCRSREAWALYLFSPQNRFRVSCQKVITHKMFDHVVLVFIFLNCVTIALERPDIDP
GSTERVFLSVSNYIFTAIFVAEMMVKVVALGLLSGEHAYLQSSWNLLDGLLVLVSLVD
IVVAMASAGGAKILGVLRVLRLLRTLRPLRVISRAPGLKLVVETLISSLRPIGNIVLI
CCAFFIIFGILGVQLFKGKFYYCEGPDTRNISTKAQCRAAHYRWVRRKYNFDNLGQAL
MSLFVLSSKDGWVNIMYDGLDAVGVDQQPVQNHNPWMLLYFISFLLIVSFFVLNMFVG
VVVENFHKCRQHQEAEEARRREEKRLRRLERRRRSTFPSPEAQRRPYYADYSPTRRSI
HSLCTSHYLDLFITFIICVNVITMSMEHYNQPKSLDEALKYCNYVFTIVFVFEAALKL
VAFGFRRFFKDRWNQLDLAIVLLSLMGITLEEIEMSAALPINPTIIRIMRVLRIARVL
KLLKMATGMRALLDTVVQALPQVGNLGLLFMLLFFIYAALGVELFGRLECSEDNPCEG
LSRHATFSNFGMAFLTLFRVSTGDNWNGIMKDTLRECSREDKHCLSYLPALSPVYFVT
FVLVAQFVLVNVVVAVLMKHLEESNKEAREDAELDAEIELEMAQGPGSARRVDADRPP
LPQESPGARDAPNLVARKVSVSRMLSLPNDSYMFRPVVPASAPHPRPLQEVEMETYGA
GTPLGSVASVHSPPAESCASLQIPLAVSSPARSGEPLHALSPRGTARSPSLSRLLCRQ
EAVHTDSLEGKIDSPRDTLDPAEPGEKTPVRPVTQGGSLQSPPRSPRPASVRTRKHTF
GQHCVSSRPAAPGGEEAEASDPADEEVSHITSSACPWQPTAEPHGPEASPVAGGERDL
RRLYSVDAQGFLDKPGRADEQWRPSAELGSGEPGEAKAWGPEAEPALGARRKKKMSPP
CISVEPPAEDEGSARPSAAEGGSTTLRRRTPSCEATPHRDSLEPTEGSGAGGDPAAKG
ERWGQASCRAEHLTVPSFAFEPLDLGVPSGDPFLDGSHSVTPESRASSSGAIVPLEPP
ESEPPMPVGDPPEKRRGLYLTVPQCPLEKPGSPSATPAPGGGADDPV

FIGURE 20A

1 gcggcggcgg ctgcggcggt ggggccgggc gaggtccgct gcggtcccgg cggctccgtg
61 gctgctccgc tctgagcgcc tggcgcgccc cgcgccctcc ctgccggggc cgctgggccg
121 gggatgcacg cggggcccgg gagccatggt ccgcttcggg gacgagctgg gcggccgcta
181 tggaggcccc ggcggcggag agcgggcccg gggcggcggg gccggcgggg cgggggccc
241 gggtcccggg gggctgcagc ccggccagcg ggtcctctac aagcaatcga tcgcgcagcg
301 cgcgcggacc atggcgctgt acaacccccat cccggtcaag cagaactgct tcaccgtcaa
361 ccgctcgctc ttcgtcttca gcgaggacaa cgtcgtccgc aaatacgcga agcgcatcac
421 cgagtggcct ccattcgagt atatgatcct ggccaccatc atcgccaact gcatcgtgct
481 ggccctggag cagcacctcc ctgatgggga caaaacgccc atgtccgagc ggctggacga
541 cacggagccc tatttcatcg ggatcttttg cttcgaggca gggatcaaaa tcatcgctct
601 gggctttgtc ttccacaagg gctcttacct gcggaacggc tggaacgtca tggacttcgt
661 ggtcgtcctc acagggatcc ttgccacggc tggaactgac ttcgacctgc gaacactgag
721 ggctgtgcgt gtgctgaggc ccctgaagct ggtgtctggg attccaagtt tgcaggtggt
781 gctcaagtcc atcatgaagg ccatggttcc actcctgcag attgggctgc ttctcttctt
841 tgccatcctc atgtttgcca tcattggcct ggagttctac atgggcaagt tccacaaggc
901 ctgtttcccc aacagcacag atgcggagcc cgtgggtgac ttcccctgtg gcaaggaggc
961 cccagcccgg ctgtgcgagg gcgacactga gtgccgggag tactggccag gacccaactt
1021 tggcatcacc aactttgaca atatcctgtt tgccatcttg acggtgttcc agtgcatcac
1081 catggagggc tggactgaca tcctctataa tacaaacgat gcggccggca acacctggaa
1141 ctggctctac ttcatccctc tcatcatcat cggctccttc ttcatgctca acctggtgct
1201 gggcgtgctc tcgggggagt ttgccaagga gcgagagagg gtggagaacc gccgcgcctt
1261 cctgaagctg cgccggcagc agcagatcga gcgagagctc aacgggtacc tggagtggat
1321 cttcaaggcg gaggaagtca tgctggccga ggaggacagg aatgcagagg agaagtcccc
1381 tttggacgtg ctgaagagag cggccaccaa gaagagcaga aatgacctga tccacgcaga
1441 ggagggagag gaccggtttg cagatctctg tgctgttgga tccccccttcg cccgcgccag
1501 cctcaagagc gggaagacag agagctcgtc atacttccgg aggaaggaga agatgttccg
1561 gtttttatc cggcgcatgg tgaaggctca gagcttctac tgggtggtgc tgtgcgtggt
1621 ggccctgaac acactgtgtg tggccatggt gcattacaac cagccgcggc ggcttaccac
1681 gaccctgtat tttgcagagt ttgttttcct gggtctcttc ctcacagaga tgtccctgaa
1741 gatgtatggc ctggggccca gaagctactt ccggtcctcc ttcaactgct tcgactttgg
1801 ggtcatcgtg gggagcgtct ttgaagtggt ctgggcggcc atcaagccgg gaagctcctt
1861 tgggatcagt gtgctgcggg ccctccgcct gctgaggatc ttcaaagtca cgaagtactg
1921 gagctccctg cggaacctgg tggtgtccct gctgaactcc atgaagtcca tcatcagcct
1981 gctcttcttg ctcttcctgt tcattgtggt cttcgccctg ctggggatgc agctgtttgg
2041 gggacagttc aacttccagg atgagactcc cacaaccaac ttcgacacct tccctgccgc
2101 catcctcact gtcttccaga tcctgacggg agaggactgg aatgcagtga tgtatcacgg
2161 gatcgaatcg caaggcggcg tcagcaaagg catgttctcg tcctttact tcattgtcct
2221 gacactgttc ggaaactaca ctctgctgaa tgtctttctg gccatcgctg tggacaacct
2281 ggccaacgcc caagagctga ccaaggatga agaggagatg gaagaagcag ccaatcagaa
2341 gcttgctctg caaaaggcca aagaagtggc tgaagtcagc cccatgtctg ccgcgaacat
2401 ctccatcgcc gccaggcagc agaactcggc caaggcgcgc tcggtgtggg agcagcgggc
2461 cagccagcta cggctgcaga acctgcgggc cagctgcgag gcgctgtaca gcgagatgga
2521 ccccgaggag cggctgcgct tcgccactac gcgccacctg cggcccgaca tgaagacgca
2581 cctggaccgg ccgctggtgg tggagctggg ccgcgacggc gcgcgggggc ccgtgggagg
2641 caaagcccga cctgaggctg cggaggcccc cgagggcgtc gaccctccgc gcaggcacca
2701 ccggcaccgc gacaaggaca agacccccgc ggcgggggac caggaccgag cagaggcccc
2761 gaaggcggag agcggggagc ccggtgcccg ggaggagcgg ccgcggccgc accgcagcca
2821 cagcaaggag gccgcggggc ccccggaggc gcggagcgag cgcggccgag gcccaggccc

FIGURE 20B

2881 cgagggcggc cggcggcacc accggcgcgg ctccccggag gaggcggccg agcgggagcc
2941 ccgacgccac cgcgcgcacc ggcaccagga tccgagcaag gagtgcgccg gcgccaaggg
3001 cgagcggcgc gcgcggcacc gcggcggccc ccgagcgggg ccccgggagg cggagagcgg
3061 ggaggagccg gcgcggcggc accgggcccg gcacaaggcg cagcctgctc acgaggctgt
3121 ggagaaggag accacggaga aggaggccac ggagaaggag gctgagatag tggaagccga
3181 caaggaaaag gagctccgga accaccagcc ccgggagcca cactgtgacc tggagaccag
3241 tgggactgtg actgtgggtc ccatgcacac actgcccagc acctgtctcc agaaggtgga
3301 ggaacagcca gaggatgcag acaatcagcg gaacgtcact cgcatgggca gtcagccccc
3361 agacccgaac actattgtac atatcccagt gatgctgacg ggccctcttg gggaagccac
3421 ggtcgttccc agtggtaacg tggacctgga aagccaagca gaggggaaga aggaggtgga
3481 agcggatgac gtgatgagga gcggcccccg gcctatcgtc ccatacagct ccatgttctg
3541 tttaagcccc accaacctgc tccgccgctt ctgccactac atcgtgacca tgaggtactt
3601 cgaggtggtc attctcgtgg tcatcgcctt gagcagcatc gccctggctg ctgaggaccc
3661 agtgcgcaca gactcgccca ggaacaacgc tctgaaatac ctggattaca ttttcactgg
3721 tgtctttacc tttgagatgg tgataaagat gatcgacttg ggactgctgc ttcaccctgg
3781 agcctatttc cgggacttgt ggaacattct ggacttcatt gtggtcagtg gcgccctggt
3841 ggcgtttgct ttctcaggat ccaaagggaa agacatcaat accatcaagt ctctgagagt
3901 ccttcgtgtc ctgcggcccc tcaagaccat caaacggctg cccaagctca aggctgtgtt
3961 tgactgtgtg gtgaactccc tgaagaatgt cctcaacatc ttgattgtct acatgctctt
4021 catgttcata tttgccgtca ttgcggtgca gctcttcaaa gggaagtttt tctactgcac
4081 agatgaatcc aaggagctgg agagggactg caggggtcag tatttggatt atgagaagga
4141 ggaagtggaa gctcagccca ggcagtggaa gaaatacgac tttcactacg acaatgtgct
4201 ctgggctctg ctgacgctgt tcacagtgtc cacgggagaa ggctggccca tggtgctgaa
4261 acactccgtg gatgccacct atgaggagca gggtccaagc cctgggtacc gcatggagct
4321 gtccatcttc tacgtggtct actttgtggt ctttcccttc ttcttcgtca acatctttgt
4381 ggctttgatc atcatcacct tccaggagca gggggacaag gtgatgtctg aatgcagcct
4441 ggagaagaac gagagggctt gcattgactt cgccatcagc gccaaacccc tgacacggta
4501 catgccccaa aaccggcagt cgttccagta taagacgtgg acatttgtgg tctccccgcc
4561 ctttgaatac ttcatcatgg ccatgatagc cctcaacact gtggtgctga tgatgaagtt
4621 ctatgatgca ccctatgagt acgagctgat gctgaaatgc ctgaacatcg tgttcacatc
4681 catgttctcc atggaatgcg tgctgaagat catcgccttt ggggtgctga actatttcag
4741 agatgcctgg aatgtctttg actttgtcac tgtgttggga agtattactg atattttagt
4801 aacagagatt gcggaaacga acaatttcat caacctcagc ttcctccgcc tctttcgagc
4861 tgcgcggctg atcaagctgc tccgccaggg ctacaccatc cgcatcctgc tgtggacctt
4921 tgtccagtcc ttcaaggccc tgccctacgt gtgtctgctc attgccatgc tgttcttcat
4981 ctacgccatc atcggcatgc aggtgtttgg gaatattgcc ctggatgatg acaccagcat
5041 caaccgccac aacaacttcc ggacgttttt gcaagccctg atgctgctgt tcaggagcgc
5101 cacgggggag gcctggcacg agatcatgct gtcctgcctg agcaaccagg cctgtgatga
5161 gcaggccaat gccaccgagt gtggaagtga ctttgcctac ttctacttcg tctccttcat
5221 cttcctgtgc tcctttctga tgttgaacct ctttgtggct gtgatcatgg acaattttga
5281 gtacctcacg cgggactctt ccatcctagg tcctcaccac ttggatgagt tcatccgggt
5341 ctgggctgaa tacgacccgg ctgcgtgtgg gcgcatcagt tacaatgaca tgtttgagat
5401 gctgaaacac atgtccccgc ctctgggggt ggggaagaaa tgccctgctc gagttgctta
5461 caagcgcctg gttcgcatga acatgcccat ctccaacgag gacatgactg ttcacttcac
5521 gtccacgctg atggccctca tccggacggc actggagatc aagctggccc cagctgggac
5581 aaagcagcat cagtgtgacg cggagttgag gaaggagatt tccgttgtgt gggccaatct

FIG. 20C

5641 gccccagaag actttggact tgctggtacc accccataag cctgatgaga tgacagtggg
5701 gaaggtttat gcagctctga tgatatttga cttctacaag cagaacaaaa ccaccagaga
5761 ccagatgcag caggctcctg gaggcctctc ccagatgggt cctgtgtccc tgttccaccc
5821 tctgaaggcc accctggagc agacacagcc ggctgtgctc cgaggagccc gggttttcct
5881 tcgacagaag agttccacct ccctcagcaa tggcggggcc atacaaaacc aagagagtgg
5941 catcaaagag tctgtctcct ggggcactca aaggacccag gatgcacccc atgaggccag
6001 gccacccctg gagcgtggcc actccacaga gatccctgtg gggcggtcag gagcactggc
6061 tgtggacgtt cagatgcaga gcataacccg gaggggccct gatggggagc cccagcctgg
6121 gctggagagc cagggtcgag cggcctccat gccccgcctt gcggccgaga ctcagcccgt
6181 cacagatgcc agccccatga agcgctccat ctccacgctg gcccagcggc cccgtgggac
6241 tcatctttgc agcaccaccc cggaccgccc acccccctagc caggcgtcgt cgcaccacca
6301 ccaccaccgc tgccaccgcc gcagggacag gaagcagagg tccctggaga aggggcccag
6361 cctgtctgcc gatatggatg gcgcaccaag cagtgctgtg gggccggggc tgcccccggg
6421 agaggggcct acaggctgcc ggcgggaacg agagcgccgg caggagcggg gccggtccca
6481 ggagcggagg cagccctcat cctcctcctc ggagaagcag cgcttctact cctgcgaccg
6541 ctttgggggc cgtgagcccc cgaagcccaa gccctccctc agcagccacc caacgtcgcc
6601 aacagctggc caggagccgg gaccccaccc acagggcagt ggttccgtga atgggagccc
6661 cttgctgtca acatctggtg ctagcacccc cggccgcggt gggcggaggc agctcccccа
6721 gacgcccctg actccccgcc ccagcatcac ctacaagacg gccaactcct cacccatcca
6781 cttcgccggg gctcagacca gcctccctgc cttctcccca ggccggctca gccgtgggct
6841 ttccgaacac aacgccctgc tgcagagaga ccccctcagc cagcccctgg ccctggctc
6901 tcgaattggc tctgacccct acctggggca gcgtctggac agtgaggcct ctgtccacgc
6961 cctgcctgag gacacgctca ctttcgagga ggctgtggcc accaactcgg gccgctcctc
7021 caggacttcc tacgtgtcct ccctgacctc ccagtctcac cctctccgcc gcgtgcccaa
7081 cggttaccac tgcaccctgg gactcagctc gggtggccga gcacggcaca gctaccacca
7141 ccctgaccaa gaccactggt gctagctgca ccgtgaccgc tcagacgcct gcatgcagca
7201 ggcgtgtgtt ccagtggatg agttttatca tccacacggg gcagtcggcc ctcgggggag
7261 gccttgccca ccttggtgag gctcctgtgg cccctccctc cccctcctcc cctcttttac
7321 tctagacgac gaataaagcc ctgttgcttg agtgtacgta ccgc

FIG. 20D

MVRFGDELGGRYGGPGGGERARGGGAGGAGGPGPGGLQPGQRVL
YKQSIAQRARTMALYNPIPVKQNCFTVNRSLFVFSEDNVVRKYAKRITEWPPFEYMIL
ATIIANCIVLALEQHLPDGDKTPMSERLDDTEPYFIGIFCFEAGIKIIALGFVFHKGS
YLRNGWNVMDFVVVLTGILATAGTDFDLRTLRAVRVLRPLKLVSGIPSLQVVLKSIMK
AMVPLLQIGLLLFFAILMFAIIGLEFYMGKFHKACFPNSTDAEPVGDFPCGKEAPARL
CEGDTECREYWPGPNFGITNFDNILFAILTVFQCITMEGWTDILYNTNDAAGNTWNWL
YFIPLIIIGSFFMLNLVLGVLSGEFAKERERVENRRAFLKLRRQQQIERELNGYLEWI
FKAEEVMLAEEDRNAEEKSPLDVLKRAATKKSRNDLIHAEEGEDRFADLCAVGSPFAR
ASLKSGKTESSSYFRRKEKMFRFFIRRMVKAQSFYWVVLCVVALNTLCVAMVHYNQPR
RLTTTLYFAEFVFLGLFLTEMSLKMYGLGPRSYFRSSFNCFDFGVIVGSVFEVVWAAI
KPGSSFGISVLRALRLLRIFKVTKYWSSLRNLVVSLLNSMKSIISLLFLLFLFIVVFA
LLGMQLFGGQFNFQDETPTTNFDTFPAAILTVFQILTGEDWNAVMYHGIESQGGVSKG
MFSSFYFIVLTLFGNYTLLNVFLAIAVDNLANAQELTKDEEEMEEAANQKLALQKAKE
VAEVSPMSAANISIAARQQNSAKARSVWEQRASQLRLQNLRASCEALYSEMDPEERLR
FATTRHLRPDMKTHLDRPLVVELGRDGARGPVGGKARPEAAEAPEGVDPPRRHHRHRD
KDKTPAAGDQDRAEAPKAESGEPGAREERPRPHRSHSKEAAGPPEARSERGRGPGPEG
GRRHHRRGSPEEAAEREPRRHRAHRHQDPSKECAGAKGERRARHRGGPRAGPREAESG
EEPARRHRARHKAQPAHEAVEKETTEKEATEKEAEIVEADKEKELRNHQPREPHCDLE
TSGTVTVGPMHTLPSTCLQKVEEQPEDADNQRNVTRMGSQPPDPNTIVHIPVMLTGPL
GEATVVPSGNVDLESQAEGKKEVEADDVMRSGPRPIVPYSSMFCLSPTNLLRRFCHYI
VTMRYFEVVILVVIALSSIALAAEDPVRTDSPRNNALKYLDYIFTGVFTFEMVIKMID
LGLLLHPGAYFRDLWNILDFIVVSGALVAFAFSGSKGKDINTIKSLRVLRVLRPLKTI
KRLPKLKAVFDCVVNSLKNVLNILIVYMLFMFIFAVIAVQLFKGKFFYCTDESKELER
DCRGQYLDYEKEEVEAQPRQWKKYDFHYDNVLWALLTLFTVSTGEGWPMVLKHSVDAT
YEEQGPSPGYRMELSIFYVVYFVVFPFFFVNIFVALIIITFQEQGDKVMSECSLEKNE
RACIDFAISAKPLTRYMPQNRQSFQYKTWTFVVSPPFEYFIMAMIALNTVVLMMKFYD
APYEYELMLKCLNIVFTSMFSMECVLKIIAFGVLNYFRDAWNVFDFVTVLGSITDILV
TEIAETNNFINLSFLRLFRAARLIKLLRQGYTIRILLWTFVQSFKALPYVCLLIAMLF
FIYAIIGMQVFGNIALDDDTSINRHNNFRTFLQALMLLFRSATGEAWHEIMLSCLSNQ
ACDEQANATECGSDFAYFYFVSFIFLCSFLMLNLFVAVIMDNFEYLTRDSSILGPHHL
DEFIRVWAEYDPAACGRISYNDMFEMLKHMSPPLGLGKKCPARVAYKRLVRMNMPISN
EDMTVHFTSTLMALIRTALEIKLAPAGTKQHQCDAELRKEISVVWANLPQKTLDLLVP
PHKPDEMTVGKVYAALMIFDFYKQNKTTRDQMQQAPGGLSQMGPVSLFHPLKATLEQT
QPAVLRGARVFLRQKSSTSLSNGGAIQNQESGIKESVSWGTQRTQDAPHEARPPLERG
HSTEIPVGRSGALAVDVQMQSITRRGPDGEPQPGLESQGRAASMPRLAAETQPVTDAS
PMKRSISTLAQRPRGTHLCSTTPDRPPPSQASSHHHHHRCHRRRDRKQRSLEKGPSLS
ADMDGAPSSAVGPGLPPGEGPTGCRRERERRQERGRSQERRQPSSSSSEKQRFYSCDR
FGGREPPKPKPSLSSHPTSPTAGQEPGPHPQGSGSVNGSPLLSTSGASTPGRGGRRQL
PQTPLTPRPSITYKTANSSPIHFAGAQTSLPAFSPGRLSRGLSEHNALLQRDPLSQPL
APGSRIGSDPYLGQRLDSEASVHALPEDTLTFEEAVATNSGRSSRTSYVSSLTSQSHP
LRRVPNGYHCTLGLSSGGRARHSYHHPDQDHWC

FIGURE 21A

1 gcggcggcgg ctgcggcggt ggggccgggc gaggtccgct gcggtcccgg cggctccgtg
61 gctgctccgc tctgagcgcc tggcgcgccc cgcgccctcc ctgccggggc cgctgggccg
121 gggatgcacg cggggcccgg gagccatggt ccgcttcggg gacgagctgg gcggccgcta
181 tggaggcccc ggcggcggag agcgggcccg gggcggcggg gccggcgggg cgggggccc
241 gggtcccggg gggctgcagc ccggccagcg ggtcctctac aagcaatcga tcgcgcagcg
301 cgcgcggacc atggcgctgt acaaccccat cccggtcaag cagaactgct tcaccgtcaa
361 ccgctcgctc ttcgtcttca gcgaggacaa cgtcgtccgc aaatacgcga agcgcatcac
421 cgagtggcct ccattcgagt atatgatcct ggccaccatc atcgccaact gcatcgtgct
481 ggccctggag cagcacctcc ctgatgggga caaaacgccc atgtccgagc ggctggacga
541 cacggagccc tatttcatcg ggatcttttg cttcgaggca gggatcaaaa tcatcgctct
601 gggctttgtc ttccacaagg gctcttacct gcggaacggc tggaacgtca tggacttcgt
661 ggtcgtcctc acagggatcc ttgccacggc tggaactgac ttcgacctgc gaacactgag
721 ggctgtgcgt gtgctgaggc ccctgaagct ggtgtctggg attccaagtt tgcaggtggt
781 gctcaagtcc atcatgaagg ccatggttcc actcctgcag attgggctgc ttctcttctt
841 tgccatcctc atgtttgcca tcattggcct ggagttctac atgggcaagt tccacaaggc
901 ctgtttcccc aacagcacag atgcggagcc cgtgggtgac ttcccctgtg gcaaggaggc
961 cccagcccgg ctgtgcgagg gcgacactga gtgccgggag tactggccag gacccaactt
1021 tggcatcacc aactttgaca atatcctgtt tgccatcttg acggtgttcc agtgcatcac
1081 catggagggc tggactgaca tcctctataa tacaaaacgat gcggccggca acacctggaa
1141 ctggctctac ttcatccctc tcatcatcat cggctccttc ttcatgctca acctggtgct
1201 gggcgtgctc tcgggggagt ttgccaagga gcgagagagg gtggagaacc gccgcgcctt
1261 cctgaagctg cgccggcagc agcagatcga gcgagagctc aacgggtacc tggagtggat
1321 cttcaaggcg gaggaagtca tgctggccga ggaggacagg aatgcagagg agaagtcccc
1381 tttggacgtg ctgaagagag cggccaccaa gaagagcaga aatgacctga tccacgcaga
1441 ggagggagag gaccggtttg cagatctctg tgctgttgga tccccccttcg cccgcgccag
1501 cctcaagagc gggaagacag agagctcgtc atacttccgg aggaaggaga agatgttccg
1561 gtttttttatc cggcgcatgg tgaaggctca gagcttctac tgggtggtgc tgtgcgtggt
1621 ggccctgaac acactgtgtg tggccatggt gcattacaac cagccgcggc ggcttaccac
1681 gaccctgtat tttgcagagt ttgttttcct gggtctcttc ctcacagaga tgtccctgaa
1741 gatgtatggc ctggggccca gaagctactt ccggtcctcc ttcaactgct tcgactttgg
1801 ggtcatcgtg gggagcgtct ttgaagtggt ctgggcggcc atcaagccgg gaagctcctt
1861 tgggatcagt gtgctgcggg ccctccgcct gctgaggatc ttcaaagtca cgaagtactg
1921 gagctccctg cggaacctgg tggtgtccct gctgaactcc atgaagtcca tcatcagcct
1981 gctcttcttg ctcttcctgt tcattgtggt cttcgccctg ctggggatgc agctgtttgg
2041 gggacagttc aacttccagg atgagactcc cacaaccaac ttcgacacct tccctgccgc
2101 catcctcact gtcttccaga tcctgacggg agaggactgg aatgcagtga tgtatcacgg
2161 gatcgaatcg caaggcggcg tcagcaaagg catgttctcg tcctttact tcattgtcct
2221 gacactgttc ggaaactaca ctctgctgaa tgtctttctg gccatcgctg tggacaacct
2281 ggccaacgcc caagagctga ccaaggatga agaggagatg gaagaagcag ccaatcagaa
2341 gcttgctctg caaaaggcca aagaagtggc tgaagtcagc cccatgtctg ccgcgaacat
2401 ctccatcgcc gccaggcagc agaactcggc caaggcgcgc tcggtgtggg agcagcgggc
2461 cagccagcta cggctgcaga acctgcgggc cagctgcgag gcgctgtaca gcgagatgga
2521 ccccgaggag cggctgcgct tcgccactac gcgccacctg cggcccgaca tgaagacgca
2581 cctggaccgg ccgctggtgg tggagctggg ccgcgacggc gcgcgggggc ccgtgggagg
2641 caaagcccga cctgaggctg cggaggcccc cgagggcgtc gaccctccgc gcaggcacca
2701 ccggcaccgc gacaaggaca agacccccgc ggcgggggac caggaccgag cagaggcccc
2761 gaaggcggag agcggggagc ccggtgcccg ggaggagcgg ccgcggccgc accgcagcca

FIGURE 21B

2821 cagcaaggag gccgcggggc ccccggaggc gcggagcgag cgcggccgag gcccaggccc
2881 cgagggcggc cggcggcacc accggcgcgg ctccccggag gaggcggccg agcgggagcc
2941 ccgacgccac cgcgcgcacc ggcaccagga tccgagcaag gagtgcgccg gcgccaaggg
3001 cgagcggcgc gcgcggcacc gcggcggccc ccgagcgggg ccccgggagg cggagagcgg
3061 ggaggagccg gcgcggcggc accgggcccg gcacaaggcg cagcctgctc acgaggctgt
3121 ggagaaggag accacggaga aggaggccac ggagaaggag gctgagatag tggaagccga
3181 caaggaaaag gagctccgga accaccagcc ccgggagcca cactgtgacc tggagaccag
3241 tgggactgtg actgtgggtc ccatgcacac actgcccagc acctgtctcc agaaggtgga
3301 ggaacagcca gaggatgcag acaatcagcg gaacgtcact cgcatgggca gtcagccccc
3361 agacccgaac actattgtac atatcccagt gatgctgacg ggcccctcttg gggaagccac
3421 ggtcgttccc agtggtaacg tggacctgga aagccaagca gaggggaaga aggaggtgga
3481 agcggatgac gtgatgagga gcggcccccg gcctatcgtc ccatacagct ccatgttctg
3541 tttaagcccc accaacctgc tccgccgctt ctgccactac atcgtgacca tgaggtactt
3601 cgaggtggtc attctcgtgg tcatcgcctt gagcagcatc gccctggctg ctgaggaccc
3661 agtgcgcaca gactcgccca ggaacaacgc tctgaaatac ctggattaca ttttcactgg
3721 tgtctttacc tttgagatgg tgataaagat gatcgacttg ggactgctgc ttcaccctgg
3781 agcctatttc cgggacttgt ggaacattct ggacttcatt gtggtcagtg gcgccctggt
3841 ggcgtttgct ttctcaggat ccaaagggaa agacatcaat accatcaagt ctctgagagt
3901 ccttcgtgtc ctgcggcccc tcaagaccat caaacggctg cccaagctca aggctgtgtt
3961 tgactgtgtg gtgaactccc tgaagaatgt cctcaacatc ttgattgtct acatgctctt
4021 catgttcata tttgccgtca ttgcggtgca gctcttcaaa gggaagtttt tctactgcac
4081 agatgaatcc aaggagctgg agagggactg caggggtcag tatttggatt atgagaagga
4141 ggaagtggaa gctcagccca ggcagtggaa gaaatacgac tttcactacg acaatgtgct
4201 ctgggctctg ctgacgctgt tcacagtgtc cacgggagaa ggctggccca tggtgctgaa
4261 acactccgtg gatgccacct atgaggagca gggtccaagc cctgggtacc gcatggagct
4321 gtccatcttc tacgtggtct actttgtggt ctttcccttc ttcttcgtca acatctttgt
4381 ggctttgatc atcatcacct tccaggagca gggggacaag gtgatgtctg aatgcagcct
4441 ggagaagaac gagagggctt gcattgactt cgccatcagc gccaaacccc tgacacggta
4501 catgccccaa aaccggcagt cgttccagta taagacgtgg acatttgtgg tctccccgcc
4561 ctttgaatac ttcatcatgg ccatgatagc cctcaacact gtggtgctga tgatgaagtt
4621 ctatgatgca ccctatgagt acgagctgat gctgaaatgc ctgaacatcg tgttcacatc
4681 catgttctcc atggaatgcg tgctgaagat catcgccttt ggggtgctga actatttcag
4741 agatgcctgg aatgtctttg actttgtcac tgtgttggga agtattactg atattttagt
4801 aacagagatt gcggaaacga acaatttcat caacctcagc ttcctccgcc tctttcgagc
4861 tgcgcggctg atcaagctgc tccgccaggg ctacaccatc cgcatcctgc tgtggacctt
4921 tgtccagtcc ttcaaggccc tgccctacgt gtgtctgctc attgccatgc tgttcttcat
4981 ctacgccatc atcggcatgc aggtgtttgg gaatattgcc ctggatgatg acaccagcat
5041 caaccgccac aacaacttcc ggacgttttt gcaagccctg atgctgctgt tcaggagcgc
5101 cacgggggag gcctggcacg agatcatgct gtcctgcctg agcaaccagg cctgtgatga
5161 gcaggccaat gccaccgagt gtggaagtga ctttgcctac ttctacttcg tctccttcat
5221 cttcctgtgc tcctttctga tgttgaacct ctttgtggct gtgatcatgg acaattttga
5281 gtacctcacg cgggactctt ccatcctagg tcctcaccac ttggatgagt tcatccgggt
5341 ctgggctgaa tacgacccgg ctgcgtgtgg gcgcatcagt tacaatgaca tgtttgagat
5401 gctgaaacac atgtccccgc ctctggggct ggggaagaaa tgccctgctc gagttgctta
5461 caagcgcctg gttcgcatga acatgcccat ctccaacgag gacatgactg ttcacttcac
5521 gtccacgctg atggccctca tccggacggc actggagatc aagctggccc cagctgggac

FIG. 21C

5581 aaagcagcat cagtgtgacg cggagttgag gaaggagatt tccgttgtgt gggccaatct
5641 gccccagaag actttggact tgctggtacc accccataag cctgatgaga tgacagtggg
5701 gaaggtttat gcagctctga tgatatttga cttctacaag cagaacaaaa ccaccagaga
5761 ccagatgcag caggctcctg gaggcctctc ccagatgggt cctgtgtccc tgttccaccc
5821 tctgaaggcc accctggagc agacacagcc ggctgtgctc cgaggagccc gggttttcct
5881 tcgacagaag agttccacct ccctcagcaa tggcggggcc atacaaaacc aagagagtgg
5941 catcaaagag tctgtctcct ggggcactca aaggacccag gatgcacccc atgaggccag
6001 gccacccctg gagcgtggcc actccacaga gatccctgtg gggcggtcag gagcactggc
6061 tgtggacgtt cagatgcaga gcataacccg gaggggccct gatggggagc cccagcctgg
6121 gctggagagc cagggtcgag cggcctccat gccccgcctt gcggccgaga ctcagcccgt
6181 cacagatgcc agccccatga agcgctccat ctccacgctg gcccagcggc cccgtgggac
6241 tcatctttgc agcaccaccc cggaccgccc acccccctagc caggcgtcgt cgcaccacca
6301 ccaccaccgc tgccaccgcc gcagggacag gaagcagagg tccctggaga aggggcccag
6361 cctgtctgcc gatatggatg gcgcaccaag cagtgctgtg gggccggggc tgcccccggg
6421 agaggggcct acaggctgcc ggcgggaacg agagcgccgg caggagcggg gccggtccca
6481 ggagcggagg cagccctcat cctcctcctc ggagaagcag cgcttctact cctgcgaccg
6541 ctttgggggc cgtgagcccc cgaagcccaa gccctccctc agcagccacc caacgtcgcc
6601 aacagctggc caggagccgg gaccccaccc acaggccggc tcagccgtgg gctttccgaa
6661 cacaacgccc tgctgcagag agacccccte agccagcccc tggcccctgg ctctcgaatt
6721 ggctctgacc cttacctggg gcagcgtctg gacagtgagg cctctgtcca cgccctgcct
6781 gaggacacgc tcactttcga ggaggctgtg gccaccaact cgggccgctc ctccaggact
6841 tcctacgtgt cctccctgac ctcccagtct caccctctcc gccgcgtgcc caacggttac
6901 cactgcaccc tgggactcag ctcgggtggc cgagcacggc acagctacca ccaccctgac
6961 caagaccact ggtgctagct gcaccgtgac cgctcagacg cctgcatgca gcaggcgtgt
7021 gttccagtgg atgagtttta tcatccacac ggggcagtcg gccctcgggg gaggccttgc
7081 ccaccttggt gaggctcctg tggcccctcc ctcccccctcc tcccctcttt tactctagac
7141 gacgaataaa gccctgttgc ttgagtgtac gtaccgc

FIG. 21D

MVRFGDELGGRYGGPGGGERARGGGAGGAGGPGPGGLQPGQRVL
YKQSIAQRARTMALYNPIPVKQNCFTVNRSLFVFSEDNVVRKYAKRITEWPPFEYMIL
ATIIANCIVLALEQHLPDGDKTPMSERLDDTEPYFIGIFCFEAGIKIIALGFVFHKGS
YLRNGWNVMDFVVVLTGILATAGTDFDLRTLRAVRVLRPLKLVSGIPSLQVVLKSIMK
AMVPLLQIGLLLFFAILMFAIIGLEFYMGKFHKACFPNSTDAEPVGDFPCGKEAPARL
CEGDTECREYWPGPNFGITNFDNILFAILTVFQCITMEGWTDILYNTNDAAGNTWNWL
YFIPLIIIGSFFMLNLVLGVLSGEFAKERERVENRRAFLKLRRQQQIERELNGYLEWI
FKAEEVMLAEEDRNAEEKSPLDVLKRAATKKSRNDLIHAEEGEDRFADLCAVGSPFAR
ASLKSGKTESSSYFRRKEKMFRFFIRRMVKAQSFYWVVLCVVALNTLCVAMVHYNQPR
RLTTTLYFAEFVFLGLFLTEMSLKMYGLGPRSYFRSSFNCFDFGVIVGSVFEVVWAAI
KPGSSFGISVLRALRLLRIFKVTKYWSSLRNLVVSLLNSMKSIISLLFLLFLFIVVFA
LLGMQLFGGQFNFQDETPTTNFDTFPAAILTVFQILTGEDWNAVMYHGIESQGGVSKG
MFSSFYFIVLTLFGNYTLLNVFLAIAVDNLANAQELTKDEEEMEEAANQKLALQKAKE
VAEVSPMSAANISIAARQQNSAKARSVWEQRASQLRLQNLRASCEALYSEMDPEERLR
FATTRHLRPDMKTHLDRPLVVELGRDGARGPVGGKARPEAAEAPEGVDPPRRHHRHRD
KDKTPAAGDQDRAEAPKAESGEPGAREERPRPHRSHSKEAAGPPEARSERGRGPGPEG
GRRHHRRGSPEEAAEREPRRHRAHRHQDPSKECAGAKGERRARHRGGPRAGPREAESG
EEPARRHRARHKAQPAHEAVEKETTEKEATEKEAEIVEADKEKELRNHQPREPHCDLE
TSGTVTVGPMHTLPSTCLQKVEEQPEDADNQRNVTRMGSQPPDPNTIVHIPVMLTGPL
GEATVVPSGNVDLESQAEGKKEVEADDVMRSGPRPIVPYSSMFCLSPTNLLRRFCHYI
VTMRYFEVVILVVIALSSIALAAEDPVRTDSPRNNALKYLDYIFTGVFTFEMVIKMID
LGLLLHPGAYFRDLWNILDFIVVSGALVAFAFSGSKGKDINTIKSLRVLRVLRPLKTI
KRLPKLKAVFDCVVNSLKNVLNILIVYMLFMFIFAVIAVQLFKGKFFYCTDESKELER
DCRGQYLDYEKEEVEAQPRQWKKYDFHYDNVLWALLTLFTVSTGEGWPMVLKHSVDAT
YEEQGPSPGYRMELSIFYVVYFVVFPFFFVNIFVALIIITFQEQGDKVMSECSLEKNE
RACIDFAISAKPLTRYMPQNRQSFQYKTWTFVVSPPFEYFIMAMIALNTVVLMMKFYD
APYEYELMLKCLNIVFTSMFSMECVLKIIAFGVLNYFRDAWNVFDFVTVLGSITDILV
TEIAETNNFINLSFLRLFRAARLIKLLRQGYTIRILLWTFVQSFKALPYVCLLIAMLF
FIYAIIGMQVFGNIALDDDTSINRHNNFRTFLQALMLLFRSATGEAWHEIMLSCLSNQ
ACDEQANATECGSDFAYFYFVSFIFLCSFLMLNLFVAVIMDNFEYLTRDSSILGPHHL
DEFIRVWAEYDPAACGRISYNDMFEMLKHMSPPLGLGKKCPARVAYKRLVRMNMPISN
EDMTVHFTSTLMALIRTALEIKLAPAGTKQHQCDAELRKEISVVWANLPQKTLDLLVP
PHKPDEMTVGKVYAALMIFDFYKQNKTTRDQMQQAPGGLSQMGPVSLFHPLKATLEQT
QPAVLRGARVFLRQKSSTSLSNGGAIQNQESGIKESVSWGTQRTQDAPHEARPPLERG
HSTEIPVGRSGALAVDVQMQSITRRGPDGEPQPGLESQGRAASMPRLAAETQPVTDAS
PMKRSISTLAQRPRGTHLCSTTPDRPPPSQASSHHHHHRCHRRRDRKQRSLEKGPSLS
ADMDGAPSSAVGPGLPPGEGPTGCRRERERRQERGRSQERRQPSSSSSEKQRFYSCDR
FGGREPPKPKPSLSSHPTSPTAGQEPGPHPQAGSAVGFPNTTPCCRETPSASPWPLAL
ELALTLTWGSVWTVRPLSTPCLRTRSLSRRLWPPTRAAPPGLPTCPP

FIGURE 22A

1 gatgtcccga gctgctatcc ccggctcggc ccgggcagcc gccttctgag cccccgaccc
61 gaggcgccga gccgccgccg cccgatgggc tgggccgtgg agcgtctccg cagtcgtagc
121 tccagccgcc gcgctcccag ccccggcagc ctcagcatca gcggcggcgg cggcggcggc
181 ggcgtcttcc gcatcgttcg ccgcagcgta acccggagcc ctttgctctt tgcagaatgg
241 cccgcttcgg agacgagatg ccggcccgct acgggggagg aggctccggg gcagccgccg
301 gggtggtcgt gggcagcgga ggcgggcgag gagccggggg cagccggcag ggcgggcagc
361 ccggggcgca aaggatgtac aagcagtcaa tggcgcagag agcgcggacc atggcactct
421 acaaccccat ccccgtccga cagaactgcc tcacggttaa ccggtctctc ttcctcttca
481 gcgaagacaa cgtggtgaga aaatacgcca aaaagatcac cgaatggcct ccctttgaat
541 atatgatttt agccaccatc atagcgaatt gcatcgtcct cgcactggag cagcatctgc
601 ctgatgatga caagaccccg atgtctgaac ggctggatga cacagaacca tacttcattg
661 gaatttttg tttcgaggct ggaattaaaa tcattgccct tgggtttgcc ttccacaaag
721 gctcctactt gaggaatggc tggaatgtca tggactttgt ggtggtgcta acgggcatct
781 tggcgacagt tgggacggag tttgacctac ggacgctgag ggcagttcga gtgctgcggc
841 cgctcaagct ggtgtctgga atcccaagtt tacaagtcgt cctgaagtcg atcatgaagg
901 cgatgatccc tttgctgcag atcggcctcc tcctattttt tgcaatcctt atttttgcaa
961 tcatagggtt agaatttat atgggaaaat ttcataccac ctgctttgaa gaggggacag
1021 atgacattca gggtgagtct ccggctccat gtgggacaga agagcccgcc cgcacctgcc
1081 ccaatgggac caaatgtcag ccctactggg aagggcccaa caacgggatc actcagttcg
1141 acaacatcct gtttgcagtg ctgactgttt tccagtgcat aaccatggaa gggtggactg
1201 atctcctcta caatagcaac gatgcctcag ggaacacttg gaactggttg tacttcatcc
1261 ccctcatcat catcggctcc ttttttatgc tgaaccttgt gctgggtgtg ctgtcagggg
1321 agtttgccaa agaaagggaa cgggtggaga accggcgggc ttttctgaag ctgaggcggc
1381 aacaacagat tgaacgtgag ctcaatgggt acatggaatg gatctcaaaa gcagaagagg
1441 tgatcctcgc cgaggatgaa actgacgggg agcagaggca tccctttgat ggagctctgc
1501 ggagaaccac ataaagaaa agcaagacag atttgctcaa ccccgaagag gctgaggatc
1561 agctggctga tatagcctct gtgggttctc ccttcgcccg agccagcatt aaaagtgcca
1621 agctggagaa ctcgaccttt tttcacaaaa aggagaggag gatgcgtttc tacatccgcc
1681 gcatggtcaa aactcaggcc ttctactgga ctgtactcag tttggtagct ctcaacacgc
1741 tgtgtgttgc tattgttcac tacaaccagc ccgagtggct ctccgacttc ctttactatg
1801 cagaattcat tttcttagga ctctttatgt ccgaaatgtt tataaaaatg tacgggcttg
1861 ggacgcggcc ttacttccac tcttccttca actgctttga ctgtggggtt atcattggga
1921 gcatcttcga ggtcatctgg gctgtcataa aacctggcac atcctttgga atcagcgtgt
1981 tacgagccct caggttattg cgtatttttca aagtcacaaa gtactgggca tctctcagaa
2041 acctggtcgt ctctctcctc aactccatga agtccatcat cagcctgttg tttctccttt
2101 tcctgttcat tgtcgtcttc gccctttttgg gaatgcaact cttcggcggc cagtttaatt
2161 tcgatgaagg gactcctccc accaacttcg atacttttcc agcagcaata atgacggtgt
2221 ttcagatcct gacgggcgaa gactggaacg aggtcatgta cgacgggatc aagtctcagg
2281 ggggcgtgca gggcggcatg gtgttctcca tctatttcat tgtactgacg ctctttggga
2341 actacaccct cctgaatgtg ttcttggcca tcgctgtgga caatctggcc aacgcccagg
2401 agctcaccaa ggtggaggcg gacgagcaag aggaagaaga agcagcgaac cagaaacttg
2461 ccctacagaa agccaaggag gtggcagaag tgagtcctct gtccgcggcc aacatgtcta
2521 tagctgtgaa agagcaacag aagaatcaaa agccagccaa gtccgtgtgg gagcagcgga
2581 ccagtgagat gcgaaagcag aacttgctgg ccagccggga ggccctgtat aacgaaatgg
2641 acccggacga gcgctggaag gctgcctaca cgcggcacct gcggccagac atgaagacgc

FIGURE 22B

2701 acttggaccg gccgctggtg gtggacccgc aggagaaccg caacaacaac accaacaaga
2761 gccgggcggc cgagcccacc gtggaccagc gcctcggcca gcagcgcgcc gaggacttcc
2821 tcaggaaaca ggcccgctac cacgatcggg cccgggaccc cagcggctcg gcgggcctgg
2881 acgcacggag gccctgggcg ggaagccagg aggccgagct gagccgggag ggaccctacg
2941 gccgcgagtc ggaccaccac gcccgggagg gcagcctgga gcaacccggg ttctgggagg
3001 gcgaggccga gcgaggcaag gccgggggacc cccaccggag gcacgtgcac cggcaggggg
3061 gcagcaggga gagccgcagc gggtccccgc gcacgggcgc ggacggggag catcgacgtc
3121 atcgcgcgca ccgcaggccc ggggaggagg gtccggagga caaggcggag cggagggcgc
3181 ggcaccgcga gggcagccgg ccggcccggg gcggcgaggg cgagggcgag ggccccgacg
3241 ggggcgagcg caggagaagg caccggcatg gcgctccagc cacgtacgag ggggacgcgc
3301 ggagggagga caaggagcgg aggcatcgga ggaggaaaga gaaccagggc tccggggtcc
3361 ctgtgtcggg ccccaacctg tcaaccaccc ggccaatcca gcaggacctg ggccgccaag
3421 acccaccccc ggcagaggat attgacaaca tgaagaacaa caagctggcc accgcggagt
3481 cggccgctcc ccacggcagc cttggccacg ccggcctgcc ccagagccca gccaagatgg
3541 gaaacagcac cgaccccggc cccatgctgg ccatccctgc catggccacc aacccccaga
3601 acgccgccag ccgccggacg cccaacaacc cggggaaccc atccaatccc ggccccccca
3661 agacccccga gaatagcctt atcgtcacca accccagcgg cacccagacc aattcagcta
3721 agactgccag gaaacccgac cacaccacag tggacatccc cccagcctgc ccaccccccc
3781 tcaaccacac cgtcgtacaa gtgaacaaaa acgccaaccc agacccactg ccaaaaaaag
3841 aggaagagaa gaaggaggag gaggaagacg accgtgggga agacggccct aagccaatgc
3901 ctccctatag ctccatgttc atcctgtcca cgaccaaccc ccttcgccgc ctgtgccatt
3961 acatcctgaa cctgcgctac tttgagatgt gcatcctcat ggtcattgcc atgagcagca
4021 tcgccctggc cgccgaggac cctgtgcagc ccaacgcacc tcggaacaac gtgctgcgat
4081 actttgacta cgtttttaca ggcgtcttca cctttgagat ggtgatcaag atgattgacc
4141 tggggctcgt cctgcatcag ggtgcctact tccgtgacct ctggaatatt ctcgacttca
4201 tagtggtcag tggggccctg gtagcctttg ccttcactgg caatagcaaa ggaaaagaca
4261 tcaacacgat taaatccctc cgagtcctcc gggtgctacg acctcttaaa accatcaagc
4321 ggctgccaaa gctcaaggct gtgtttgact gtgtggtgaa ctcacttaaa aacgtcttca
4381 acatcctcat cgtctacatg ctattcatgt tcatcttcgc cgtggtggct gtgcagctct
4441 tcaaggggaa attcttccac tgcactgacg agtccaaaga gtttgagaaa gattgtcgag
4501 gcaaatacct cctctacgag aagaatgagg tgaaggcgcg agaccgggag tggaagaagt
4561 atgaattcca ttacgacaat gtgctgtggg ctctgctgac cctcttcacc gtgtccacgg
4621 gagaaggctg gccacaggtc ctcaagcatt cggtggacgc caccttgag aaccagggcc
4681 ccagccccgg gtaccgcatg gagatgtcca ttttctacgt cgtctacttt gtggtgttcc
4741 ccttcttctt tgtcaatatc tttgtggcct tgatcatcat caccttccag gagcaagggg
4801 acaagatgat ggaggaatac agcctggaga aaaatgagag ggcctgcatt gatttcgcca
4861 tcagcgccaa gccgctgacc cgacacatgc cgcagaacaa gcagagcttc cagtaccgca
4921 tgtggcagtt cgtggtgtct ccgcctttcg agtacacgat catggccatg atcgccctca
4981 acaccatcgt gcttatgatg aagttctatg gggcttctgt tgcttatgaa aatgccctgc
5041 gggtgttcaa catcgtcttc acctccctct tctctctgga atgtgtgctg aaagtcatgg
5101 cttttgggat tctgaattat ttccgcgatg cctggaacat cttcgacttt gtgactgttc
5161 tgggcagcat caccgatatc ctcgtgactg agtttgggaa tccgaataac ttcatcaacc
5221 tgagctttct ccgcctcttc cgagctgccc ggctcatcaa acttctccgt cagggttaca
5281 ccatccgcat tcttctctgg acctttgtgc agtccttcaa ggccctgcct tatgtctgtc
5341 tgctgatcgc catgctcttc ttcatctatg ccatcattgg gatgcaggtg tttggtaaca

FIG. 22C

5401 ttggcatcga cgtggaggac gaggacagtg atgaagatga gttccaaatc actgagcaca
5461 ataacttccg gaccttcttc caggccctca tgcttctctt ccggagtgcc accggggaag
5521 cttggcacaa catcatgctt tcctgcctca gcgggaaacc gtgtgataag aactctggca
5581 tcctgactcg agagtgtggc aatgaatttg cttattttta ctttgtttcc ttcatcttcc
5641 tctgctcgtt tctgatgctg aatctctttg tcgccgtcat catggacaac tttgagtacc
5701 tcacccgaga ctcctccatc ctgggccccc accacctgga tgagtacgtg cgtgtctggg
5761 ccgagtatga ccccgcagct tggggccgca tgccttacct ggacatgtat cagatgctga
5821 gacacatgtc tccgcccctg ggtctgggga agaagtgtcc ggccagagtg gcttacaagc
5881 ggcttctgcg gatggacctg cccgtcgcag atgacaacac cgtccacttc aattccaccc
5941 tcatggctct gatccgcaca gccctggaca tcaagattgc caagggagga gccgacaaac
6001 agcagatgga cgctgagctg cggaaggaga tgatggcgat ttggcccaat ctgtcccaga
6061 agacgctaga cctgctggtc acacctcaca agtccacgga cctcaccgtg gggaagatct
6121 acgcagccat gatgatcatg gagtactacc ggcagagcaa ggccaagaag ctgcaggcca
6181 tgcgcgagga gcaggaccgg acacccctca tgttccagcg catggagccc ccgtccccaa
6241 cgcaggaagg gggacctggc cagaacgccc tcccctccac ccagctggac ccaggaggag
6301 ccctgatggc tcacgaaagc ggcctcaagg agagcccgtc ctgggtgacc cagcgtgccc
6361 aggagatgtt ccagaagacg ggcacatgga gtccggaaca aggcccccct accgacatgc
6421 ccaacagcca gcctaactct cagtccgtgg agatgcgaga gatgggcaga gatggctact
6481 ccgacagcga gcactacctc cccatggaag gccagggccg ggctgcctcc atgccccgcc
6541 tccctgcaga gaaccagagg agaaggggcc ggccacgtgg gaataacctc agtaccatct
6601 cagacaccag ccccatgaag cgttcagcct ccgtgctggg ccccaaggcc cgacgcctgg
6661 acgattactc gctggagcgg gtcccgcccg aggagaacca gcggcaccac cagcggcgcc
6721 gcgaccgcag ccaccgcgcc tctgagcgct ccctgggccg ctacaccgat gtggacacag
6781 gcttggggac agacctgagc atgaccaccc aatccgggga cctgccgtcg aaggagcggg
6841 accaggagcg gggccggccc aaggatcgga agcatcgaca gcaccaccac caccaccacc
6901 accaccacca tccccccgcc cccgacaagg accgctatgc ccaggaacgg ccggaccacg
6961 gccgggcacg ggctcgggac cagcgctggt cccgctcgcc cagcgagggc cgagagcaca
7021 tggcgcaccg gcagggcagt agttccgtaa gtggaagccc agccccctca acatctggta
7081 ccagcactcc gcggcggggc cgccgccagc tcccccagac cccctccacc ccccggccac
7141 acgtgtccta ttcccctgtg atccgtaagg ccggcggctc ggggcccccg cagcagcagc
7201 agcagcagca gcagcagcag caggcggtgg ccaggccggg ccgggcggcc accagcggcc
7261 ctcggaggta cccaggcccc acggccgagc ctctggccgg agatcggccg cccacggggg
7321 gccacagcag cggccgctcg cccaggatgg agaggcgggt cccaggcccg gcccggagcg
7381 agtcccccag ggcctgtcga cacggcgggg cccggtggcc ggcatctggc ccgcacgtgt
7441 ccgaggggcc cccgggtccc cggcaccatg gctactaccg gggctccgac tacgacgagg
7501 ccgatggccc gggcagcggg ggcggcgagg aggccatggc cggggcctac gacgcgccac
7561 cccccgtacg acacgcgtcc tcgggcgcca ccgggcgctc gcccaggact ccccgggcct
7621 cgggcccggc ctgcgcctcg ccttctcggc acggccggcg actccccaac ggctactacc
7681 cggcgcacgg actggccagg ccccgcgggc cgggctccag gaagggcctg cacgaaccct
7741 acagcgagag tgacgatgat tggtgctaag cccgggcgag gtggcgcccg cccggccccc
7801 cacgcacc

FIGURE 22D

MARFGDEMPARYGGGGSGAAAGVVVGSGGGRGAGGSRQGGQPGA
QRMYKQSMAQRARTMALYNPIPVRQNCLTVNRSLFLFSEDNVVRKYAKKITEWPPFEY
MILATIIANCIVLALEQHLPDDDKTPMSERLDDTEPYFIGIFCFEAGIKIIALGFAFH
KGSYLRNGWNVMDFVVVLTGILATVGTEFDLRTLRAVRVLRPLKLVSGIPSLQVVLKS
IMKAMIPLLQIGLLLFFAILIFAIIGLEFYMGKFHTTCFEEGTDDIQGESPAPCGTEE
PARTCPNGTKCQPYWEGPNNGITQFDNILFAVLTVFQCITMEGWTDLLYNSNDASGNT
WNWLYFIPLIIIGSFFMLNLVLGVLSGEFAKERERVENRRAFLKLRRQQQIERELNGY
MEWISKAEEVILAEDETDGEQRHPFDGALRRTTIKKSKTDLLNPEEAEDQLADIASVG
SPFARASIKSAKLENSTFFHKKERRMRFYIRRMVKTQAFYWTVLSLVALNTLCVAIVH
YNQPEWLSDFLYYAEFIFLGLFMSEMFIKMYGLGTRPYFHSSFNCFDCGVIIGSIFEV
IWAVIKPGTSFGISVLRALRLLRIFKVTKYWASLRNLVVSLLNSMKSIISLLFLLFLF
IVVFALLGMQLFGGQFNFDEGTPPTNFDTFPAAIMTVFQILTGEDWNEVMYDGIKSQG
GVQGGMVFSIYFIVLTLFGNYTLLNVFLAIAVDNLANAQELTKVEADEQEEEEAANQK
LALQKAKEVAEVSPLSAANMSIAVKEQQKNQKPAKSVWEQRTSEMRKQNLLASREALY
NEMDPDERWKAAYTRHLRPDMKTHLDRPLVVDPQENRNNNTNKSRAAEPTVDQRLGQQ
RAEDFLRKQARYHDRARDPSGSAGLDARRPWAGSQEAELSREGPYGRESDHHAREGSL
EQPGFWEGEAERGKAGDPHRRHVHRQGGSRESRSGSPRTGADGEHRRHRAHRRPGEEG
PEDKAERRARHREGSRPARGGEGEGEGPDGGERRRRHRHGAPATYEGDARREDKERRH
RRRKENQGSGVPVSGPNLSTTRPIQQDLGRQDPPLAEDIDNMKNNKLATAESAAPHGS
LGHAGLPQSPAKMGNSTDPGPMLAIPAMATNPQNAASRRTPNNPGNPSNPGPPKTPEN
SLIVTNPSGTQTNSAKTARKPDHTTVDIPPACPPPLNHTVVQVNKNANPDPLPKKEEE
KKEEEEDDRGEDGPKPMPPYSSMFILSTTNPLRRLCHYILNLRYFEMCILMVIAMSSI
ALAAEDPVQPNAPRNNVLRYFDYVFTGVFTFEMVIKMIDLGLVLHQGAYFRDLWNILD
FIVVSGALVAFAFTGNSKGKDINTIKSLRVLRVLRPLKTIKRLPKLKAVFDCVVNSLK
NVFNILIVYMLFMFIFAVVAVQLFKGKFFHCTDESKEFEKDCRGKYLLYEKNEVKARD
REWKKYEFHYDNVLWALLTLFTVSTGEGWPQVLKHSVDATFENQGPSPGYRMEMSIFY
VVYFVVFPFFFVNIFVALIITFQEQGDKMMEEYSLEKNERACIDFAISAKPLTRHMP
QNKQSFQYRMWQFVVSPPFEYTIMAMIALNTIVLMMKFYGASVAYENALRVFNIVFTS
LFSLECVLKVMAFGILNYFRDAWNIFDFVTVLGSITDILVTEFGNPNNFINLSFLRLF
RAARLIKLLRQGYTIRILLWTFVQSFKALPYVCLLIAMLFFIYAIIGMQVFGNIGIDV
EDEDSDEDEFQITEHNNFRTFFQALMLLFRSATGEAWHNIMLSCLSGKPCDKNSGILT
RECGNEFAYFYFVSFIFLCSFLMLNLFVAVIMDNFEYLTRDSSILGPHHLDEYVRVWA
EYDPAAWGRMPYLDMYQMLRHMSPPLGLGKKCPARVAYKRLLRMDLPVADDNTVHFNS
TLMALIRTALDIKIAKGGADKQQMDAELRKEMMAIWPNLSQKTLDLLVTPHKSTDLTV
GKIYAAMMIMEYYRQSKAKKLQAMREEQDRTPLMFQRMEPPSPTQEGGPGQNALPSTQ
LDPGGALMAHESGLKESPSWVTQRAQEMFQKTGTWSPEQGPPTDMPNSQPNSQSVEMR
EMGRDGYSDSEHYLPMEGQGRAASMPRLPAENQRRRGRPRGNNLSTISDTSPMKRSAS
VLGPKARRLDDYSLERVPPEENQRHHQRRRDRSHRASERSLGRYTDVDTGLGTDLSMT
TQSGDLPSKERDQERGRPKDRKHRQHHHHHHHHHHHPPPPDKDRYAQERPDHGRARARD
QRWSRSPSEGREHMAHRQGSSSVSGSPAPSTSGTSTPRRGRRQLPQTPSTPRPHVSYS
PVIRKAGGSGPPQQQQQQQQQQQAVARPGRAATSGPRRYPGPTAEPLAGDRPPTGGHS
SGRSPRMERRVPGPARSESPRACRHGGARWPASGPHVSEGPPGPRHHGYYRGSDYDEA
DGPGSGGGEEAMAGAYDAPPPVRHASSGATGRSPRTPRASGPACASPSRHGRRLPNGY
YPAHGLARPRGPGSRKGLHEPYSESDDDWC

FIGURE 23A

1 gatgtcccga gctgctatcc ccggctcggc ccgggcagcc gccttctgag cccccgaccc
61 gaggcgccga gccgccgccg cccgatgggc tgggccgtgg agcgtctccg cagtcgtagc
121 tccagccgcc gcgctcccag ccccggcagc ctcagcatca gcggcggcgg cggcggcggc
181 ggcgtcttcc gcatcgttcg ccgcagcgta acccggagcc ctttgctctt tgcagaatgg
241 cccgcttcgg agacgagatg ccggcccgct acgggggagg aggctccggg gcagccgccg
301 gggtggtcgt gggcagcgga ggcgggcgag gagccggggg cagccggcag ggcgggcagc
361 ccgggggcgca aaggatgtac aagcagtcaa tggcgcagag agcgcggacc atggcactct
421 acaaccccat ccccgtccga cagaactgcc tcacggttaa ccggtctctc ttcctcttca
481 gcgaagacaa cgtggtgaga aaatacgcca aaaagatcac cgaatggcct ccctttgaat
541 atatgatttt agccaccatc atagcgaatt gcatcgtcct cgcactggag cagcatctgc
601 ctgatgatga caagaccccg atgtctgaac ggctggatga cacagaacca tacttcattg
661 gaatttttg tttcgaggct ggaattaaaa tcattgccct tgggtttgcc ttccacaaag
721 gctcctactt gaggaatggc tggaatgtca tggactttgt ggtggtgcta acgggcatct
781 tggcgacagt tgggacggag tttgacctac ggacgctgag ggcagttcga gtgctgcggc
841 cgctcaagct ggtgtctgga atcccaagtt tacaagtcgt cctgaagtcg atcatgaagg
901 cgatgatccc tttgctgcag atcggcctcc tcctattttt tgcaatcctt atttttgcaa
961 tcatagggtt agaattttat atgggaaaat ttcataccac ctgctttgaa gaggggacag
1021 atgacattca gggtgagtct ccggctccat gtgggacaga agagcccgcc cgcacctgcc
1081 ccaatgggac caaatgtcag ccctactggg aagggcccaa caacgggatc actcagttcg
1141 acaacatcct gtttgcagtg ctgactgttt tccagtgcat aaccatggaa gggtggactg
1201 atctcctcta caatagcaac gatgcctcag ggaacacttg gaactggttg tacttcatcc
1261 ccctcatcat catcggctcc ttttttatgc tgaaccttgt gctgggtgtg ctgtcagggg
1321 agtttgccaa agaaagggaa cgggtggaga accggcgggc ttttctgaag ctgaggcggc
1381 aacaacagat tgaacgtgag ctcaatgggt acatggaatg gatctcaaaa gcagaagagg
1441 tgatcctcgc cgaggatgaa actgacgggg agcagaggca tccctttgat ggagctctgc
1501 ggagaaccac cataaagaaa agcaagacag atttgctcaa ccccgaagag gctgaggatc
1561 agctggctga tatagcctct gtgggttctc ccttcgcccg agccagcatt aaaagtgcca
1621 agctggagaa ctcgaccttt tttcacaaaa aggagaggag gatgcgtttc tacatccgcc
1681 gcatggtcaa aactcaggcc ttctactgga ctgtactcag tttggtagct ctcaacacgc
1741 tgtgtgttgc tattgttcac tacaaccagc ccgagtggct ctccgacttc ctttactatg
1801 cagaattcat tttcttagga ctctttatgt ccgaaatgtt tataaaaatg tacgggcttg
1861 ggacgcggcc ttacttccac tcttccttca actgctttga ctgtggggtt atcattggga
1921 gcatcttcga ggtcatctgg gctgtcataa aacctggcac atcctttgga atcagcgtgt
1981 tacgagccct caggttattg cgtattttca aagtcacaaa gtactgggca tctctcagaa
2041 acctggtcgt ctctctcctc aactccatga agtccatcat cagcctgttg tttctccttt
2101 tcctgttcat tgtcgtcttc gccctttgg gaatgcaact cttcggcggc cagtttaatt
2161 tcgatgaagg gactcctccc accaacttcg atacttttcc agcagcaata atgacggtgt
2221 ttcagatcct gacgggcgaa gactggaacg aggtcatgta cgacgggatc aagtctcagg
2281 ggggcgtgca gggcggcatg gtgttctcca tctatttcat tgtactgacg ctctttggga
2341 actacaccct cctgaatgtg ttcttggcca tcgctgtgga caatctggcc aacgcccagg
2401 agctcaccaa ggtggaggcg gacgagcaag aggaagaaga agcagcgaac cagaaacttg
2461 ccctacagaa agccaaggag gtggcagaag tgagtcctct gtccgcggcc aacatgtcta
2521 tagctgtgaa agagcaacag aagaatcaaa agccagccaa gtccgtgtgg gagcagcgga
2581 ccagtgagat gcgaaagcag aacttgctgg ccagccggga ggccctgtat aacgaaatgg
2641 acccggacga gcgctggaag gctgcctaca cgcggcacct gcggccagac atgaagacgc
2701 acttggaccg gccgctggtg gtggacccgc aggagaaccg caacaacaac accaacaaga

FIGURE 23B

2761 gccgggcggc cgagcccacc gtggaccagc gcctcggcca gcagcgcgcc gaggacttcc
2821 tcaggaaaca ggcccgctac cacgatcggg cccgggaccc cagcggctcg gcgggcctgg
2881 acgcacggag gccctgggcg ggaagccagg aggccgagct gagccgggag ggaccctacg
2941 gccgcgagtc ggaccaccac gcccgggagg gcagcctgga gcaacccggg ttctgggagg
3001 gcgaggccga gcgaggcaag gccggggacc cccaccggag gcacgtgcac cggcaggggg
3061 gcagcaggga gagccgcagc gggtccccgc gcacgggcgc ggacggggag catcgacgtc
3121 atcgcgcgca ccgcaggccc ggggaggagg gtccggagga caaggcggag cggagggcgc
3181 ggcaccgcga gggcagccgg ccggcccggg gcggcgaggg cgagggcgag ggccccgacg
3241 ggggcgagcg caggagaagg caccggcatg gcgctccagc cacgtacgag ggggacgcgc
3301 ggagggagga caaggagcgg aggcatcgga ggaggaaaga gaaccagggc tccggggtcc
3361 ctgtgtcggg ccccaacctg tcaaccaccc ggccaatcca gcaggacctg ggccgccaag
3421 acccaccccc ggcagaggat attgacaaca tgaagaacaa caagctggcc accgcggagt
3481 cggccgctcc ccacggcagc cttggccacg ccggcctgcc ccagagccca gccaagatgg
3541 gaaacagcac cgaccccggc cccatgctgg ccatccctgc catggccacc aacccccaga
3601 acgccgccag ccgccggacg cccaacaacc cggggaaccc atccaatccc ggccccccca
3661 agacccccga gaatagcctt atcgtcacca accccagcgg cacccagacc aattcagcta
3721 agactgccag gaaacccgac cacaccacag tggacatccc cccagcctgc ccaccccccc
3781 tcaaccacac cgtcgtacaa gtgaacaaaa acgccaaccc agacccactg ccaaaaaaag
3841 aggaagagaa gaaggaggag gaggaagacg accgtgggga agacggccct aagccaatgc
3901 ctccctatag ctccatgttc atcctgtcca cgaccaaccc ccttcgccgc ctgtgccatt
3961 acatcctgaa cctgcgctac tttgagatgt gcatcctcat ggtcattgcc atgagcagca
4021 tcgccctggc cgccgaggac cctgtgcagc ccaacgcacc tcggaacaac gtgctgcgat
4081 actttgacta cgtttttaca ggcgtcttca cctttgagat ggtgatcaag atgattgacc
4141 tggggctcgt cctgcatcag ggtgcctact tccgtgacct ctggaatatt ctcgacttca
4201 tagtggtcag tggggccctg gtagcctttg ccttcactgg caatagcaaa ggaaaagaca
4261 tcaacacgat taaatccctc cgagtcctcc gggtgctacg acctcttaaa accatcaagc
4321 ggctgccaaa gctcaaggct gtgtttgact gtgtggtgaa ctcacttaaa aacgtcttca
4381 acatcctcat cgtctacatg ctattcatgt tcatcttcgc cgtggtggct gtgcagctct
4441 tcaaggggaa attcttccac tgcactgacg agtccaaaga gtttgagaaa gattgtcgag
4501 gcaaatacct cctctacgag aagaatgagg tgaaggcgcg agaccgggag tggaagaagt
4561 atgaattcca ttacgacaat gtgctgtggg ctctgctgac cctcttcacc gtgtccacgg
4621 gagaaggctg gccacaggtc ctcaagcatt cggtggacgc caccttgag aaccagggcc
4681 ccagccccgg gtaccgcatg gagatgtcca ttttctacgt cgtctacttt gtggtgttcc
4741 ccttcttctt tgtcaatatc tttgtggcct tgatcatcat caccttccag gagcaagggg
4801 acaagatgat ggaggaatac agcctggaga aaaatgagag ggcctgcatt gatttcgcca
4861 tcagcgccaa gccgctgacc cgacacatgc cgcagaacaa gcagagcttc cagtaccgca
4921 tgtggcagtt cgtggtgtct ccgcctttcg agtacacgat catggccatg atcgccctca
4981 acaccatcgt gcttatgatg aagttctatg gggcttctgt tgcttatgaa aatgccctgc
5041 gggtgttcaa catcgtcttc acctccctct tctctctgga atgtgtgctg aaagtcatgg
5101 cttttgggat tctgaattat ttccgcgatg cctggaacat cttcgacttt gtgactgttc
5161 tgggcagcat caccgatatc ctcgtgactg agtttgggaa tccgaataac ttcatcaacc
5221 tgagctttct ccgcctcttc cgagctgccc ggctcatcaa acttctccgt cagggttaca
5281 ccatccgcat tcttctctgg acctttgtgc agtccttcaa ggccctgcct tatgtctgtc
5341 tgctgatcgc catgctcttc ttcatctatg ccatcattgg gatgcaggtg tttggtaaca

FIG. 23C

5401 ttggcatcga cgtggaggac gaggacagtg atgaagatga gttccaaatc actgagcaca
5461 ataacttccg gaccttcttc caggccctca tgcttctctt ccggagtgcc accggggaag
5521 cttggcacaa catcatgctt tcctgcctca gcgggaaacc gtgtgataag aactctggca
5581 tcctgactcg agagtgtggc aatgaatttg cttattttta ctttgtttcc ttcatcttcc
5641 tctgctcgtt tctgatgctg aatctctttg tcgccgtcat catggacaac tttgagtacc
5701 tcacccgaga ctcctccatc ctgggccccc accacctgga tgagtacgtg cgtgtctggg
5761 ccgagtatga ccccgcagct tggggccgca tgccttacct ggacatgtat cagatgctga
5821 gacacatgtc tccgcccctg ggtctgggga agaagtgtcc ggccagagtg gcttacaagc
5881 ggcttctgcg gatggacctg cccgtcgcag atgacaacac cgtccacttc aattccaccc
5941 tcatggctct gatccgcaca gccctggaca tcaagattgc caagggagga gccgacaaac
6001 agcagatgga cgctgagctg cggaaggaga tgatggcgat ttggcccaat ctgtcccaga
6061 agacgctaga cctgctggtc acacctcaca agtccacgga cctcaccgtg gggaagatct
6121 acgcagccat gatgatcatg gagtactacc ggcagagcaa ggccaagaag ctgcaggcca
6181 tgcgcgagga gcaggaccgg acacccctca tgttccagcg catggagccc ccgtccccaa
6241 cgcaggaagg gggacctggc cagaacgccc tcccctccac ccagctggac ccaggaggag
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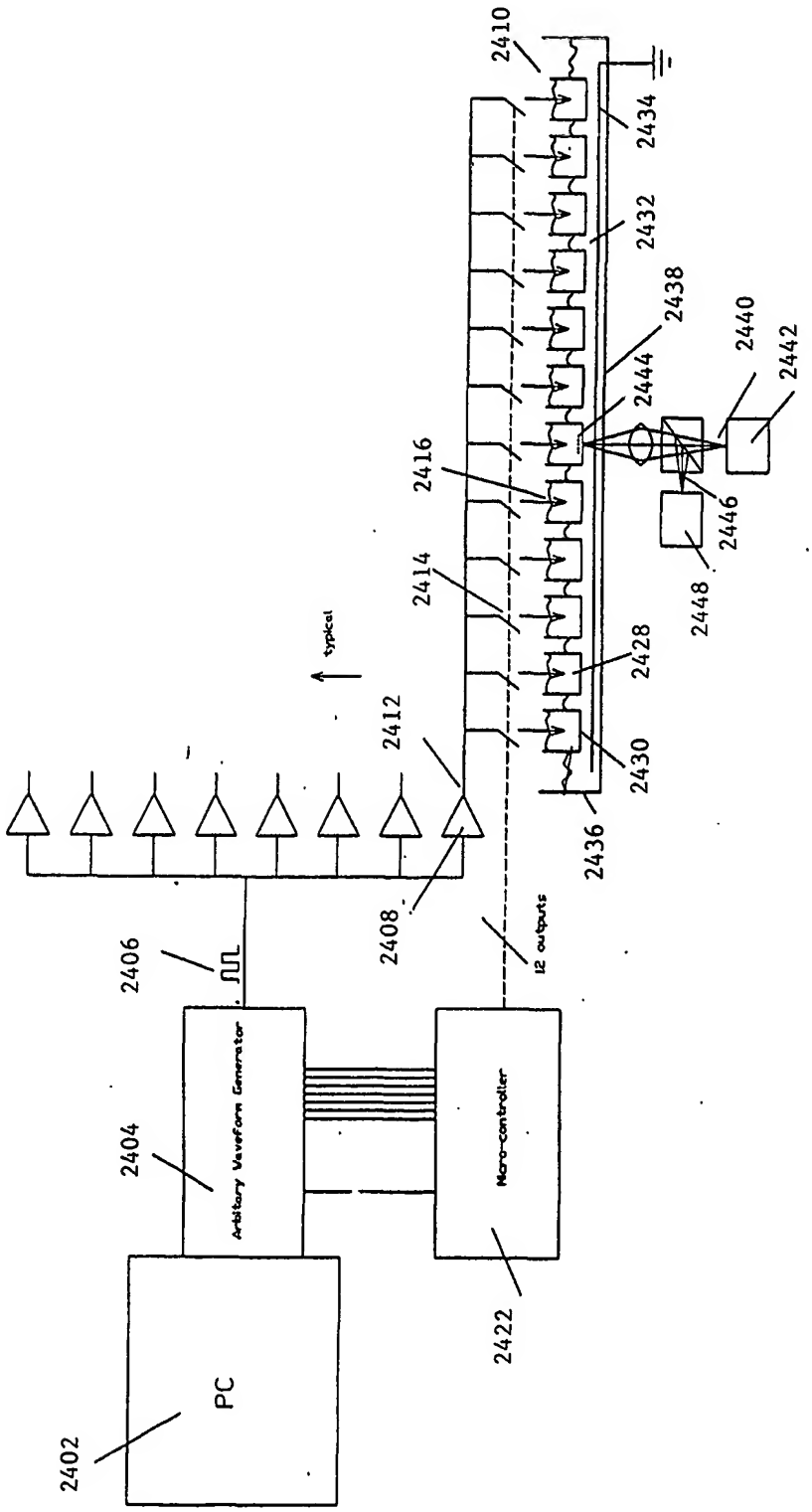
6421 ccaacagcca gcctaacict cagtccgtgg agatgcgaga gatgggcaga gatggctact
6481 ccgacagcga gcactacctc cccatggaag gccagggccg ggctgcctcc atgccccgcc
6541 tccctgcaga gaaccagagg agaaggggcc ggccacgtgg gaataacctc agtaccatct
6601 cagacaccag ccccatgaag cgttcagcct ccgtgctggg ccccaaggcc cgacgcctgg
6661 acgattactc gctggagcgg gtcccgcccg aggagaacca gcggcaccac cagcggcgcc
6721 gcgaccgcag ccaccgcgcc tctgagcgct ccctgggccg ctacaccgat gtggacacag
6781 gcttggggac agacctgagc atgaccaccc aatccgggga cctgccgtcg aaggagcggg
6841 accaggagcg gggccggccc aaggatcgga agcatcgaca gcaccaccac caccaccacc
6901 accaccacca tcccccgccc cccgacaagg accgctatgc ccaggaacgg ccggaccacg
6961 gccgggcacg ggctcgggac cagcgctggt cccgctcgcc cagcgagggc cgagagcaca
7021 tggcgcaccg gcagtagttc cgtaagtgga agcccagccc cctcaacatc tggtaccagc
7081 actccgcggc ggggccgccg ccagctcccc cagacccccc ccacccccgg gccacacgtg
7141 tcctattccc ctgtgatccg taaggccggc ggctcggggc ccccgcagca gcagcagcag
7201 cagcaggcgg tggccaggcc gggccgggcg gccaccagcg gccctcggag gtacccaggc
7261 cccacggccg agcctctggc cggagatcgg ccgcccacgg ggggccacag cagcggccgc
7321 tcgcccagga tggagaggcg ggtcccaggc ccggcccgga gcgagtcccc cagggcctgt
7381 cgacacggcg gggcccggtg gccggcatct ggcccgcacg tgtccgaggg gcccccgggt
7441 ccccggcacc atggctacta ccggggctcc gactacgacg aggccgatgg cccgggcagc
7501 gggggcggcg aggaggccat ggccgggggc tacgacgcgc cacccccccgt acgacacgcg
7561 tcctcgggcg ccaccgggcg ctcgcccagg actccccggg cctcgggccc ggcctgcgcc
7621 tcgccttctc ggcacggccg gcgactcccc aacggctact acccggcgca cggactggcc
7681 aggccccgcg ggccgggctc caggaagggc ctgcacgaac cctacagcga gagtgacgat
7741 gattggtgct aagcccgggc gaggtggcgc ccgcccggcc ccccacgcac c

FIG. 23D

MARFGDEMPARYGGGGSGAAAGVVVGSGGGRGAGGSRQGGQPGA
QRMYKQSMAQRARTMALYNPIPVRQNCLTVNRSLFLFSEDNVVRKYAKKITEWPPFEY
MILATIIANCIVLALEQHLPDDDKTPMSERLDDTEPYFIGIFCFEAGIKIIALGFAFH
KGSYLRNGWNVMDFVVVLTGILATVGTEFDLRTLRAVRVLRPLKLVSGIPSLQVVLKS
IMKAMIPLLQIGLLLFFAILIFAIIGLEFYMGKFHTTCFEEGTDDIQGESPAPCGTEE
PARTCPNGTKCQPYWEGPNNGITQFDNILFAVLTVFQCITMEGWTDLLYNSNDASGNT
WNWLYFIPLIIIGSFFMLNLVLGVLSGEFAKERERVENRRAFLKLRRQQQIERELNGY
MEWISKAEEVILAEDETDGEQRHPFDGALRRTTIKKSKTDLLNPEEAEDQLADIASVG
SPFARASIKSAKLENSTFFHKKERRMRFYIRRMVKTQAFYWTVLSLVALNTLCVAIVH
YNQPEWLSDFLYYAEFIFLGLFMSEMFIKMYGLGTRPYFHSSFNCFDCGVIIGSIFEV
IWAVIKPGTSFGISVLRALRLLRIFKVTKYWASLRNLVVSLLNSMKSIISLLFLLFLF
IVVFALLGMQLFGGQFNFDEGTPPTNFDTFPAAIMTVFQILTGEDWNEVMYDGIKSQG
GVQGGMVFSIYFIVLTLFGNYTLLNVFLAIAVDNLANAQELTKVEADEQEEEEAANQK
LALQKAKEVAEVSPLSAANMSIAVKEQQKNQKPAKSVWEQRTSEMRKQNLLASREALY
NEMDPDERWKAAYTRHLRPDMKTHLDRPLVVDPQENRNNNTNKSRAAEPTVDQRLGQQ
RAEDFLRKQARYHDRARDPSGSAGLDARRPWAGSQEAELSREGPYGRESDHHAREGSL
EQPGFWEGEAERGKAGDPHRRHVHRQGGSRESRSGSPRTGADGEHRRHRAHRRPGEEG
PEDKAERRARHREGSRPARGGEGEGEGPDGGERRRRHRHGAPATYEGDARREDKERRH
RRRKENQGSGVPVSGPNLSTTRPIQQDLGRQDPPLAEDIDNMKNNKLATAESAAPHGS
LGHAGLPQSPAKMGNSTDPGPMLAIPAMATNPQNAASRRTPNNPGNPSNPGPPKTPEN
SLIVTNPSGTQTNSAKTARKPDHTTVDIPPACPPPLNHTVVQVNKNANPDPLPKKEEE
KKEEEEDDRGEDGPKPMPPYSSMFILSTTNPLRRLCHYILNLRYFEMCILMVIAMSSI
ALAAEDPVQPNAPRNNVLRYFDYVFTGVFTFEMVIKMIDLGLVLHQGAYFRDLWNILD
FIVVSGALVAFAFTGNSKGKDINTIKSLRVLRVLRPLKTIKRLPKLKAVFDCVVNSLK
NVFNILIVYMLFMFIFAVVAVQLFKGKFFHCTDESKEFEKDCRGKYLLYEKNEVKARD
REWKKYEFHYDNVLWALLTLFTVSTGEGWPQVLKHSVDATFENQGPSPGYRMEMSIFY
VVYFVVFPFFFVNIFVALIIITFQEQGDKMMEEYSLEKNERACIDFAISAKPLTRHMP
QNKQSFQYRMWQFVVSPPFEYTIMAMIALNTIVLMMKFYGASVAYENALRVFNIVFTS
LFSLECVLKVMAFGILNYFRDAWNIFDFVTVLGSITDILVTEFGNPNNFINLSFLRLF
RAARLIKLLRQGYTIRILLWTFVQSFKALPYVCLLIAMLFFIYAIIGMQVFGNIGIDV
EDEDSDEDEFQITEHNNFRTFFQALMLLFRSATGEAWHNIMLSCLSGKPCDKNSGILT
RECGNEFAYFYFVSFIFLCSFLMLNLFVAVIMDNFEYLTRDSSILGPHHLDEYVRVWA
EYDPAAWGRMPYLDMYQMLRHMSPPLGLGKKCPARVAYKRLLRMDLPVADDNTVHFNS
TLMALIRTALDIKIAKGGADKQQMDAELRKEMMAIWPNLSQKTLDLLVTPHKSTDLTV
GKIYAAMMIMEYYRQSKAKKLQAMREEQDRTPLMFQRMEPPSPTQEGGPGQNALPSTQ
LDPGGALMAHESGLKESPSWVTQRAQEMFQKTGTWSPEQGPPTDMPNSQPNSQSVEMR
EMGRDGYSDSEHYLPMEGQGRAASMPRLPAENQRRRGRPRGNNLSTISDTSPMKRSAS
VLGPKARRLDDYSLERVPPEENQRHHQRRRDRSHRASERSLGRYTDVDTGLGTDLSMT
TQSGDLPSKERDQERGRPKDRKHRQHHHHHHHHHHHPPPPDKDRYAQERPDHGRARARD
QRWSRSPSEGREHMAHRQ

FIG. 24


2410
2434
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2416
2446
2414
2448
typical
2428
2412
2430
2436
2408
12 outputs
2406
2404
Arbitary Waveform Generator
Micro-controller
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PC
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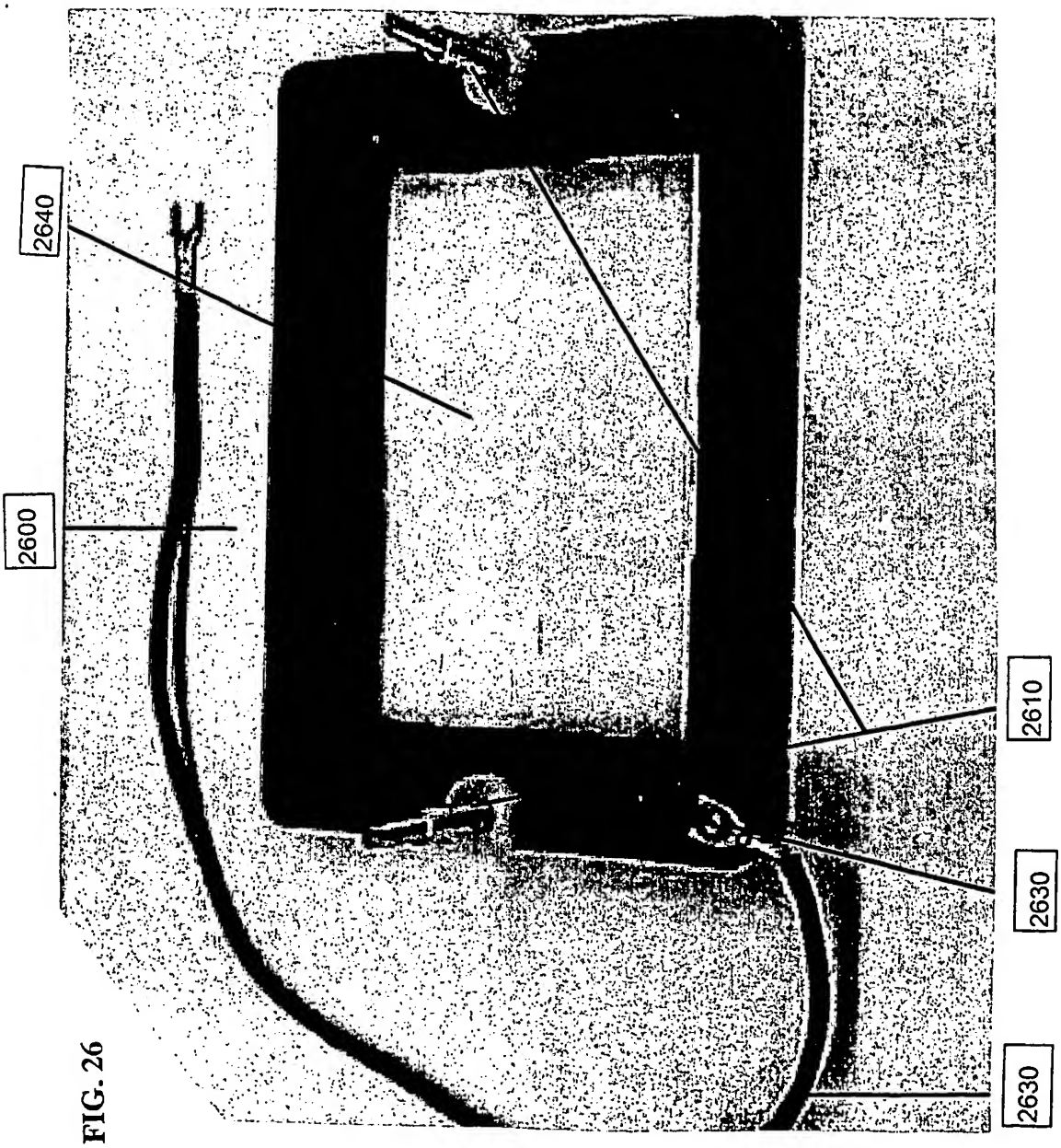


FIG. 26

FIG. 27

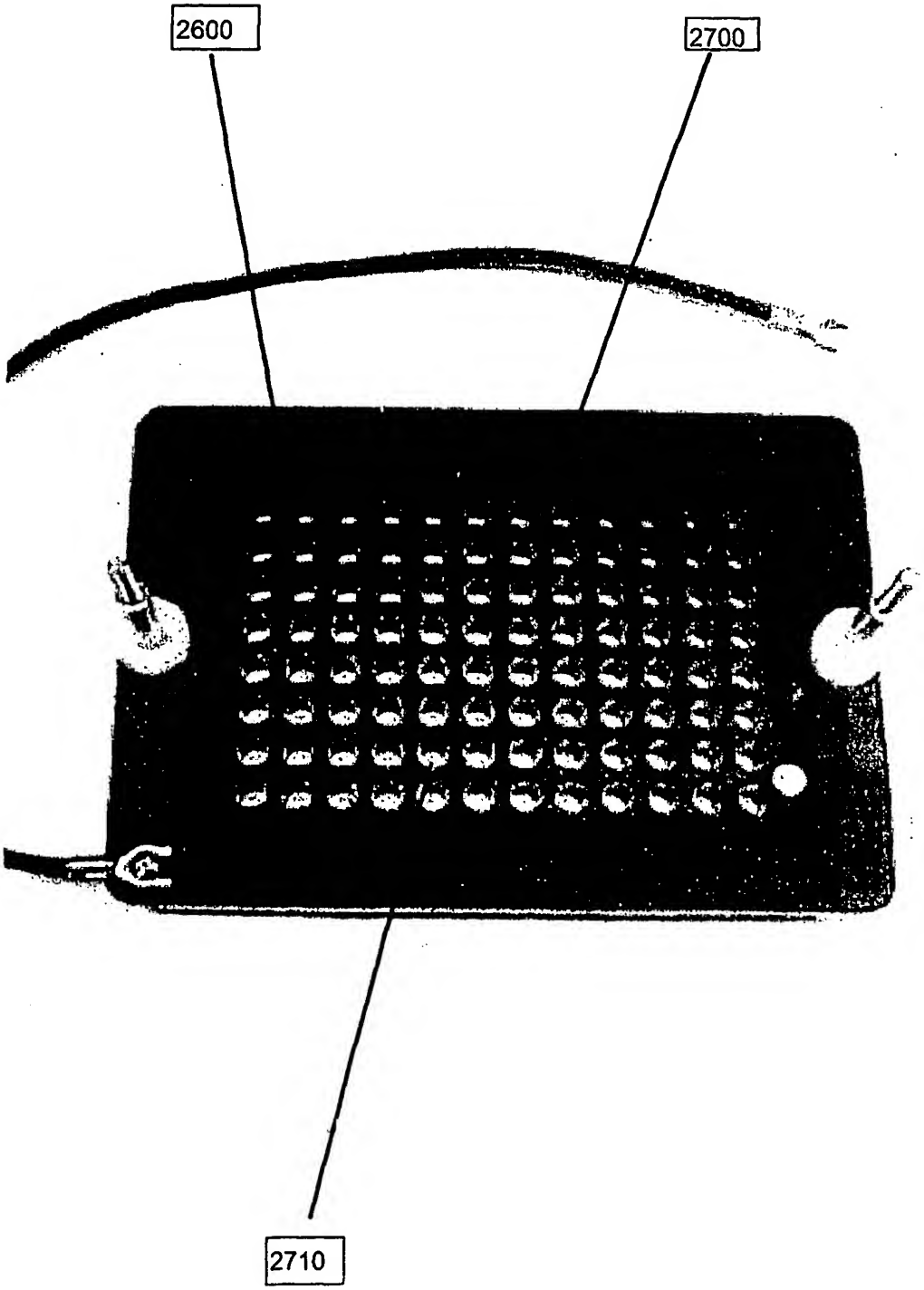
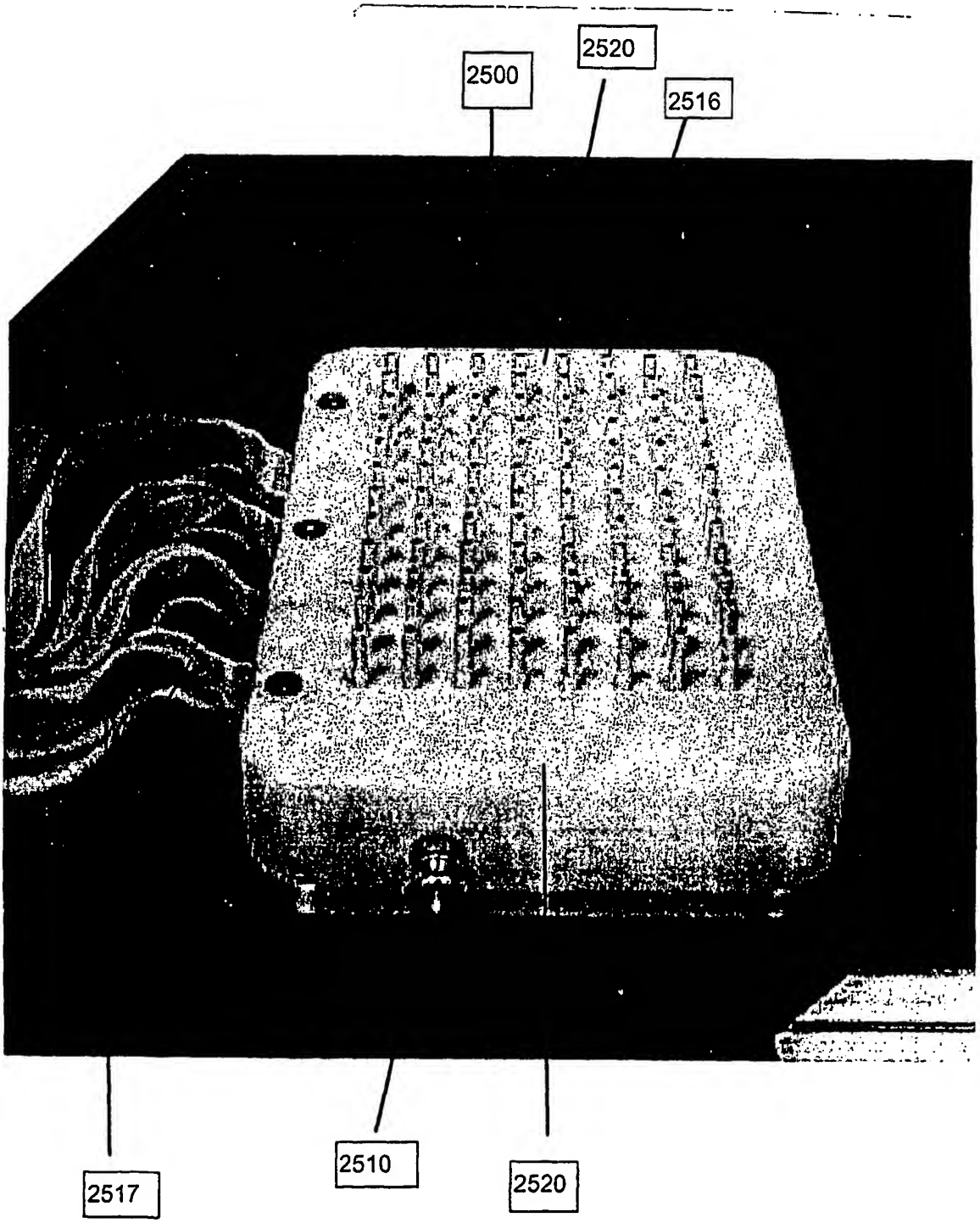


FIG. 25



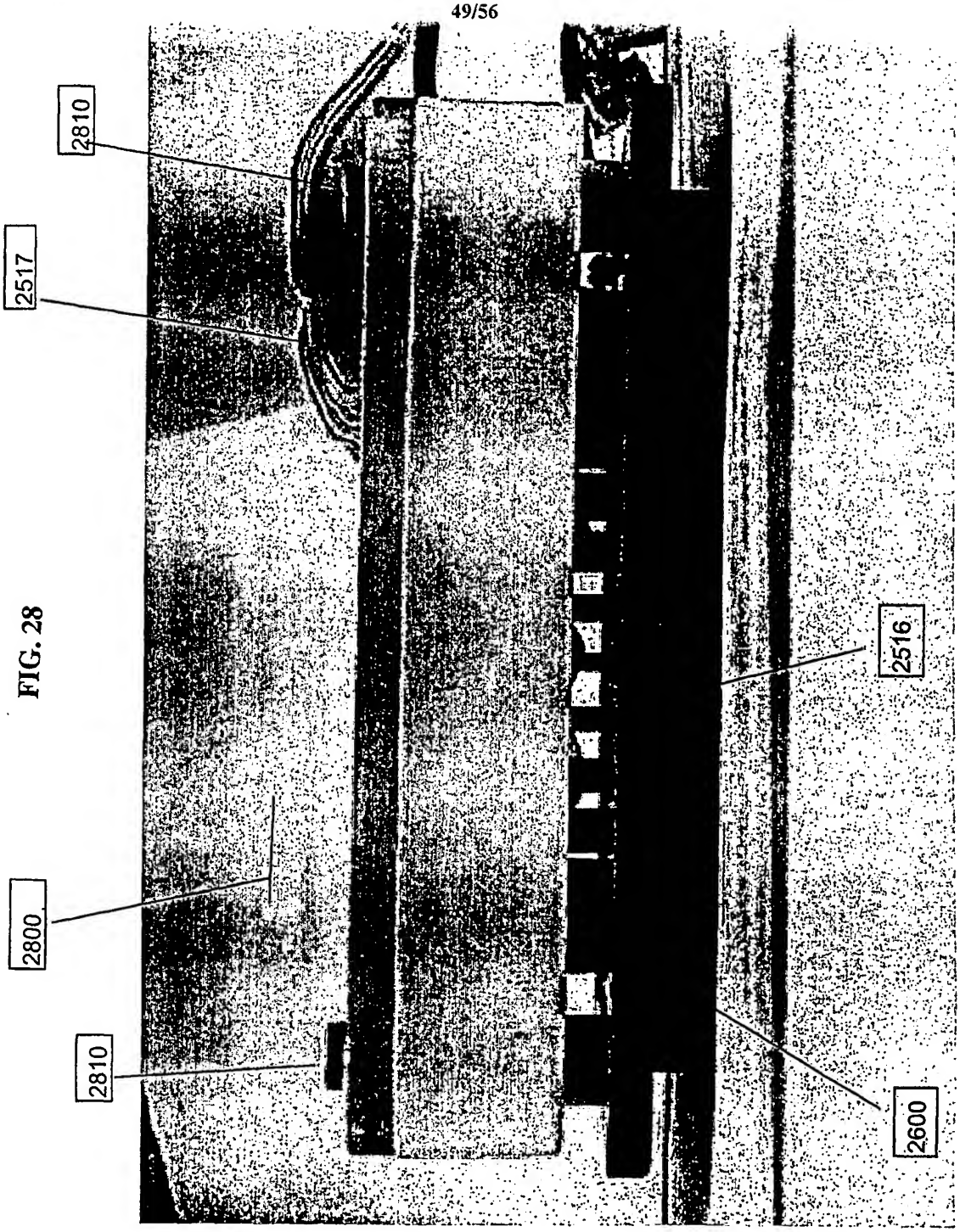


FIG. 28

FIG. 29

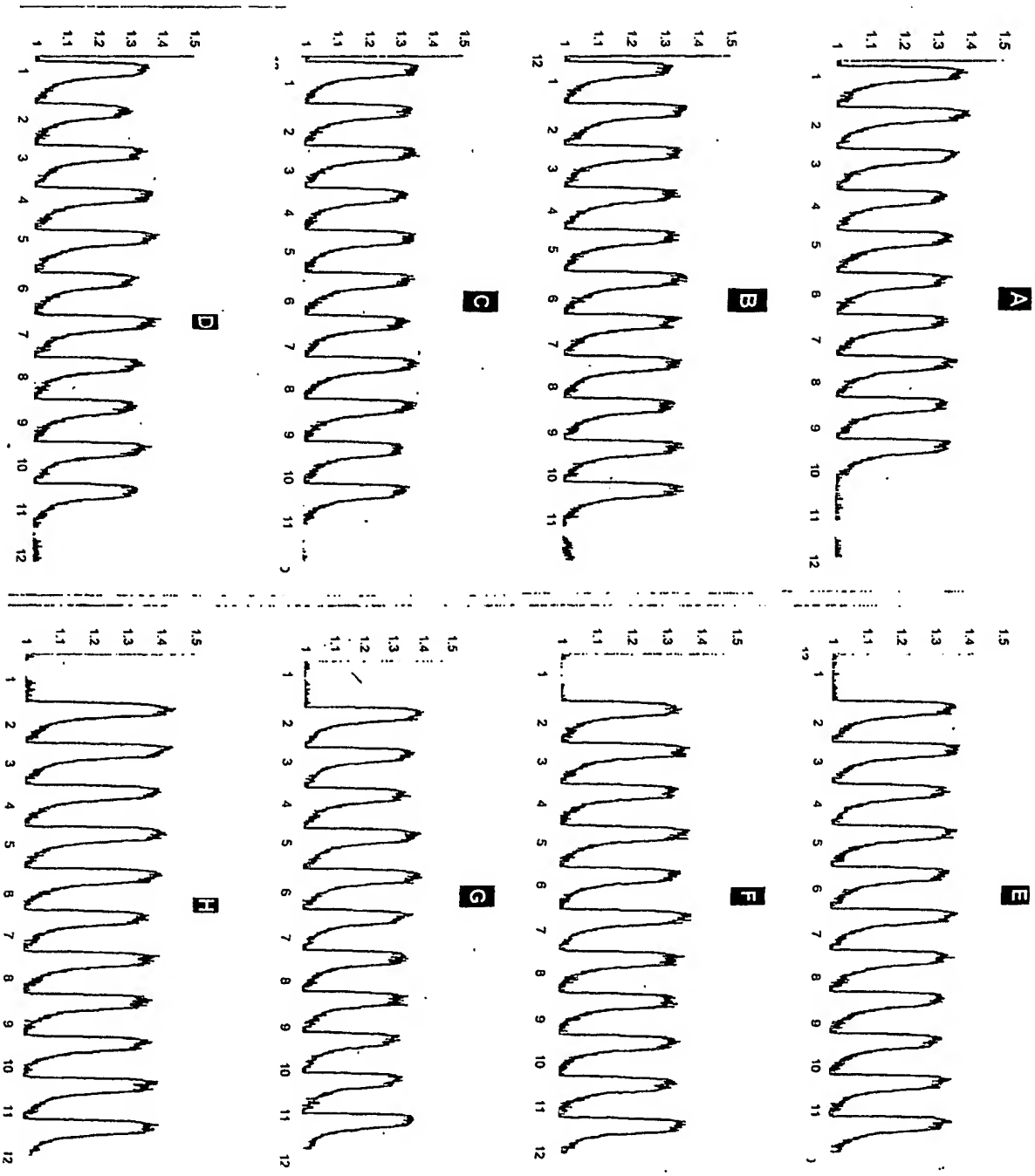
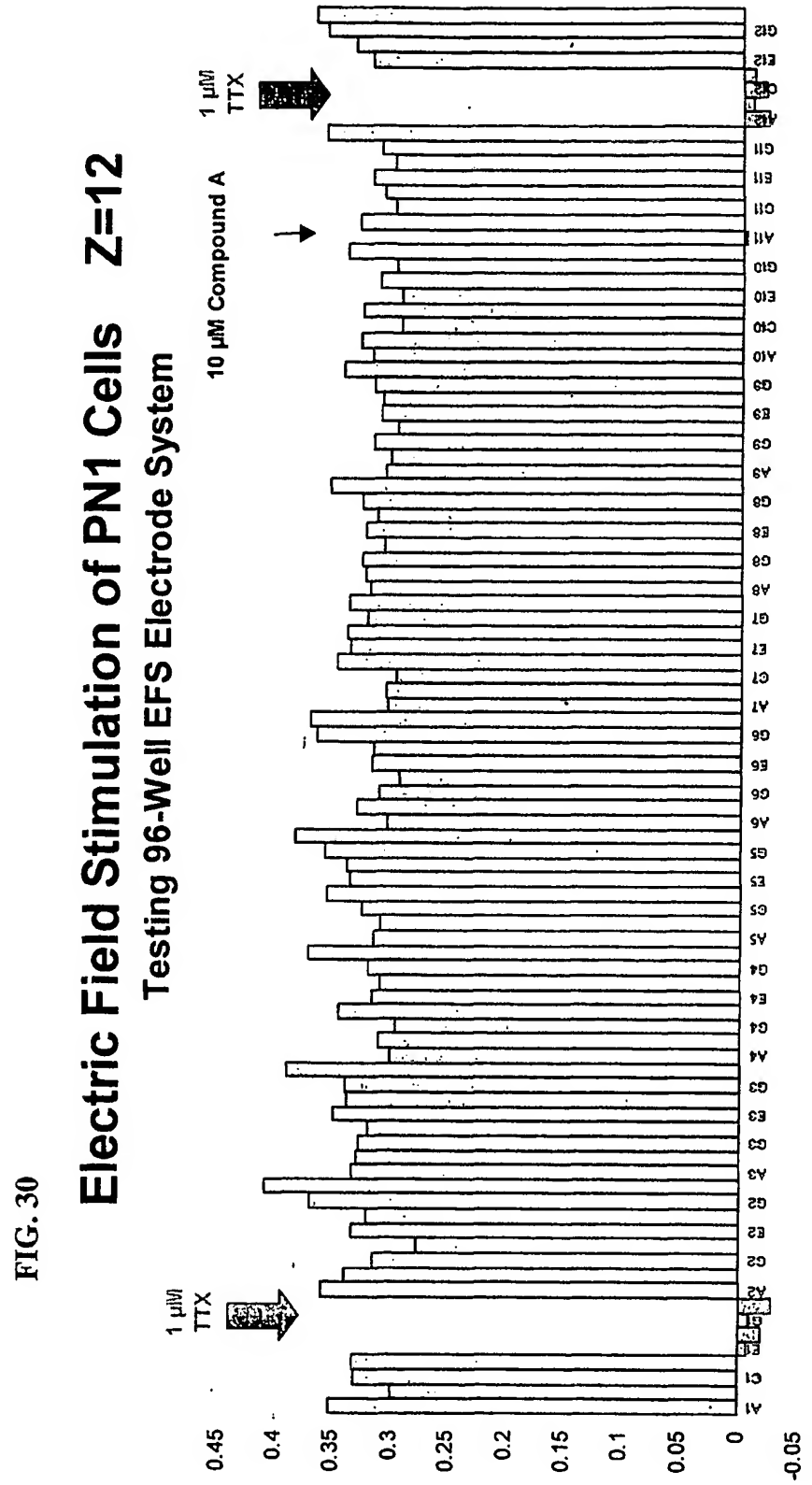


FIG. 30


Electric Field Stimulation of PN1 Cells Z=12
Testing 96-Well EFS Electrode System
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10 µM Compound A
1 µM TTX
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0.4
0.35
0.3
0.25
0.2
0.15
0.1
0.05
0
-0.05

FIG. 31

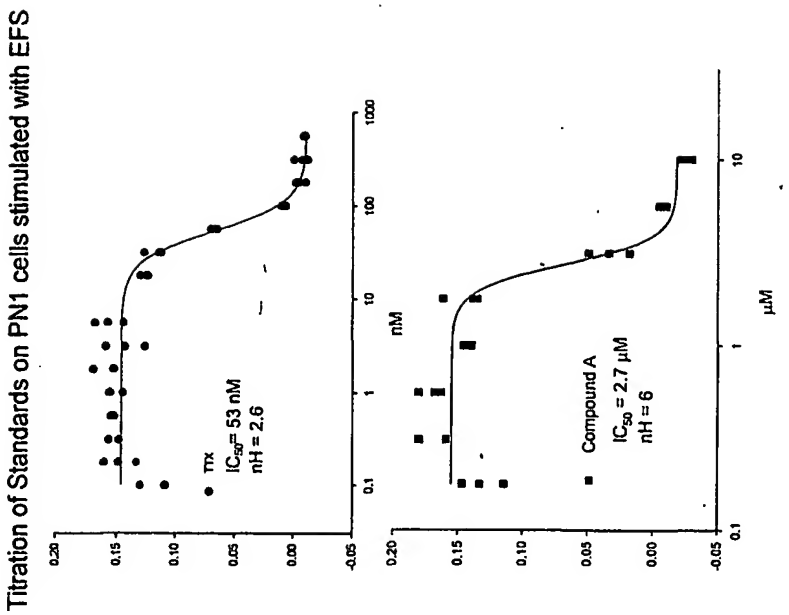

Titration of Standards on PN1 cells stimulated with EFS
Tx
IC50= 53 nM
nH = 2.6
nM
Compound A
IC50 = 2.7 µM
nH = 6
µM

FIG. 32

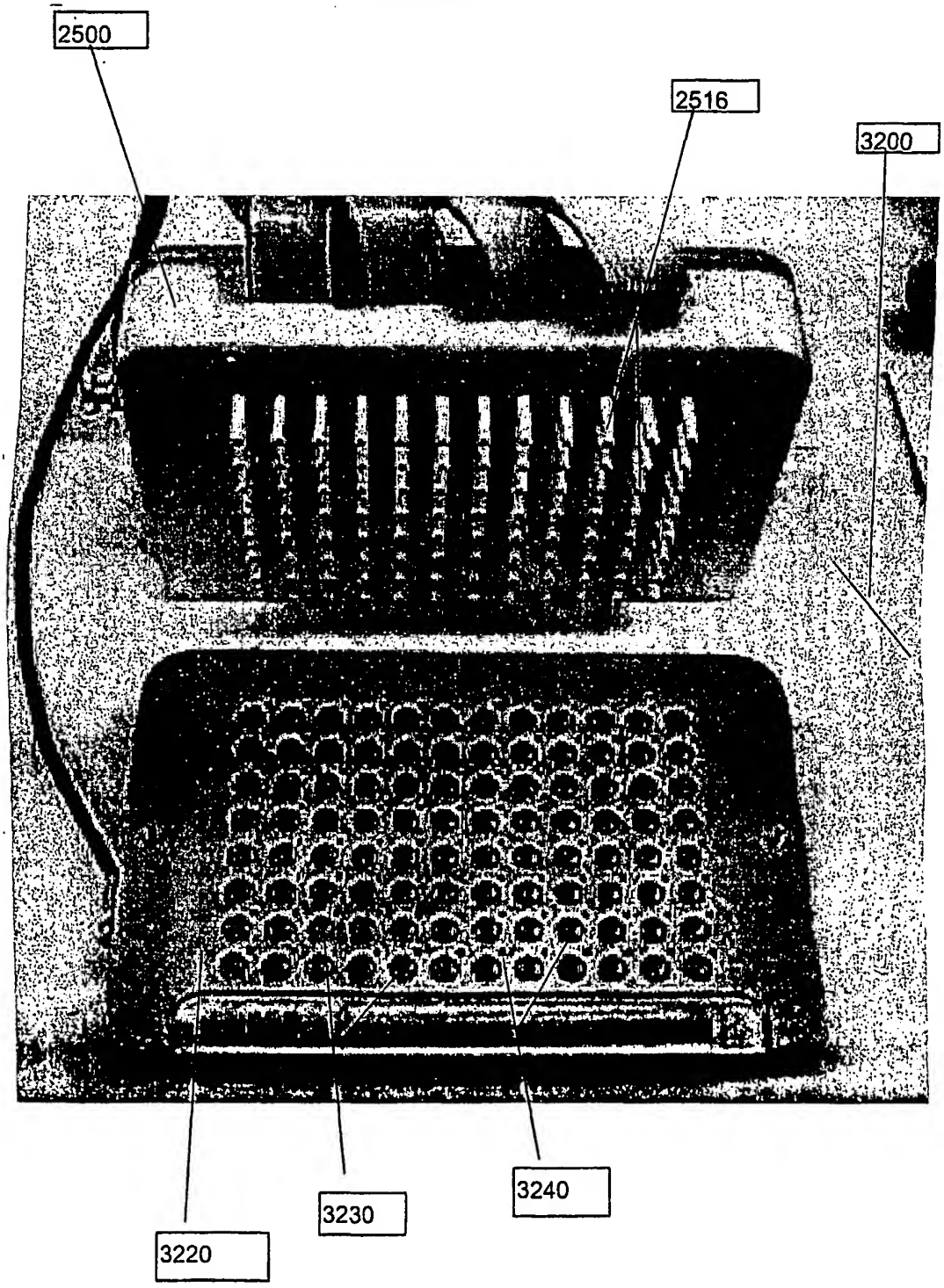


FIG. 33

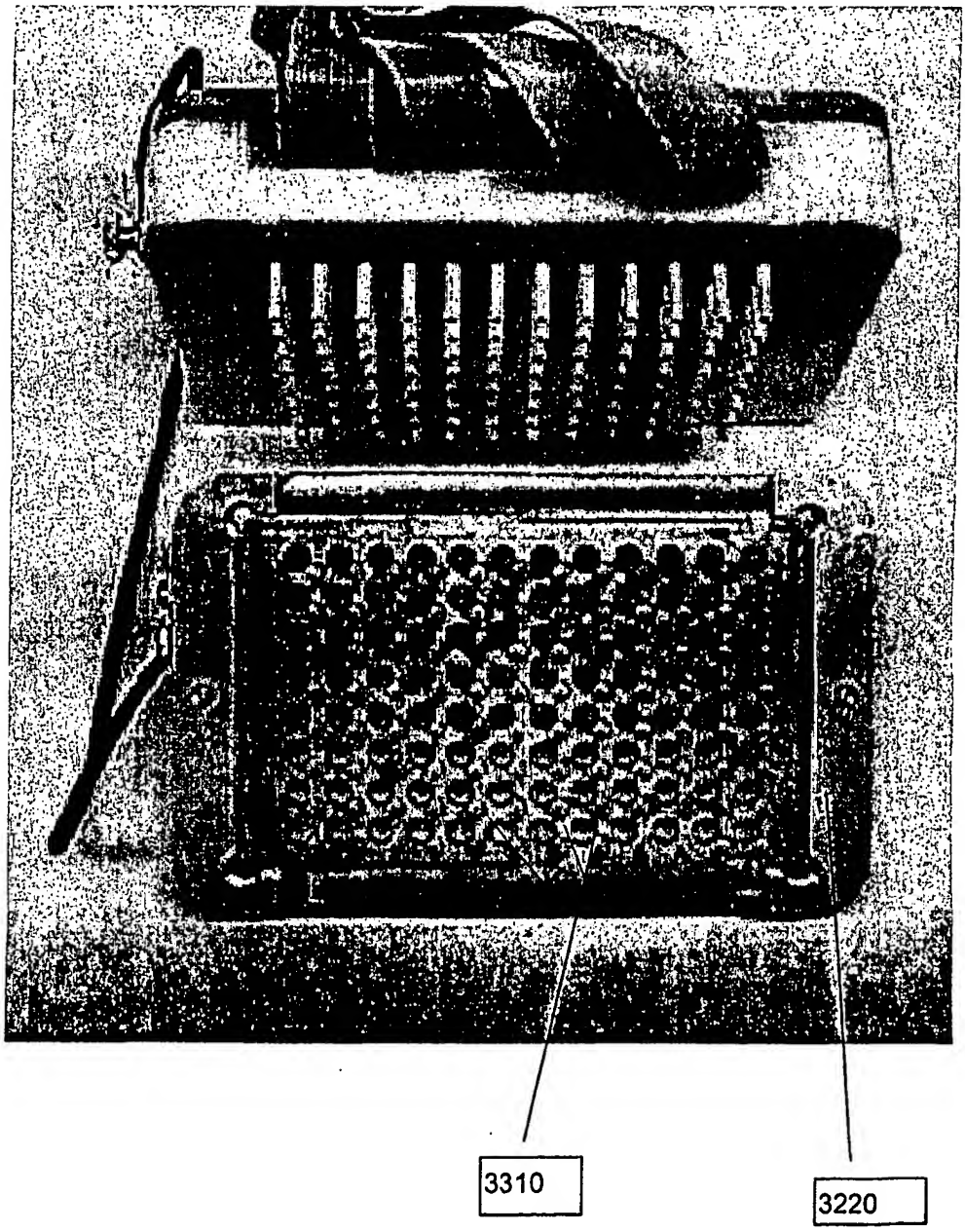


FIG. 34

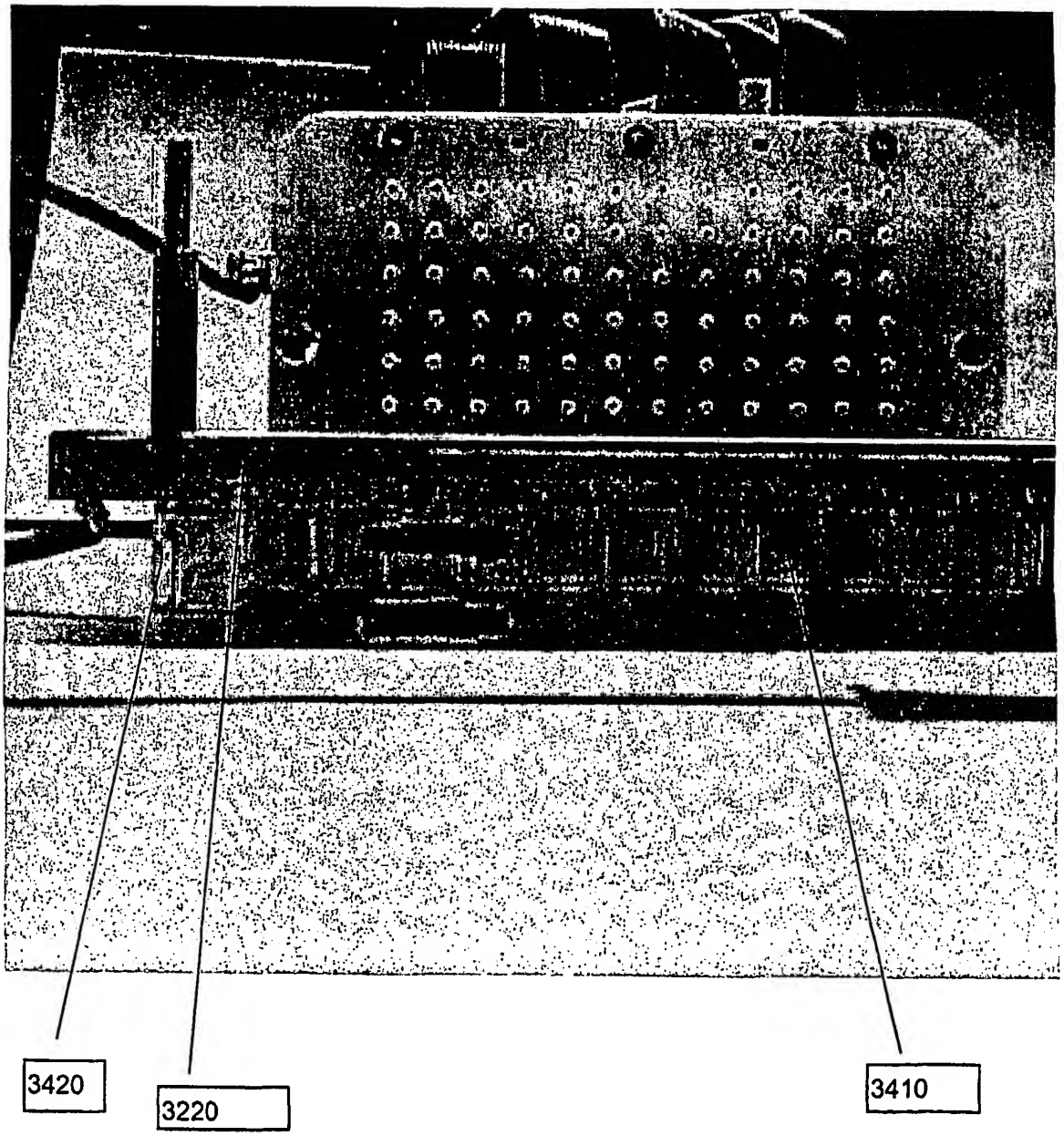
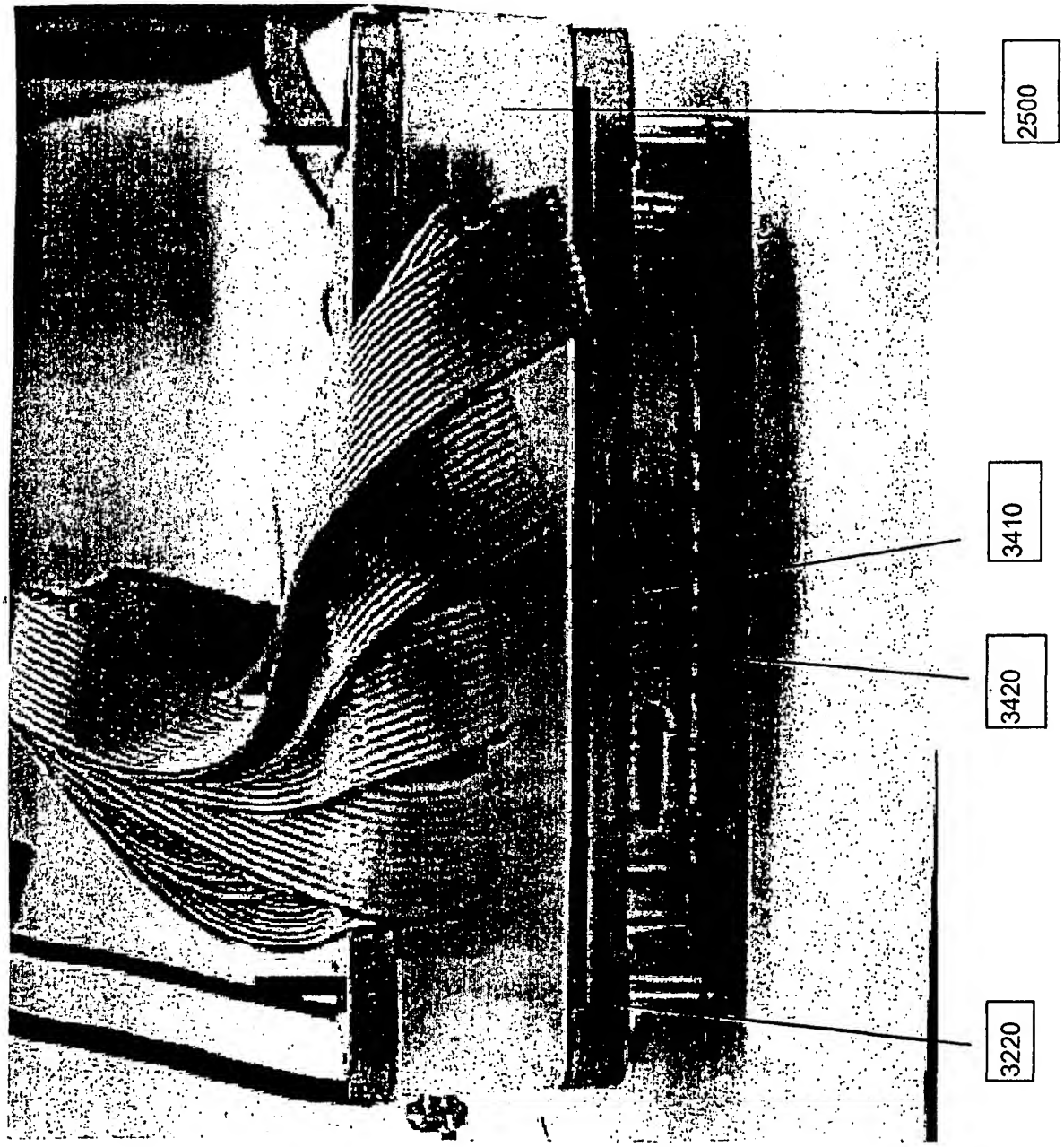


FIG. 35



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Glu Lys Pro Arg Pro Gln Leu Asp Leu Lys Ala Cys Asn Gln Leu Pro
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Lys Phe Tyr Gly Glu Leu Pro Ala Glu Leu Ile Gly Glu Pro Leu Glu
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Lys Gly Arg Thr Ile Ser Arg Phe Ser Ala Thr Arg Ala Leu Trp Leu
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Phe Ser Pro Phe Asn Leu Ile Arg Arg Thr Ala Ile Lys Val Ser Val
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Lys Lys Lys Leu Gly Gly Gln Asp Ile Phe Met Thr Glu Glu Gln Lys
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Pro Ile Pro Arg Pro Leu Asn Lys Phe Gln Gly Phe Val Phe Asp Ile
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Val Thr Arg Gln Ala Phe Asp Ile Thr Ile Met Val Leu Ile Cys Leu
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Asn Met Ile Thr Met Met Val Glu Thr Asp Asp Gln Ser Glu Glu Lys
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Thr Lys Ile Leu Gly Lys Ile Asn Gln Phe Phe Val Ala Val Phe Thr
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Gly Glu Cys Val Met Lys Met Phe Ala Leu Arg Gln Tyr Tyr Phe Thr
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Pro Thr Leu Phe Arg Val Ile Arg Leu Ala Arg Ile Gly Arg Ile Leu
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Arg Leu Ile Arg Ala Ala Lys Gly Ile Arg Thr Leu Leu Phe Ala Leu
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Val Pro Gly Asp Lys Ile His Cys Leu Asp Ile Leu Phe Ala Phe Thr
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Met Glu Glu Lys Phe Met Ala Thr Asn Leu Ser Lys Ser Ser Tyr Glu
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Pro Ile Ala Thr Thr Leu Arg Trp Lys Gln Glu Asp Ile Ser Ala Thr
                1845                1850                1855
Val Ile Gln Lys Ala Tyr Arg Ser Tyr Val Leu His Arg Ser Met Ala
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Leu Ser Asn Thr Pro Cys Val Pro Arg Ala Glu Glu Glu Ala Ala Ser
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Leu Pro Asp Glu Gly Phe Val Ala Phe Thr Ala Asn Glu Asn Cys Val
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Leu Pro Asp Lys Ser Glu Thr Ala Ser Ala Thr Ser Phe Pro Pro Ser
1905                1910                1915                1920
Tyr Glu Ser Val Thr Arg Gly Leu Ser Asp Arg Val Asn Met Arg Thr
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        1955
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        35                  40                  45
Ser Pro Ser Glu Ser Pro Ala Ala Glu Arg Gly Ala Glu Leu Gly Ala
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Asp Glu Glu Gln Arg Val Pro Tyr Pro Ala Leu Ala Ala Thr Val Phe
65                  70                  75                  80
Phe Cys Leu Gly Gln Thr Thr Arg Pro Arg Ser Trp Cys Leu Arg Leu
                85                  90                  95
Val Cys Asn Pro Trp Phe Glu His Val Ser Met Leu Val Ile Met Leu
            100                 105                 110
Asn Cys Val Thr Leu Gly Met Phe Arg Pro Cys Glu Asp Val Glu Cys
        115                 120                 125
Gly Ser Glu Arg Cys Asn Ile Leu Glu Ala Phe Asp Ala Phe Ile Phe
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Ala Phe Phe Ala Val Glu Met Val Ile Lys Met Val Ala Leu Gly Leu
145                 150                 155                 160
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Phe Ile Val Val Ala Gly Met Met Glu Tyr Ser Leu Asp Gly His Asn
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Val Ser Leu Ser Ala Ile Arg Thr Val Arg Val Leu Arg Pro Leu Arg
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Arg Asn Arg Cys Phe Leu Asp Ser Ala Phe Val Arg Asn Asn Asn Leu
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Thr Phe Leu Arg Pro Tyr Tyr Gln Thr Glu Glu Gly Glu Glu Asn Pro
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Phe Ile Cys Ser Ser Arg Arg Asp Asn Gly Met Gln Lys Cys Ser His
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Ile Pro Gly Arg Arg Glu Leu Arg Met Pro Cys Thr Leu Gly Trp Glu
305                 310                 315                 320
Ala Tyr Thr Gln Pro Gln Ala Glu Gly Val Gly Ala Ala Arg Asn Ala
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Cys Ile Asn Trp Asn Gln Tyr Tyr Asn Val Cys Arg Ser Gly Asp Ser
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Asn Pro His Asn Gly Ala Ile Asn Phe Asp Asn Ile Gly Tyr Ala Trp
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Ile Ala Ile Phe Gln Val Ile Thr Leu Glu Gly Trp Val Asp Ile Met
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Tyr Tyr Val Met Asp Ala His Ser Phe Tyr Asn Phe Ile Tyr Phe Ile
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Leu Leu Ile Ile Val Gly Ser Phe Phe Met Ile Asn Leu Cys Leu Val
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Met Arg Glu Gln Arg Ala Arg His Leu Ser Asn Asp Ser Thr Leu Ala
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Ser Phe Ser Glu Pro Gly Ser Cys Tyr Glu Glu Leu Leu Lys Tyr Val
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Gly Gln Gly Pro Gly His Arg Gln Arg Arg Ala Gly Arg His Thr Ala
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Ser Val His His Leu Val Tyr His His His His His His His His His
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Tyr His Phe Ser His Gly Ser Pro Arg Arg Pro Gly Pro Glu Pro Gly
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Pro Gly Thr Gly Gly His Gly Pro Leu Ser Leu Asn Ser Pro Asp Pro
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Tyr Glu Lys Ile Pro His Val Val Gly Glu His Gly Leu Gly Gln Ala
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Pro Gly His Leu Ser Gly Leu Ser Val Pro Cys Pro Leu Pro Ser Pro
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Pro Ala Gly Thr Leu Thr Cys Glu Leu Lys Ser Cys Pro Tyr Cys Thr
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Arg Ala Leu Glu Asp Pro Glu Gly Glu Leu Ser Gly Ser Glu Ser Gly
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740 745 750
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755 760 765
Pro Gly Glu Pro Gly Trp Met Gly Arg Leu Trp Val Thr Phe Ser Gly
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915 920 925
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Trp Cys Arg Ser Arg Glu Ala Trp Ala Leu Tyr Leu Phe Ser Pro Gln
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Ser Val Ser Asn Tyr Ile Phe Thr Ala Ile Phe Val Ala Glu Met Met
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Val Lys Val Val Ala Leu Gly Leu Leu Ser Gly Glu His Ala Tyr Leu
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Gln Ser Ser Trp Asn Leu Leu Asp Gly Leu Leu Val Leu Val Ser Leu
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Val Ile Ser Arg Ala Pro Gly Leu Lys Leu Val Val Glu Thr Leu Ile
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Phe Ile Ile Phe Gly Ile Leu Gly Val Gln Leu Phe Lys Gly Lys Phe
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Tyr Tyr Cys Glu Gly Pro Asp Thr Arg Asn Ile Ser Thr Lys Ala Gln
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Cys Arg Ala Ala His Tyr Arg Trp Val Arg Arg Lys Tyr Asn Phe Asp
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Asn Leu Gly Gln Ala Leu Met Ser Leu Phe Val Leu Ser Ser Lys Asp
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Ser Phe Leu Leu Ile Val Ser Phe Phe Val Leu Asn Met Phe Val Gly
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Val Val Val Glu Asn Phe His Lys Cys Arg Gln His Gln Glu Ala Glu
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Glu Ala Arg Arg Arg Glu Glu Lys Arg Leu Arg Arg Leu Glu Arg Arg
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His Tyr Leu Asp Leu Phe Ile Thr Phe Ile Ile Cys Val Asn Val Ile
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Thr Met Ser Met Glu His Tyr Asn Gln Pro Lys Ser Leu Asp Glu Ala
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Leu Lys Tyr Cys Asn Tyr Val Phe Thr Ile Val Phe Val Phe Glu Ala
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Trp Asn Gln Leu Asp Leu Ala Ile Val Leu Leu Ser Leu Met Gly Ile
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Ser Glu Asp Asn Pro Cys Glu Gly Leu Ser Arg His Ala Thr Phe Ser
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Asp Lys His Cys Leu Ser Tyr Leu Pro Ala Leu Ser Pro Val Tyr Phe
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Gly Ser Ala Arg Arg Val Asp Ala Asp Arg Pro Pro Leu Pro Gln Glu
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Ser Pro Gly Ala Arg Asp Ala Pro Asn Leu Val Ala Arg Lys Val Ser
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Val Val Pro Ala Ser Ala Pro His Pro Arg Pro Leu Gln Glu Val Glu
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His Ser Pro Pro Ala Glu Ser Cys Ala Ser Leu Gln Ile Pro Leu Ala
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Val Ser Ser Pro Ala Arg Ser Gly Glu Pro Leu His Ala Leu Ser Pro
1985                1990                1995                2000
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Glu Ala Val His Thr Asp Ser Leu Glu Gly Lys Ile Asp Ser Pro Arg
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Gly Ser Gly Glu Pro Gly Glu Ala Lys Ala Trp Gly Pro Glu Ala Glu
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Pro Ala Leu Gly Ala Arg Arg Lys Lys Lys Met Ser Pro Pro Cys Ile
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Arg Ala Glu His Leu Thr Val Pro Ser Phe Ala Phe Glu Pro Leu Asp
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Leu Gly Val Pro Ser Gly Asp Pro Phe Leu Asp Gly Ser His Ser Val
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Thr Pro Glu Ser Arg Ala Ser Ser Ser Gly Ala Ile Val Pro Leu Glu
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Pro Pro Glu Ser Glu Pro Pro Met Pro Val Gly Asp Pro Pro Glu Lys
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Lys Gln Asn Cys Phe Thr Val Asn Arg Ser Leu Phe Val Phe Ser Glu
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Arg Leu Asp Asp Thr Glu Pro Tyr Phe Ile Gly Ile Phe Cys Phe Glu
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Ala Glu Glu Asp Arg Asn Ala Glu Glu Lys Ser Pro Leu Asp Val Leu
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Lys Arg Ala Ala Thr Lys Lys Ser Arg Asn Asp Leu Ile His Ala Glu
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Glu Gly Glu Asp Arg Phe Ala Asp Leu Cys Ala Val Gly Ser Pro Phe
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Ala Arg Ala Ser Leu Lys Ser Gly Lys Thr Glu Ser Ser Ser Tyr Phe
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Arg Arg Lys Glu Lys Met Phe Arg Phe Phe Ile Arg Arg Met Val Lys
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Ala Gln Ser Phe Tyr Trp Val Val Leu Cys Val Val Ala Leu Asn Thr
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Leu Cys Val Ala Met Val His Tyr Asn Gln Pro Arg Arg Leu Thr Thr
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Thr Leu Tyr Phe Ala Glu Phe Val Phe Leu Gly Leu Phe Leu Thr Glu
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Met Ser Leu Lys Met Tyr Gly Leu Gly Pro Arg Ser Tyr Phe Arg Ser
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Ser Phe Asn Cys Phe Asp Phe Gly Val Ile Val Gly Ser Val Phe Glu
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Val Val Trp Ala Ala Ile Lys Pro Gly Ser Ser Phe Gly Ile Ser Val
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Leu Arg Ala Leu Arg Leu Leu Arg Ile Phe Lys Val Thr Lys Tyr Trp
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Ser Ser Leu Arg Asn Leu Val Val Ser Leu Leu Asn Ser Met Lys Ser
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Ile Ile Ser Leu Leu Phe Leu Leu Phe Leu Phe Ile Val Val Phe Ala
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Leu Leu Gly Met Gln Leu Phe Gly Gly Gln Phe Asn Phe Gln Asp Glu
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Thr Pro Thr Thr Asn Phe Asp Thr Phe Pro Ala Ala Ile Leu Thr Val
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Phe Gln Ile Leu Thr Gly Glu Asp Trp Asn Ala Val Met Tyr His Gly
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Ile Glu Ser Gln Gly Gly Val Ser Lys Gly Met Phe Ser Ser Phe Tyr
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Phe Ile Val Leu Thr Leu Phe Gly Asn Tyr Thr Leu Leu Asn Val Phe
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Asp Glu Glu Glu Met Glu Glu Ala Ala Asn Gln Lys Leu Ala Leu Gln
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Lys Ala Lys Glu Val Ala Glu Val Ser Pro Met Ser Ala Ala Asn Ile
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Ser Ile Ala Ala Arg Gln Gln Asn Ser Ala Lys Ala Arg Ser Val Trp
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Glu Gln Arg Ala Ser Gln Leu Arg Leu Gln Asn Leu Arg Ala Ser Cys
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Glu Ala Leu Tyr Ser Glu Met Asp Pro Glu Glu Arg Leu Arg Phe Ala
785                 790                 795                 800
Thr Thr Arg His Leu Arg Pro Asp Met Lys Thr His Leu Asp Arg Pro
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Leu Val Val Glu Leu Gly Arg Asp Gly Ala Arg Gly Pro Val Gly Gly
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Lys Ala Arg Pro Glu Ala Ala Glu Ala Pro Glu Gly Val Asp Pro Pro
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Arg Arg His His Arg His Arg Asp Lys Asp Lys Thr Pro Ala Ala Gly
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Pro His Cys Asp Leu Glu Thr Ser Gly Thr Val Thr Val Gly Pro Met
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His Thr Leu Pro Ser Thr Cys Leu Gln Lys Val Glu Glu Gln Pro Glu
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Asp Ala Asp Asn Gln Arg Asn Val Thr Arg Met Gly Ser Gln Pro Pro
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Asp Pro Asn Thr Ile Val His Ile Pro Val Met Leu Thr Gly Pro Leu
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Gly Glu Ala Thr Val Val Pro Ser Gly Asn Val Asp Leu Glu Ser Gln
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Pro Arg Pro Ile Val Pro Tyr Ser Ser Met Phe Cys Leu Ser Pro Thr
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Glu Val Val Ile Leu Val Val Ile Ala Leu Ser Ser Ile Ala Leu Ala
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Ala Glu Asp Pro Val Arg Thr Asp Ser Pro Arg Asn Asn Ala Leu Lys
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Tyr Leu Asp Tyr Ile Phe Thr Gly Val Phe Thr Phe Glu Met Val Ile
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1265 1270 1275 1280
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Ser Asp Phe Ala Tyr Phe Tyr Phe Val Ser Phe Ile Phe Leu Cys Ser
1685 1690 1695

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Pro Pro Leu Glu Arg Gly His Ser Thr Glu Ile Pro Val Gly Arg Ser
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Gly Ala Leu Ala Val Asp Val Gln Met Gln Ser Ile Thr Arg Arg Gly
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Pro Asp Gly Glu Pro Gln Pro Gly Leu Glu Ser Gln Gly Arg Ala Ala
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Ser Met Pro Arg Leu Ala Ala Glu Thr Gln Pro Val Thr Asp Ala Ser
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Pro Met Lys Arg Ser Ile Ser Thr Leu Ala Gln Arg Pro Arg Gly Thr
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Ile Ala Gln Arg Ala Arg Thr Met Ala Leu Tyr Asn Pro Ile Pro Val
    50                  55                  60
Lys Gln Asn Cys Phe Thr Val Asn Arg Ser Leu Phe Val Phe Ser Glu
65                  70                  75                  80
Asp Asn Val Val Arg Lys Tyr Ala Lys Arg Ile Thr Glu Trp Pro Pro
                85                  90                  95
Phe Glu Tyr Met Ile Leu Ala Thr Ile Ile Ala Asn Cys Ile Val Leu
            100                 105                 110
Ala Leu Glu Gln His Leu Pro Asp Gly Asp Lys Thr Pro Met Ser Glu
        115                 120                 125
Arg Leu Asp Asp Thr Glu Pro Tyr Phe Ile Gly Ile Phe Cys Phe Glu
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Ala Gly Ile Lys Ile Ile Ala Leu Gly Phe Val Phe His Lys Gly Ser
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Tyr Leu Arg Asn Gly Trp Asn Val Met Asp Phe Val Val Val Leu Thr
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Gly Ile Leu Ala Thr Ala Gly Thr Asp Phe Asp Leu Arg Thr Leu Arg
            180                 185                 190
Ala Val Arg Val Leu Arg Pro Leu Lys Leu Val Ser Gly Ile Pro Ser
        195                 200                 205
Leu Gln Val Val Leu Lys Ser Ile Met Lys Ala Met Val Pro Leu Leu
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Gln Ile Gly Leu Leu Leu Phe Phe Ala Ile Leu Met Phe Ala Ile Ile
225                 230                 235                 240
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Gly Leu Glu Phe Tyr Met Gly Lys Phe His Lys Ala Cys Phe Pro Asn
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Ser Thr Asp Ala Glu Pro Val Gly Asp Phe Pro Cys Gly Lys Glu Ala
            260                 265                 270
Pro Ala Arg Leu Cys Glu Gly Asp Thr Glu Cys Arg Glu Tyr Trp Pro
        275                 280                 285
Gly Pro Asn Phe Gly Ile Thr Asn Phe Asp Asn Ile Leu Phe Ala Ile
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Leu Thr Val Phe Gln Cys Ile Thr Met Glu Gly Trp Thr Asp Ile Leu
305                 310                 315                 320
Tyr Asn Thr Asn Asp Ala Ala Gly Asn Thr Trp Asn Trp Leu Tyr Phe
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Ile Pro Leu Ile Ile Ile Gly Ser Phe Phe Met Leu Asn Leu Val Leu
            340                 345                 350
Gly Val Leu Ser Gly Glu Phe Ala Lys Glu Arg Glu Arg Val Glu Asn
        355                 360                 365
Arg Arg Ala Phe Leu Lys Leu Arg Arg Gln Gln Gln Ile Glu Arg Glu
    370                 375                 380
Leu Asn Gly Tyr Leu Glu Trp Ile Phe Lys Ala Glu Glu Val Met Leu
385                 390                 395                 400
Ala Glu Glu Asp Arg Asn Ala Glu Glu Lys Ser Pro Leu Asp Val Leu
                405                 410                 415
Lys Arg Ala Ala Thr Lys Lys Ser Arg Asn Asp Leu Ile His Ala Glu
            420                 425                 430
Glu Gly Glu Asp Arg Phe Ala Asp Leu Cys Ala Val Gly Ser Pro Phe
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Ala Arg Ala Ser Leu Lys Ser Gly Lys Thr Glu Ser Ser Ser Tyr Phe
    450                 455                 460
Arg Arg Lys Glu Lys Met Phe Arg Phe Phe Ile Arg Arg Met Val Lys
465                 470                 475                 480
Ala Gln Ser Phe Tyr Trp Val Val Leu Cys Val Val Ala Leu Asn Thr
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Leu Cys Val Ala Met Val His Tyr Asn Gln Pro Arg Arg Leu Thr Thr
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Thr Leu Tyr Phe Ala Glu Phe Val Phe Leu Gly Leu Phe Leu Thr Glu
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Met Ser Leu Lys Met Tyr Gly Leu Gly Pro Arg Ser Tyr Phe Arg Ser
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Ser Phe Asn Cys Phe Asp Phe Gly Val Ile Val Gly Ser Val Phe Glu
545                 550                 555                 560
Val Val Trp Ala Ala Ile Lys Pro Gly Ser Ser Phe Gly Ile Ser Val
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Leu Arg Ala Leu Arg Leu Leu Arg Ile Phe Lys Val Thr Lys Tyr Trp
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Ser Ser Leu Arg Asn Leu Val Val Ser Leu Leu Asn Ser Met Lys Ser
        595                 600                 605
Ile Ile Ser Leu Leu Phe Leu Leu Phe Leu Phe Ile Val Val Phe Ala
    610                 615                 620
Leu Leu Gly Met Gln Leu Phe Gly Gly Gln Phe Asn Phe Gln Asp Glu
625                 630                 635                 640
Thr Pro Thr Thr Asn Phe Asp Thr Phe Pro Ala Ala Ile Leu Thr Val
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Phe Gln Ile Leu Thr Gly Glu Asp Trp Asn Ala Val Met Tyr His Gly
            660                 665                 670
Ile Glu Ser Gln Gly Gly Val Ser Lys Gly Met Phe Ser Ser Phe Tyr
        675                 680                 685
Phe Ile Val Leu Thr Leu Phe Gly Asn Tyr Thr Leu Leu Asn Val Phe
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Leu Ala Ile Ala Val Asp Asn Leu Ala Asn Ala Gln Glu Leu Thr Lys
705                 710                 715                 720
Asp Glu Glu Glu Met Glu Glu Ala Ala Asn Gln Lys Leu Ala Leu Gln
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Ser Ile Ala Ala Arg Gln Gln Asn Ser Ala Lys Ala Arg Ser Val Trp
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Glu Gln Arg Ala Ser Gln Leu Arg Leu Gln Asn Leu Arg Ala Ser Cys
    770                 775                 780
Glu Ala Leu Tyr Ser Glu Met Asp Pro Glu Glu Arg Leu Arg Phe Ala
785                 790                 795                 800
Thr Thr Arg His Leu Arg Pro Asp Met Lys Thr His Leu Asp Arg Pro
                805                 810                 815
Leu Val Val Glu Leu Gly Arg Asp Gly Ala Arg Gly Pro Val Gly Gly
            820                 825                 830
Lys Ala Arg Pro Glu Ala Ala Glu Ala Pro Glu Gly Val Asp Pro Pro
        835                 840                 845
Arg Arg His His Arg His Arg Asp Lys Asp Lys Thr Pro Ala Ala Gly
    850                 855                 860
Asp Gln Asp Arg Ala Glu Ala Pro Lys Ala Glu Ser Gly Glu Pro Gly
865                 870                 875                 880
Ala Arg Glu Glu Arg Pro Arg Pro His Arg Ser His Ser Lys Glu Ala
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Ala Gly Pro Pro Glu Ala Arg Ser Glu Arg Gly Arg Gly Pro Gly Pro
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Glu Gly Gly Arg Arg His His Arg Arg Gly Ser Pro Glu Glu Ala Ala
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Glu Arg Glu Pro Arg Arg His Arg Ala His Arg His Gln Asp Pro Ser
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Lys Glu Cys Ala Gly Ala Lys Gly Glu Arg Arg Ala Arg His Arg Gly
945                 950                 955                 960
Gly Pro Arg Ala Gly Pro Arg Glu Ala Glu Ser Gly Glu Glu Pro Ala
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Arg Arg His Arg Ala Arg His Lys Ala Gln Pro Ala His Glu Ala Val
            980                 985                 990
Glu Lys Glu Thr Thr Glu Lys Glu Ala Thr Glu Lys Glu Ala Glu Ile
        995                 1000                1005
Val Glu Ala Asp Lys Glu Lys Glu Leu Arg Asn His Gln Pro Arg Glu
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Pro His Cys Asp Leu Glu Thr Ser Gly Thr Val Thr Val Gly Pro Met
1025                1030                1035                1040
His Thr Leu Pro Ser Thr Cys Leu Gln Lys Val Glu Glu Gln Pro Glu
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Asp Ala Asp Asn Gln Arg Asn Val Thr Arg Met Gly Ser Gln Pro Pro
            1060                1065                1070
Asp Pro Asn Thr Ile Val His Ile Pro Val Met Leu Thr Gly Pro Leu
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Gly Glu Ala Thr Val Val Pro Ser Gly Asn Val Asp Leu Glu Ser Gln
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Ala Glu Gly Lys Lys Glu Val Glu Ala Asp Asp Val Met Arg Ser Gly
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Pro Arg Pro Ile Val Pro Tyr Ser Ser Met Phe Cys Leu Ser Pro Thr
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Asn Leu Leu Arg Arg Phe Cys His Tyr Ile Val Thr Met Arg Tyr Phe
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Glu Val Val Ile Leu Val Val Ile Ala Leu Ser Ser Ile Ala Leu Ala
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Tyr Leu Asp Tyr Ile Phe Thr Gly Val Phe Thr Phe Glu Met Val Ile
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Lys Met Ile Asp Leu Gly Leu Leu Leu His Pro Gly Ala Tyr Phe Arg
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Asp Leu Trp Asn Ile Leu Asp Phe Ile Val Val Ser Gly Ala Leu Val
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Leu Pro Lys Leu Lys Ala Val Phe Asp Cys Val Val Asn Ser Leu Lys
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Asn Val Leu Asn Ile Leu Ile Val Tyr Met Leu Phe Met Phe Ile Phe
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Ala Val Ile Ala Val Gln Leu Phe Lys Gly Lys Phe Phe Tyr Cys Thr
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Asp Glu Ser Lys Glu Leu Glu Arg Asp Cys Arg Gly Gln Tyr Leu Asp
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Tyr Glu Lys Glu Glu Val Glu Ala Gln Pro Arg Gln Trp Lys Lys Tyr
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Asp Phe His Tyr Asp Asn Val Leu Trp Ala Leu Leu Thr Leu Phe Thr
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Val Ser Thr Gly Glu Gly Trp Pro Met Val Leu Lys His Ser Val Asp
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Ala Thr Tyr Glu Glu Gln Gly Pro Ser Pro Gly Tyr Arg Met Glu Leu
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Ser Ile Phe Tyr Val Val Tyr Phe Val Val Phe Pro Phe Phe Phe Val
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Lys Val Met Ser Glu Cys Ser Leu Glu Lys Asn Glu Arg Ala Cys Ile
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Asp Phe Ala Ile Ser Ala Lys Pro Leu Thr Arg Tyr Met Pro Gln Asn
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Arg Gln Ser Phe Gln Tyr Lys Thr Trp Thr Phe Val Val Ser Pro Pro
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Phe Glu Tyr Phe Ile Met Ala Met Ile Ala Leu Asn Thr Val Val Leu
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Met Met Lys Phe Tyr Asp Ala Pro Tyr Glu Tyr Glu Leu Met Leu Lys
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Cys Leu Asn Ile Val Phe Thr Ser Met Phe Ser Met Glu Cys Val Leu
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Val Phe Asp Phe Val Thr Val Leu Gly Ser Ile Thr Asp Ile Leu Val
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Leu Phe Arg Ala Ala Arg Leu Ile Lys Leu Leu Arg Gln Gly Tyr Thr
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Ile Arg Ile Leu Leu Trp Thr Phe Val Gln Ser Phe Lys Ala Leu Pro
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Tyr Val Cys Leu Leu Ile Ala Met Leu Phe Phe Ile Tyr Ala Ile Ile
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Ser Asp Phe Ala Tyr Phe Tyr Phe Val Ser Phe Ile Phe Leu Cys Ser
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Phe Leu Met Leu Asn Leu Phe Val Ala Val Ile Met Asp Asn Phe Glu
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Tyr Leu Thr Arg Asp Ser Ser Ile Leu Gly Pro His His Leu Asp Glu
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Gly Leu Gly Lys Lys Cys Pro Ala Arg Val Ala Tyr Lys Arg Leu Val
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Arg Met Asn Met Pro Ile Ser Asn Glu Asp Met Thr Val His Phe Thr
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Ser Thr Leu Met Ala Leu Ile Arg Thr Ala Leu Glu Ile Lys Leu Ala
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Pro Ala Gly Thr Lys Gln His Gln Cys Asp Ala Glu Leu Arg Lys Glu
    1810                1815                1820
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Gln Met Gln Gln Ala Pro Gly Gly Leu Ser Gln Met Gly Pro Val Ser
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Leu Phe His Pro Leu Lys Ala Thr Leu Glu Gln Thr Gln Pro Ala Val
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Leu Arg Gly Ala Arg Val Phe Leu Arg Gln Lys Ser Ser Thr Ser Leu
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Ser Asn Gly Gly Ala Ile Gln Asn Gln Glu Ser Gly Ile Lys Glu Ser
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Val Ser Trp Gly Thr Gln Arg Thr Gln Asp Ala Pro His Glu Ala Arg
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Pro Pro Leu Glu Arg Gly His Ser Thr Glu Ile Pro Val Gly Arg Ser
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Gly Ala Leu Ala Val Asp Val Gln Met Gln Ser Ile Thr Arg Arg Gly
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Pro Asp Gly Glu Pro Gln Pro Gly Leu Glu Ser Gln Gly Arg Ala Ala
1985                1990                1995                2000
Ser Met Pro Arg Leu Ala Ala Glu Thr Gln Pro Val Thr Asp Ala Ser
                2005                2010                2015
Pro Met Lys Arg Ser Ile Ser Thr Leu Ala Gln Arg Pro Arg Gly Thr
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His Leu Cys Ser Thr Thr Pro Asp Arg Pro Pro Pro Ser Gln Ala Ser
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Ser His His His His His Arg Cys His Arg Arg Arg Asp Arg Lys Gln
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Pro Ser Ser Ala Val Gly Pro Gly Leu Pro Pro Gly Glu Gly Pro Thr
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Ser Cys Asp Arg Phe Gly Gly Arg Glu Pro Pro Lys Pro Lys Pro Ser
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Leu Ser Ser His Pro Thr Ser Pro Thr Ala Gly Gln Glu Pro Gly Pro
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His Pro Gln Ala Gly Ser Ala Val Gly Phe Pro Asn Thr Thr Pro Cys
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Cys Arg Glu Thr Pro Ser Ala Ser Pro Trp Pro Leu Ala Leu Glu Leu
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Ala Leu Thr Leu Thr Trp Gly Ser Val Trp Thr Val Arg Pro Leu Ser
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Lys Gln Ser Met Ala Gln Arg Ala Arg Thr Met Ala Leu Tyr Asn Pro
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Ile Pro Val Arg Gln Asn Cys Leu Thr Val Asn Arg Ser Leu Phe Leu
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Phe Ser Glu Asp Asn Val Val Arg Lys Tyr Ala Lys Lys Ile Thr Glu
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Trp Pro Pro Phe Glu Tyr Met Ile Leu Ala Thr Ile Ile Ala Asn Cys
            100                 105                 110
Ile Val Leu Ala Leu Glu Gln His Leu Pro Asp Asp Asp Lys Thr Pro
        115                 120                 125
Met Ser Glu Arg Leu Asp Asp Thr Glu Pro Tyr Phe Ile Gly Ile Phe
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Cys Phe Glu Ala Gly Ile Lys Ile Ile Ala Leu Gly Phe Ala Phe His
145                 150                 155                 160
Lys Gly Ser Tyr Leu Arg Asn Gly Trp Asn Val Met Asp Phe Val Val
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Val Leu Thr Gly Ile Leu Ala Thr Val Gly Thr Glu Phe Asp Leu Arg
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Thr Leu Arg Ala Val Arg Val Leu Arg Pro Leu Lys Leu Val Ser Gly
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Pro Leu Leu Gln Ile Gly Leu Leu Leu Phe Phe Ala Ile Leu Ile Phe
225                 230                 235                 240
Ala Ile Ile Gly Leu Glu Phe Tyr Met Gly Lys Phe His Thr Thr Cys
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Phe Glu Glu Gly Thr Asp Asp Ile Gln Gly Glu Ser Pro Ala Pro Cys
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Gly Thr Glu Glu Pro Ala Arg Thr Cys Pro Asn Gly Thr Lys Cys Gln
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Pro Tyr Trp Glu Gly Pro Asn Asn Gly Ile Thr Gln Phe Asp Asn Ile
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Trp Leu Tyr Phe Ile Pro Leu Ile Ile Ile Gly Ser Phe Phe Met Leu
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Glu Trp Leu Ser Asp Phe Leu Tyr Tyr Ala Glu Phe Ile Phe Leu Gly
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Pro Tyr Phe His Ser Ser Phe Asn Cys Phe Asp Cys Gly Val Ile Ile
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Phe Gly Ile Ser Val Leu Arg Ala Leu Arg Leu Leu Arg Ile Phe Lys
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Val Thr Lys Tyr Trp Ala Ser Leu Arg Asn Leu Val Val Ser Leu Leu
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Asn Ser Met Lys Ser Ile Ile Ser Leu Leu Phe Leu Leu Phe Leu Phe
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Ile Val Val Phe Ala Leu Leu Gly Met Gln Leu Phe Gly Gly Gln Phe
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Val Met Tyr Asp Gly Ile Lys Ser Gln Gly Gly Val Gln Gly Gly Met
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Val Phe Ser Ile Tyr Phe Ile Val Leu Thr Leu Phe Gly Asn Tyr Thr
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Lys Asn Gln Lys Pro Ala Lys Ser Val Trp Glu Gln Arg Thr Ser Glu
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Pro Asp Met Lys Thr His Leu Asp Arg Pro Leu Val Val Asp Pro Gln
820 825 830
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Val Asp Gln Arg Leu Gly Gln Gln Arg Ala Glu Asp Phe Leu Arg Lys
850 855 860
Gln Ala Arg Tyr His Asp Arg Ala Arg Asp Pro Ser Gly Ser Ala Gly
865 870 875 880
Leu Asp Ala Arg Arg Pro Trp Ala Gly Ser Gln Glu Ala Glu Leu Ser
885 890 895
Arg Glu Gly Pro Tyr Gly Arg Glu Ser Asp His His Ala Arg Glu Gly
900 905 910
Ser Leu Glu Gln Pro Gly Phe Trp Glu Gly Glu Ala Glu Arg Gly Lys
915 920 925
Ala Gly Asp Pro His Arg Arg His Val His Arg Gln Gly Gly Ser Arg
930 935 940
Glu Ser Arg Ser Gly Ser Pro Arg Thr Gly Ala Asp Gly Glu His Arg
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Arg His Arg Ala His Arg Arg Pro Gly Glu Glu Gly Pro Glu Asp Lys
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995 1000 1005
His Arg His Gly Ala Pro Ala Thr Tyr Glu Gly Asp Ala Arg Arg Glu
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His Thr Thr Val Asp Ile Pro Pro Ala Cys Pro Pro Pro Leu Asn His
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Thr Val Val Gln Val Asn Lys Asn Ala Asn Pro Asp Pro Leu Pro Lys
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Lys Tyr Glu Phe His Tyr Asp Asn Val Leu Trp Ala Leu Leu Thr Leu
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Lys Cys Pro Ala Arg Val Ala Tyr Lys Arg Leu Leu Arg Met Asp Leu
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Leu Ile Arg Thr Ala Leu Asp Ile Lys Ile Ala Lys Gly Gly Ala Asp
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1985                1990                1995                2000
Pro Thr Gln Glu Gly Gly Pro Gly Gln Asn Ala Leu Pro Ser Thr Gln
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        35                  40                  45
Lys Gln Ser Met Ala Gln Arg Ala Arg Thr Met Ala Leu Tyr Asn Pro
    50                  55                  60
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For two-letter codes and other abbreviations, refer to the "Guidance Notes on Codes and Abbreviations" appearing at the beginning of each regular issue of the PCT Gazette.

(54) Title: ELECTRICAL FIELD STIMULATION OF EUKARYOTIC CELLS

(57) Abstract: Methods of identifying activators and inhibitors of voltage-gated ion channels are provided in which the methods employ electrical field stimulation of the cells in order to manipulate the open/close state transition of the voltage-gated ion channels. This allows for more convenient, more precise experimental manipulation of these transitions, and, coupled with efficient methods of detecting the result of ion flux through the channels, provides methods that are especially suitable for high throughput screening.

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INTERNATIONAL SEARCH REPORT

International application No.

PCT/US02/22161

A. CLASSIFICATION OF SUBJECT MATTER

IPC(7) : C07H 21/04; C12N 5/00, 15/00; C12P 1/06; G01N 33/53, 33/554, 35/00; G01R 27/00

US CL : 324/600; 422/50, 55, 67; 435/69.1, 320.1, 325; 436/43, 519, 800, 807, 809; 536/23.5

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

U.S. : 324/600; 422/50, 55, 67; 435/69.1, 320.1, 325; 436/43, 519, 800, 807, 809; 536/23.5

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practicable, search terms used)

USPT, PGPB, JPAB, EPAB, DWPI, REGISTRY, HCAPLUS

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
Y	US 6,057,114 A (AKONG et al.) 02 May 2000 (02.05.2000), Abstract, Column 1, Lines 12-15; Column 4, Lines 4-22; Column 9, Lines 63-68; Column 17, Lines 22-40; Column 17, Line 63 to Column 18, Line 15; Column 20, Lines 32-54; Column 22, Line 57 to Column 23, Line 30; Column 23, Line 54 to Column 24, Line 9; Column 26, Lines 9-20; Column 27, Lines 14-22; Column 33, Line 61 to Column 34 Line 27; Column 41, Line 43 to Column 42, Line 9; Column 42, Lines 28-67; Column 43, Lines 35-56.	1-16 and 20-60
Y	CONNOLLY, P. et al. An Extracellular Microelecrode Array for Monitoring Electrogenic Cells in Culture. Biosensors and Bioelectronics, 1990, Vol. 5, Pages 223-	17-19 and 61-74
Y, P	US 6,377,057 B1 (BORKHOLDER) 23 April 2002 (23.04.2002), entire document.	17-19 and 61-74

☐ Further documents are listed in the continuation of Box C. ☐ See patent family annex.

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"Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art

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